

[Click here to view linked References](#)

1
2
3 **Membranous nephropathy after multiple Hymenoptera stings: a case**
4
5
6 **report**
7
8
9

10
11
12 Kenichi Morii^{1,2}, Toshiki Doi^{1,2}, Yoshio Yuba¹, Aiko Okubo¹, Kazuomi Yamashita¹,
13
14
15
16 Sonoo Mizuiri¹, Yoshiko Nishizawa¹, Kenichiro Shigemoto¹, Akira Shimizu³, Takao
17
18
19 Masaki⁴
20
21
22
23
24

25 1 Department of Nephrology, Ichiyokai Harada Hospital, 7-10 Kairoyamacho, Saeki-ku,
26
27
28 Hiroshima, Japan
29
30

31
32 2 Department of Kidney Disease and Community Medicine, Hiroshima University
33
34
35 Hospital, 1-2-3 Kasumi, Minami-ku, Hiroshima, Japan
36
37

38 3 Department of Analytic Human Pathology, Nippon Medical School, 1-25-16 Nezu,
39
40
41 Bunkyo-ku, Tokyo, Japan
42
43

44 4 Department of Nephrology, Hiroshima University Hospital, 1-2-3 Kasumi, Minami-ku,
45
46
47 Hiroshima, Japan
48
49

50
51
52
53
54 Corresponding Author
55
56

57 Toshiki Doi
58
59
60
61
62
63
64
65

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

E-mail: doitoshi@hiroshima-u.ac.jp

Telephone/fax number: +81 82-923-5161/+81 82-921-8035

Number of words: 2877 words

1
2
3 **Abstract**
4
5

6 An association between Hymenoptera (bee and wasp) stings and nephrotic
7
8
9 syndrome has been rarely reported. We report a case of nephrotic syndrome after multiple
10
11
12 Hymenoptera stings, and membranous nephropathy was later diagnosed by a kidney
13
14
15 biopsy. The patient was a 79-year-old woman who was stung by Hymenoptera at seven
16
17
18 sites on her body. A weight gain of 3.7 kg was observed in the patient at 1 week after
19
20
21 being stung, and she had considerable edema in both lower extremities. A urine protein
22
23
24 concentration of 14.8 g/g creatinine and a serum albumin concentration of 1.7 g/dL led to
25
26
27 the diagnosis of nephrotic syndrome. A percutaneous kidney biopsy 8 days after the
28
29
30 Hymenoptera stings showed stage I membranous nephropathy. She was in complete
31
32
33 remission 1 week after the administration of oral prednisolone 40 mg/day, which was
34
35
36 started 14 days after Hymenoptera stings, and had no relapse of nephrotic syndrome. To
37
38
39
40
41 the best of our knowledge, this is the first report of biopsy-proven membranous
42
43
44 nephropathy caused by Hymenoptera stings.
45
46
47
48
49
50

51 **Keywords:** Hymenoptera sting, membranous nephropathy, nephrotic syndrome
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 **Introduction**
4
5

6 Hymenoptera (bee or wasp) stings lead to local reactions characterized by
7
8
9 redness and swelling at the stinging site and systemic allergic reactions or anaphylaxis
10
11
12 [1]. There are various other unusual reactions to Hymenoptera stings, such as hemolysis,
13
14
15 rhabdomyolysis, acute renal failure, and nephrotic syndrome [2,3]. Minimal change
16
17
18 nephrotic syndrome [4–8], proliferative glomerulonephropathy [9], mesangial
19
20
21 proliferative glomerulonephritis [10], and mesangial proliferative glomerulonephropathy
22
23
24 with possible early membranous nephropathy [11] have been reported as nephrotic
25
26
27 syndrome caused by Hymenoptera stings. However, the reports providing histological
28
29
30 results of nephrotic syndrome caused by Hymenoptera stings are limited. In this report,
31
32
33 we present a case of stage I membranous nephropathy induced by Hymenoptera stings in
34
35
36 which complete remission was achieved by oral prednisolone.
37
38
39
40
41
42
43
44

45 **Case report**
46

47 A 79-year-old Japanese woman was stung by Hymenoptera at seven sites on her
48
49
50 body, with one on her right arm and six on her left arm, while working in the garden. On
51
52
53 the same day, a dermatologist prescribed an anti-histamine, but the patient experienced
54
55
56 mild edema in both lower extremities in the evening. Her symptoms did not improve after
57
58
59
60
61
62
63
64
65

1
2
3 taking the anti-histamine. Subsequently, a physician noted hypoalbuminemia and
4
5
6 proteinuria (3+) on day 7 after she was stung by Hymenoptera. On the same day, she was
7
8
9 referred to our hospital with the suspicion of nephrotic syndrome.
10

11
12 The patient's past medical history was notable for dyslipidemia, insomnia,
13
14 gastroesophageal reflux disease, and allergic bronchitis. She was taking pravastatin 5
15
16 mg/day, zolpidem 10 mg/day, esomeprazole 10 mg/day, tranexamic acid 750 mg/day,
17
18
19 carbocysteine 1500 mg/day, and rebamipide 300 mg/day for these conditions. Eight years
20
21
22 before the current hospital admission, she was diagnosed with non-tuberculous
23
24
25 mycobacteriosis and was followed up with no medication.
26
27
28
29
30

31
32 On admission, a physical examination showed no localized swelling at the sting
33
34 sites, but strong edema was observed in both lower extremities, and her blood pressure
35
36 was 137/74 mmHg. She weighed 52.7 kg, which indicated a weight gain of 3.7 kg for 7
37
38 days. Her laboratory data on admission are shown in Table 1. A urinalysis showed 4+
39
40
41 proteinuria (14.18 g/g creatinine [Cr]). Quantitative protein excretion on admission was
42
43
44 7.16 g/day. Urinary sediment showed 1–4 red blood cells/high power field, and 1–4
45
46
47 granular casts/whole field. A laboratory investigation showed that the blood urea nitrogen
48
49
50 concentration was 10 mg/dL, serum creatinine concentration was 0.54 mg/dL, total
51
52
53 protein concentration was 4.9 g/dL, albumin concentration was 1.7 g/dL, total cholesterol
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 concentration was 401 mg/dL, low-density lipoprotein cholesterol concentration was 266
4
5
6 mg/dL, and selectivity index of urinary protein was 0.11. The clinical data of proteinuria
7
8
9 ≥ 3.5 g/day and serum albumin concentration ≤ 3.0 g/dL led to a diagnosis of nephrotic
10
11
12 syndrome.
13

14
15
16 An ultrasound-guided percutaneous kidney biopsy was performed on the
17
18 second day of hospitalization. Light microscopy showed minor glomerular
19
20
21 abnormalities with moderate atherosclerotic lesions in the arterioles and small arteries,
22
23
24 and slight tubulointerstitial changes. Neither spikes nor a bubbly appearance of the
25
26
27 capillary wall of glomeruli was observed (Fig. 1). An immunofluorescence examination
28
29
30 using fresh frozen sections showed moderate, irregular and segmental staining for
31
32
33 immunoglobulin (Ig) G (Fig. 2a), IgM (Fig. 2b), C3c (Fig. 2c), and fibrinogen (Fig. 2d)
34
35
36 on the glomerular capillary loops. Staining of M-type phospholipase A₂ receptor
37
38
39 (PLA₂R) on paraffin sections was negative in the glomerular capillary loops, although
40
41
42 autofluorescence from red blood cells was positive for segmental staining (Fig. 2e, 2f).
43
44
45
46
47 On electron microscopy, there were diffuse and partially segmental podocyte foot
48
49
50 process effacement with microvillous formation and segmental irregular subepithelial
51
52
53 electron-dense deposits in glomeruli without apparent formation of spikes, consistent
54
55
56 with stage I membranous nephropathy (Fig. 3). Laser microdissection and mass
57
58
59
60
61
62
63
64
65

1
2
3 spectrometry (LC-MS/MS) [12] in this case did not show the presence of PLA₂R,
4
5
6 thrombospondin type-1 domain-containing 7A (THSD7A) [13], nerve epidermal growth
7
8
9 factor-like 1 (NELL-1) [14], exostosin 1/exostosin 2 (EXT1/EXT2) [15], semaphorin
10
11
12 3B (SEMA3B) [16], protocadherin 7 (PCDH7) [17], and neural cell adhesion molecule
13
14
15 1 (NCAM1) [18] (data not shown). Serum PLA₂R antibody was negative at 24 days
16
17
18 after steroid treatment. Secondary causes of membranous nephropathy were
19
20
21 investigated. Serology was negative for hepatitis B and C, and a serological test for
22
23
24 syphilis and the *Treponema pallidum* hemagglutination test were both negative.
25
26
27
28 Angiotensin-converting enzyme, anti-nuclear antibodies, antibody to Sjögren's
29
30
31 syndrome A and B, and anti-U1 RNP antibody were normal. Carcinoembryonic antigen,
32
33
34 carbohydrate antigen 19-9, carbohydrate antigen 125, α -fetoprotein, and soluble
35
36
37 interleukin-2 receptor concentrations were normal, and did not indicate malignancy. A
38
39
40 stool occult blood test was negative. Esophagogastroduodenoscopy also showed no
41
42
43 evidence of malignancy. Computed tomography scans of the chest showed lobular
44
45
46 central granular shadows, and enlarged bronchioles and nodules in both lung fields,
47
48
49 consistent with non-tuberculous mycobacteriosis. An abdominal computed tomography
50
51
52 scan showed a small amount of ascites, with no renal enlargement or atrophy. We
53
54
55
56
57 diagnosed membranous nephropathy secondary to Hymenoptera stings on the basis of
58
59

1
2
3 the renal biopsy findings, clinical and laboratory data, and exclusion of other causes of
4
5
6 membranous nephropathy.
7

8
9 On the seventh day of hospitalization (14 days after Hymenoptera stings),
10
11 prednisolone was started at an oral dose of 40 mg/day (0.8 mg/kg body weight), and the
12
13 dose was decreased every 2 weeks. The clinical course of the patient is shown in Fig. 4.
14
15

16
17 On the 13th day of hospitalization, urinalysis revealed no proteinuria (0.00 g/gCr). She
18
19 had complete remission (6 days after steroid treatment) and no relapse of nephrotic
20
21 syndrome. Laboratory data 6 months after steroid therapy were as follows: urinary protein,
22
23 0.00 g/gCr; no abnormalities in urinary sediment; serum total protein concentration, 6.3
24
25 g/dL; serum albumin concentration, 4.0 g/dL; serum creatinine concentration, 0.48
26
27 mg/dL; estimated glomerular filtration rate, 91 mL/min/1.73 m²; total cholesterol
28
29 concentration, 155 mg/dL, and low-density lipoprotein-cholesterol concentration, 68
30
31 mg/dL.
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47

48 **Discussion**

49
50 Since the first report on nephrotic syndrome due to Hymenoptera (wasp or bee)
51
52 stings by Rytand in 1955 [3], approximately 20 related case reports have been published.
53
54

55
56 A renal biopsy was performed in nine patients with nephrotic syndrome due to
57
58

1
2
3 Hymenoptera. Table 2 shows clinical features, pathology of kidney, treatment and
4
5
6 outcome of these 9 patients. Six patients were diagnosed with minimal change nephrotic
7
8
9 syndrome (MCNS) [4–8]. One patient had proliferative glomerulonephropathy [9], one
10
11
12 had mesangial proliferative glomerulonephritis [10], and one had mesangial proliferative
13
14
15 glomerulonephropathy with possible early membranous nephropathy [11]. Although the
16
17
18 majority case (4/6) with MCNS after Hymenoptera stings resulted in complete remission
19
20
21 with steroid treatment [5–8], cases with proliferative glomerulonephritis did not resolve
22
23
24 promptly [9–11]. The outcome of 2 MCNS cases was unknown [4]. A case with
25
26
27 proliferative glomerulonephritis resulted in chronic dialysis [9], a case with membranous
28
29
30 proliferative glomerulonephritis relapsed twice [10], and another case with mesangial
31
32
33 proliferative glomerulonephritis resulted in incomplete remission [11]. To the best of our
34
35
36 knowledge, this is the first case report of definite membranous nephropathy with
37
38
39 nephrotic syndrome after multiple Hymenoptera stings resulted in complete remission
40
41
42 with steroid therapy.
43
44
45

46
47
48 IgG4-dominance had the highest specificity in the differentiation of membranous
49
50
51 nephropathy, just as high as that for anti-PLA₂R seropositivity [19]. Staining for IgG
52
53
54 subclasses from the renal biopsy was not performed in this case because of missing fresh
55
56
57 frozen tissue. M-type PLA₂R on glomerular podocytes has been identified as a major
58
59
60

1
2
3 antigen in idiopathic membranous nephropathy [20]. PLA₂ is associated with diverse
4
5
6 physiological processes, such as toxicity (neurotoxicity and myotoxicity), pathology
7
8
9 (inflammation and cancer), and physiology (proliferation, contraction, and secretion) [21].
10
11
12 Bee venom contains PLA₂, which is a major allergen, and the peptide melittin and
13
14
15 hyaluronidase, whereas wasp venom contains antigen 5, different phospholipases, and
16
17
18 hyaluronidase [1]. PLA₂ accounts for up to 12% of bee venom contents [22]. It is reported
19
20
21 that there was a strong direct correlation between anti-PLA₂R and anti-secreted PLA₂
22
23
24 antibodies [23]. PLA₂ injected by Hymenoptera stings may have contributed to the
25
26
27 development of membranous nephropathy in the current case. However, serum PLA₂R
28
29
30 antibody after steroid treatment in this case was negative. In addition,
31
32
33 immunofluorescence using paraffin section for PLA₂R antigen and LC-MS/MS findings
34
35
36 of renal biopsy specimen could not be detected in this case (Fig. 2e). We cannot conclude
37
38
39 that there was a relationship between PLA₂, which is contained in Hymenoptera venom,
40
41
42 and the antigen of this patient's membranous nephropathy. However, we could not rule
43
44
45 out the possibility that treatment with steroid stabilized the proteinuria, and circulating
46
47
48 anti-PLA₂R concentrations became undetectable, as previously reported [24]. Segmental
49
50
51 and irregular deposition of IgG and electron-dense deposits in stage I membranous
52
53
54 nephropathy were observed, but PLA₂R was not detected by LC-MS/MS and
55
56
57
58
59
60
61
62
63
64
65

1
2
3 immunofluorescence using paraffin-embedded sections. However, one possibility is still
4
5
6 remained that the reduced sensitivity of antigen detection in paraffin-embedded sections
7
8
9 and the low levels of antigen in segmental deposition of immune complexes, which may
10
11
12 fall below the sensitivity of detection of LC-MS/MS, may have contributed to the failure
13
14
15 to detect PLA₂R in this study. The reason for the different renal pathological findings in
16
17
18 nephrotic syndrome after Hymenoptera (wasp or bee) stings is unclear, but may be related
19
20
21 to toxic envenomation between bees and wasps and between a few stings and thousands
22
23
24 of stings [8]. Further case reports are necessary to determine the cause and reason for
25
26
27 different renal pathological findings in nephrotic syndrome after Hymenoptera stings.
28
29
30

31
32 The onset of this case was rapid (edema was observed on the same day, and
33
34
35 hypoalbuminemia and urinary protein were observed on day 7 after the Hymenoptera
36
37
38 stings), and the reaction against steroid therapy was also rapid. These clinical courses
39
40
41 were different from general membranous nephropathy. A rapid onset of edema and
42
43
44 nephrotic syndrome, but not a rapid response against steroid therapy, may be
45
46
47 characteristics of nephropathy after Hymenoptera stings. In previous studies, the timing
48
49
50 of edema was 2 days to 2 weeks [5,7–11], and the timing of nephrotic syndrome was
51
52
53 several days to 4 weeks [5,7–11] after Hymenoptera stings. The rapid reaction against
54
55
56 steroid therapy has not been reported in previous cases of nephropathy after Hymenoptera
57
58
59

1
2
3 stings [5–8]. The prognosis of nephrotic syndrome with segmental membranous
4
5
6 glomerulopathy is favorable; 50% of patients achieve complete remission and 21%
7
8
9 achieve partial remission [25]. We consider that the rapid response to steroid therapy in
10
11
12 our patient may be related to segmental deposition of IgG on the capillary walls of
13
14
15
16 glomeruli.
17

18
19 The requirement of performing hemodialysis, despite corticosteroid treatment,
20
21
22 has been reported in some cases of nephrotic syndrome after Hymenoptera stings [8,9].
23
24
25 This case highlights the importance of an early renal biopsy and early steroid therapy in
26
27
28 managing nephrotic syndrome after Hymenoptera stings.
29
30
31
32
33
34
35
36
37

38 **Compliance with Ethical Standards**

39
40
41 Funding: None.
42

43
44 Conflict of interest: The authors have declared that no conflict of interest exists.
45
46

47
48 Consent for publication: Informed consent was obtained from the patient.
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

References

1. Hoffman DR. Hymenoptera venom allergens. *Clin Rev Allergy Immunol* 2006;30:109–28. <https://doi.org/10.1385/CRIAI:30:2:109>.
2. Vetter RS, Visscher PK, Camazine S. Mass envenomations by honey bees and wasps. *West J Med* 1999;170:223–7 (PMID: 10344177).
3. Rytand DA. Onset of the nephrotic syndrome during a reaction to bee sting. *Stanford Med Bull* 1955;13:224–33 (PMID: 14386174).
4. Venters HD, Vernier RL, Worthen HG GR. Bee sting nephrosis: A study of the immunopathologic mechanisms. *Am J Dis Child* 1961;102:688–9.
5. Olivero JJ, Ayus JC, Eknayan G. Nephrotic syndrome developing after bee stings. *South Med J* 1981;74:82–3. <https://doi.org/10.1097/00007611-198101000-00030>.
6. Révai T, Harnos G. Simvastatin treatment in nephrotic syndrome associated with a bee sting. *J R Soc Med* 1999;92:23–4. <https://doi.org/10.1177/014107689909200110>.
7. Zaman F, Saccaro S, Latif S, Atray N, Abreo K. Minimal change glomerulonephritis following a wasp sting. *Am J Nephrol* 2001;21:486–9. <https://doi.org/10.1159/000046653>.
8. Humeda YS, Clapp WL, Humeda H. Minimal change disease after multiple wasp stings. *Clin Nephrol – Case Stud* 2022;10:16–20. <https://doi.org/10.5414/cncs110369>.
9. Taber TE, King LH, Aust CH. Bee sting nephropathy in a transplant patient. *Indiana Med* 1986;79:778–9 (PMID: 3534070).
10. Sensirivatana R, Sukvichai P, Futrakul P. Nephrotic syndrome following a bee sting. *J Med Assoc Thai* 1984;67:525–8 (PMID: 6520578).
11. Tauk B, Hachem H, Bastani B. Nephrotic syndrome with mesangial proliferative glomerulonephritis induced by multiple wasp stings. *Am J Nephrol* 1999;19:70–2. <https://doi.org/10.1159/000013429>.
12. Bobart SA, Tehranian S, Sethi S, Alexander MP, Nasr SH, Moura Marta C, et al. A target antigen–based approach to the classification of membranous nephropathy. *Mayo Clin Proc* 2021;96:577–91. <https://doi.org/10.1016/j.mayocp.2020.11.028>.
13. Ren S, Wu C, Zhang Y, Wang AY, Li G, Wang L, et al. An update on clinical significance of use of THSD7A in diagnosing idiopathic membranous nephropathy : a systematic review and meta- analysis of THSD7A in IMN. *Ren*

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
- Fail 2018;0:306–13. <https://doi.org/10.1080/0886022X.2018.1456457>.
14. Sethi S, Debiec H, Madden B, Charlesworth MC, Morelle J, Gross LA, et al. Neural epidermal growth factor-like 1 protein (NELL-1) associated membranous nephropathy. *Kidney Int* 2020;97:163–74. <https://doi.org/10.1016/j.kint.2019.09.014>.
15. Sethi S, Madden BJ, Debiec H, Cristine Charlesworth M, Gross L, Ravindran A, et al. Exostosin 1/exostosin 2-associated membranous nephropathy. *J Am Soc Nephrol* 2019;30:1123–36. <https://doi.org/10.1681/ASN.2018080852>.
16. Sethi S, Debiec H, Madden B, Vivarelli M, Charlesworth MC, Ravindran A, et al. Semaphorin 3B-associated membranous nephropathy is a distinct type of disease predominantly present in pediatric patients. *Kidney Int* 2020;98:1253–64. <https://doi.org/10.1016/j.kint.2020.05.030>.
17. Sethi S, Madden B, Debiec H, Morelle J, Charlesworth MC, Gross L, et al. Protocadherin 7-associated membranous nephropathy. *J Am Soc Nephrol* 2021;32:1249–61. <https://doi.org/10.1681/ASN.2020081165>.
18. Caza TN, Hassen SI, Kuperman M, Sharma SG, Dvanajscak Z, Arthur J, et al. Neural cell adhesion molecule 1 is a novel autoantigen in membranous lupus nephritis. *Kidney Int* 2021;100:171–81. <https://doi.org/10.1016/j.kint.2020.09.016>.
19. Bajcsi D, Bitó L, Turkevi-Nagy S, Nyári T, Kemény É, Légrády P, et al. The value of PLA2R antigen and IgG subclass staining relative to anti-PLA2R seropositivity in the differential diagnosis of membranous nephropathy. *BMC Nephrol* 2023;24:230. <https://doi.org/10.1186/s12882-023-03273-4>.
20. Beck LH, Bonegio RGB, Lambeau G, Beck DM, Powell DW, Cummins TD, et al. M-type phospholipase A2 receptor as target antigen in idiopathic membranous nephropathy. *N Engl J Med* 2009;361:11–21. <https://doi.org/10.1056/NEJMoa0810457>.
21. Ishizaki J, Suzuki N, Higashino KI, Yokota Y, Ono T, Kawamoto K, et al. Cloning and characterization of novel mouse and human secretory phospholipase A2S. *J Biol Chem* 1999;274:24973–9. <https://doi.org/10.1074/jbc.274.35.24973>.
22. Hossen MS, Shapla UM, Gan SH, Khalil MI. Impact of bee venom enzymes on diseases and immune responses. *Molecules* 2017;22:1–16. <https://doi.org/10.3390/molecules22010025>.
23. Ardalan MR, Ghafari A, Hamzavi F, Nasri H, Baradaran B, Majidi J, et al. Anti-phospholipase A2 receptor antibody in idiopathic membranous nephropathy: A report from Iranian population. *J Nephrothol* 2013;2:241–8.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

<https://doi.org/10.12860/JNP.2013.38>.

24. Debiec H, Hanoy M, Francois A, Guerrot D, Ferlicot S, Johanet C, et al. Recurrent membranous nephropathy in an allograft caused by IgG3k targeting the PLA2 receptor. *J Am Soc Nephrol* 2012;23:1949–54. <https://doi.org/10.1681/ASN.2012060577>.

25. Kudose S, Santoriello D, Debiec H, Canetta PA, Bomback AS, Stokes MB, et al. The clinicopathologic spectrum of segmental membranous glomerulopathy. *Kidney Int* 2021;99:247–55. <https://doi.org/10.1016/j.kint.2020.06.014>.

1
2
3 **Figure legends**
4
5

6 **Fig. 1** Light microscopy of a renal biopsy. (a) Moderate atherosclerotic lesions can be
7
8
9
10 seen (periodic acid–Schiff stain, × 200). (b) A normocellular glomerulus with patent
11
12
13 capillaries without thickening of glomerular capillary walls can be seen (periodic acid–
14
15
16 Schiff stain, ×1000). (c) The apparent spikes or stippling of glomerular basement
17
18
19 membranes cannot be seen in glomerulus (periodic acid–silver methenamine–
20
21
22 hematoxylin and eosin stain, ×1000).
23
24
25
26
27

28 **Fig. 2** Immunofluorescence shows moderate staining for IgG (a), and staining for IgM
29
30
31 (b), C3c (c), and fibrinogen (d). Immunofluorescence using paraffin sections for
32
33
34 phospholipase A₂ receptor (PLA₂R) (e, f: high magnification of the area indicated by the
35
36
37
38 arrow in e) is negative on the glomerular capillary loops, although autofluorescence from
39
40
41 red blood cells was detected.
42
43
44
45
46
47

48 **Fig. 3** Electron microscopy shows the diffuse and partially segmental effacement of the
49
50
51 foot processes of podocytes with microvillous formation and irregular segmental
52
53
54 subepithelial electron-dense deposits (arrowheads) in glomeruli without apparent spike
55
56
57 formation (a,c: ×1500; b,d: ×7000).
58
59
60
61
62
63
64
65

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Fig. 4 Clinical course of the patient

PSL, prednisolone; Cr, creatinine; U-protein, urinary protein; Alb, serum albumin

Fig1

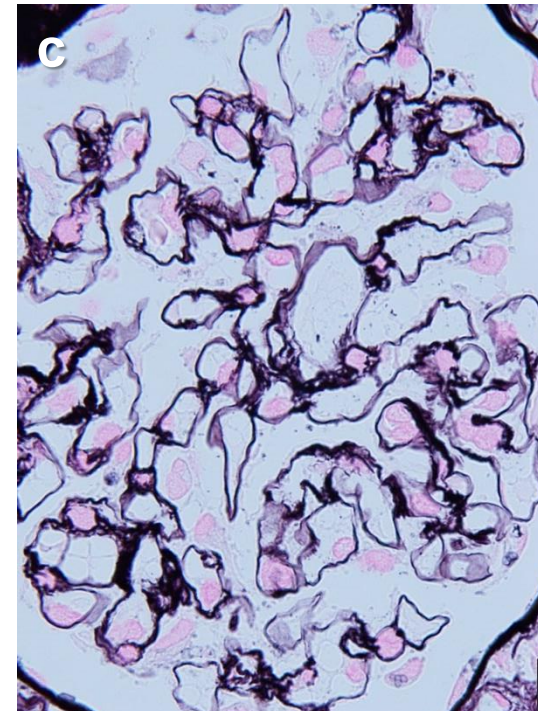
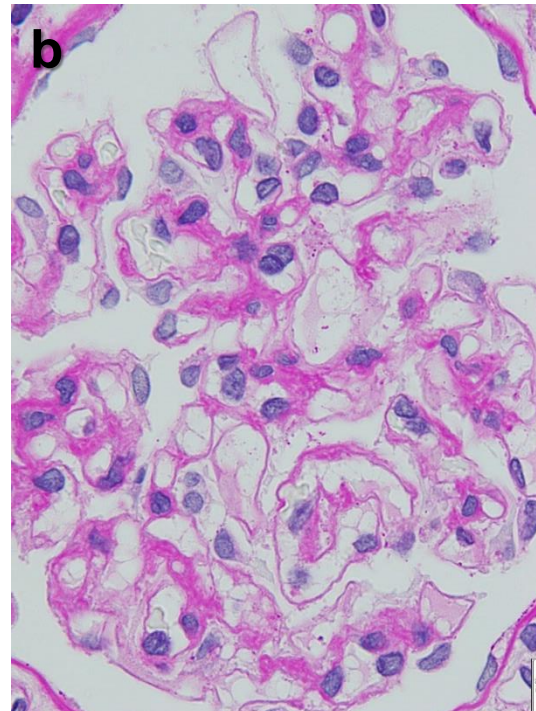
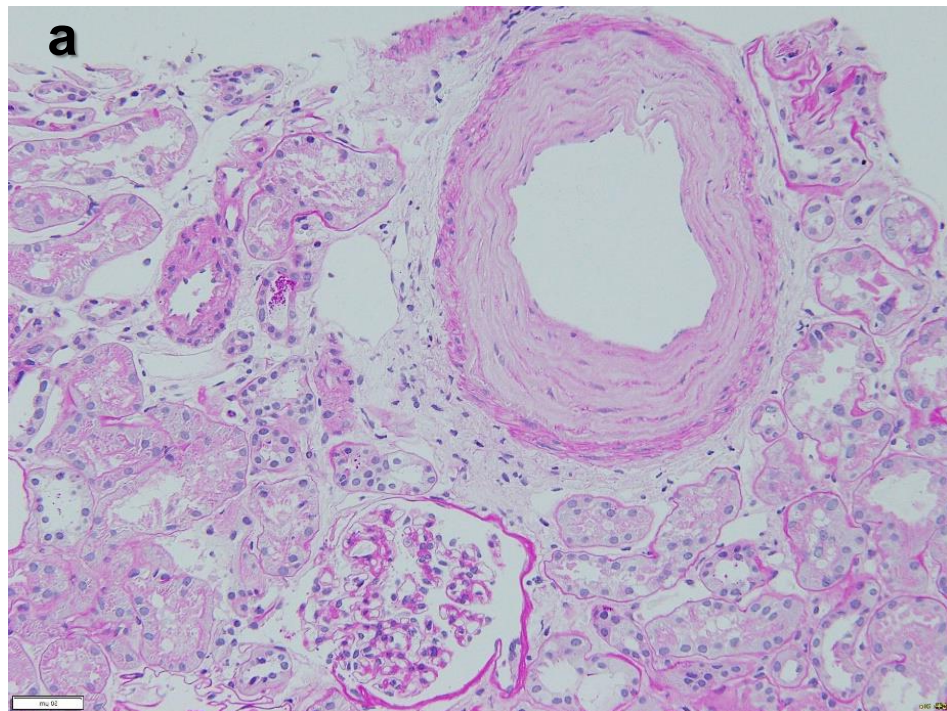


Fig2

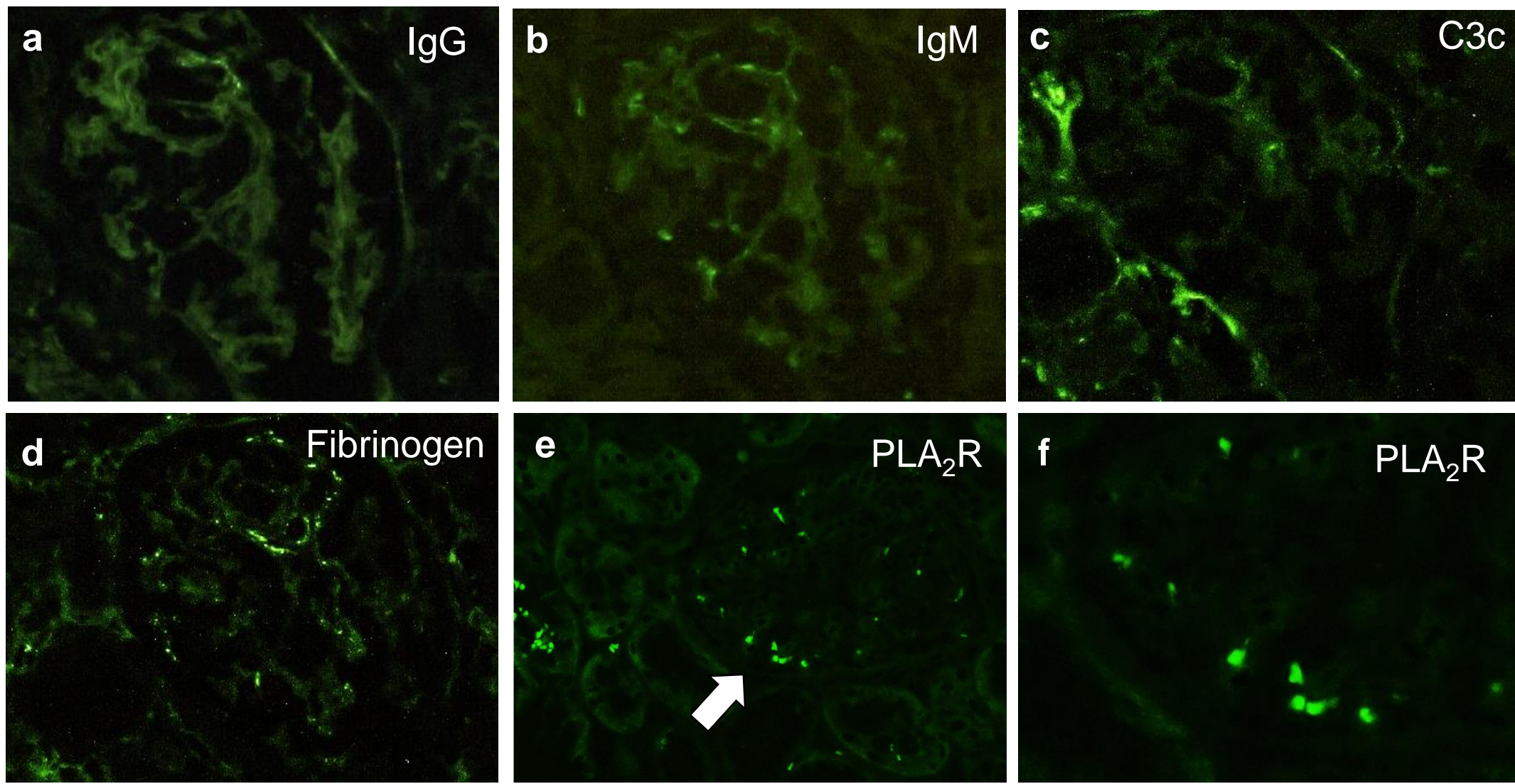


Fig3

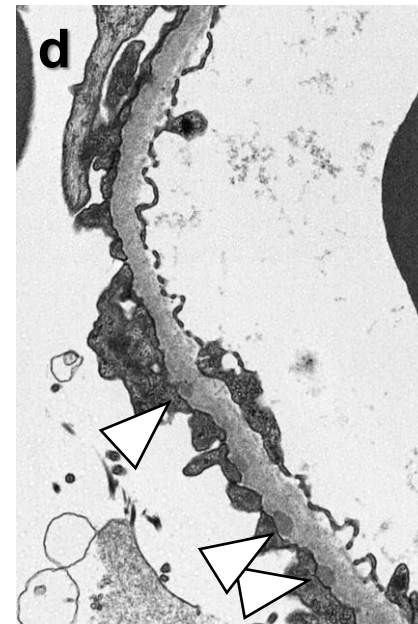
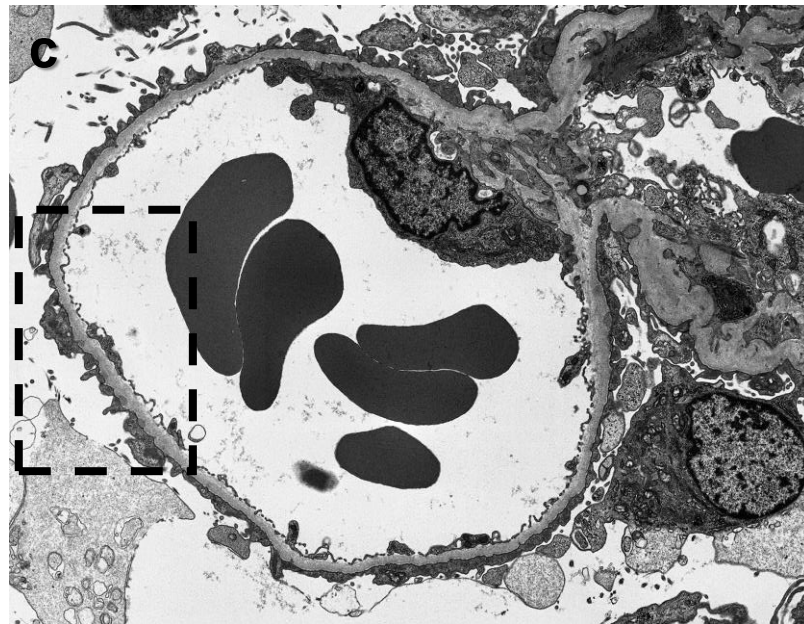
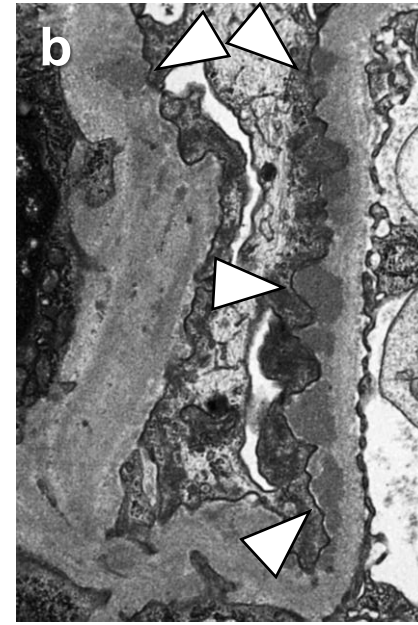
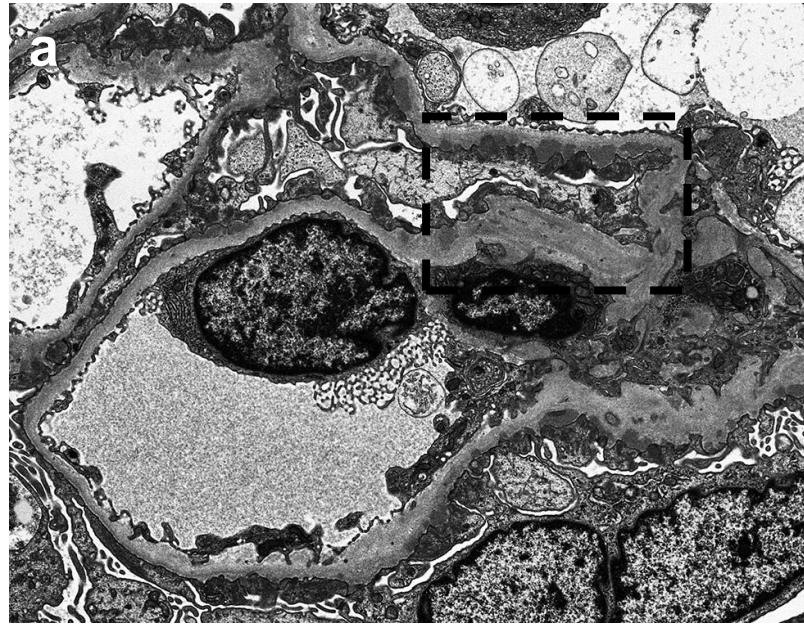


Fig4

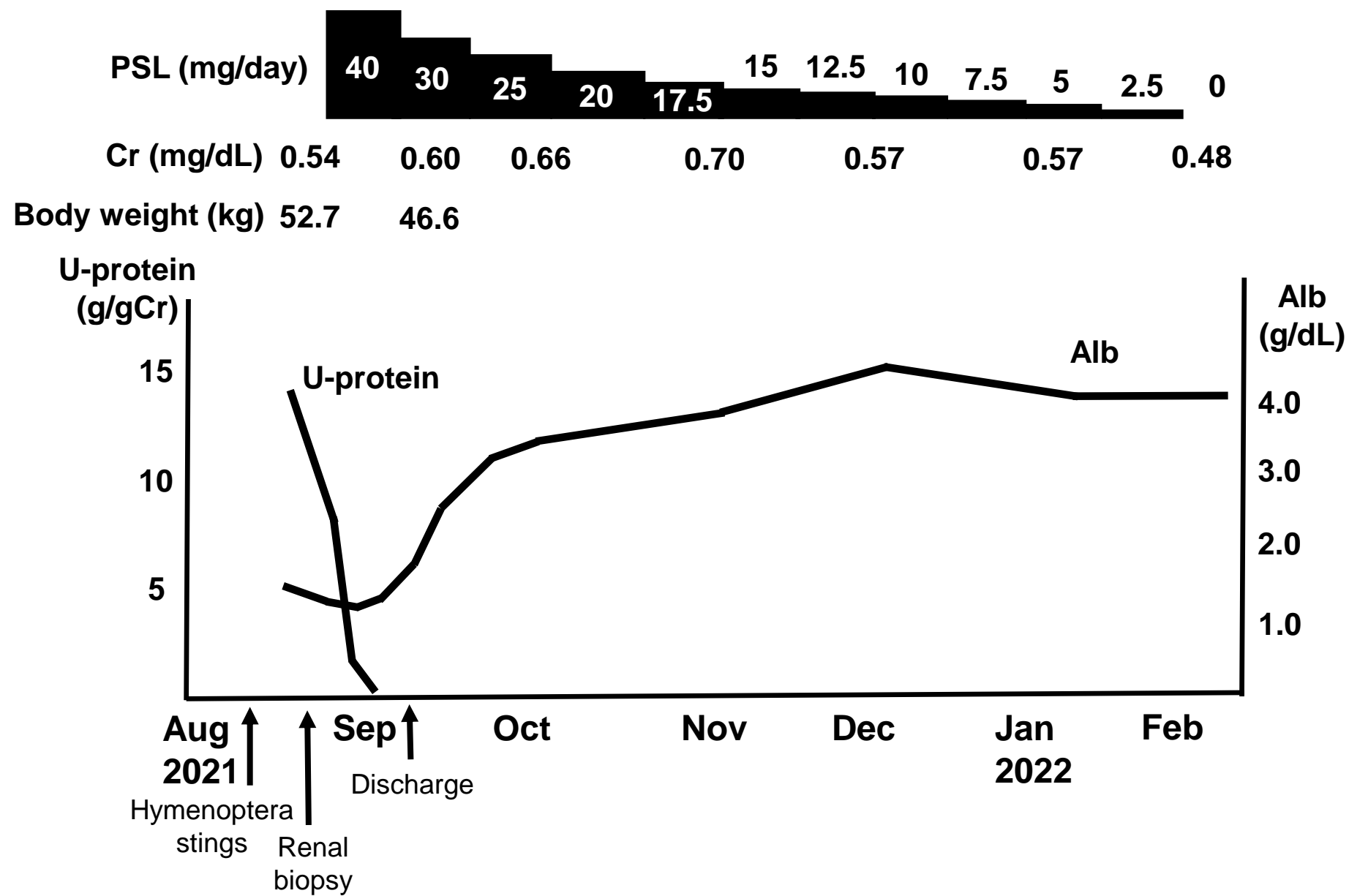


Table 1

Table 1 Laboratory data on admission

<i>Urinalysis</i>		<i>Peripheral blood</i>	
Protein	14.18 g/gCr	WBCs	7800 /mm ³
Occult blood	+ -	Neut	75.7%
Urine sediment		Lymp	17.4%
RBCs	1-4 /HPF	Mono	3.5%
WBCs	0-1 /HPF	Eosino	2.6%
Granular cast	1-4 /WF	Baso	0.8%
Hyaline cast	1-4/WF	RBCs	4.57 x 10 ⁶ /mm ³
β2-MG	402 μg/L	Hb	13.9 g/dL
		Hct	42.5%
<i>Proteinuria selectivity index</i>	0.11	Plt	30.0 x 10 ⁴ /mm ³
<i>Blood chemistry</i>		<i>Immunological studies</i>	
TP	4.9 g/dL	IgG	1128 mg/dL
Alb	1.7 g/dL	IgA	231 mg/dL
TC	401 mg/dL	IgM	92 mg/dL
LDL-C	266 mg/dL	C3	266 mg/dL
HDL-C	107 mg/dL	C4	107 mg/dL
TGs	186 mg/dL	CH50	186 mg/dL
BUN	10 mg/dL	ANA	-
Cr	0.54 mg/dL	HBsAg	-
Na	137 mEq/L	HCVAb	-
K	4.7 mEq/L	Fraction of serum protein	
Cl	106 mEq/L	Alb	44.8%
Ca	10.8 mg/dL	α1-glb	3.6%
P	3.1 mg/dL	α2-glb	21.1%
eGFR	80 mL/min/1.73 m ²	β-glb	12.7%
CRP	0.13 mg/dL	γ-glb	17.8%
FBG	97 mg/dL		

RBCs, red blood cells; WBCs, white blood cells; HPF, high-power field; WF, whole field; β2-MG, β2-microglobulin; TP, total protein; Alb, albumin; TC, total cholesterol; LDL-C, low-density lipoprotein-cholesterol; HDL-C, high-density lipoprotein-cholesterol; TGs, triglycerides; BUN, blood urea nitrogen; Cr, creatinine; Na, sodium; K, potassium; Cl, chloride; Ca, albumin-adjusted calcium; P, phosphorus; eGFR, estimated glomerular filtration rate; CRP, C-reactive protein; FBG, fasting blood glucose; Neut, neutrophils; Lymp, lymphocytes; Mono, monocytes; Eosino, eosinophils; Baso, basophils; Hb, hemoglobin; Hct, hematocrit; Plt, platelets; Ig, immunoglobulin; C, complement; CH50, 50% hemolytic unit of complement; ANA, anti-nuclear antibody; HBsAg, hepatitis B virus s antigen; HCVAb, hepatitis C virus antibody; glb, globulin.

Table 2 Clinical features, pathology of kidney, treatment and outcome of patients with nephrotic syndrome after Hymenoptera stings

Author [reference]	Year	Age (y) / sex	Proteinuria (g/day)	Creatinine (mg/dL)	Pathological findings				Treatment	Outcome
					Diagnosis	LM	IF	EM		
Venters et al. [4]	1961	N.A.	N.A.	N.A.	MCNS (2 cases)	Normal	N.A.	FPE	Cortisone	N.A.
Olivero et al. [5]	1981	22 / F	18	N.A.	MCNS	Normal	All negative	FPE No EDD	PSL	CR
Sensirivatana et al. [10]	1984	5 / M	N.A.	Normal	MPG	Mesangial proliferation	IgM, mesangial area	N.A.	PSL	Relapsed twice
Taber et al. [9]	1986	63 / F	3-4	7.1	PGN	Chronic rejection*	All negative	Proliferating glomerular epithelial cells	Prednisone, HD to PD	Chronic dialysis
Revai et al. [6]	1999	16 / F	49 g/L	N.A.	MCNS	Normal	IgM and C1q, mesangial area	N.A.	mPSL + PSL + CPA + CB	CR
Tauk et al. [11]	1999	28 / M	14	0.7	MPGN Possible early MN	Mesangial increase	IgG, IgM, C3, and C4: mesangial area IgG: capillary loops	FPE Subepithelial deposits	PSL + CPA	ICR
Zaman et al. [7]	2001	21 / M	11.5	0.8	MCNS	Normal	All negative	FPE No EDD	PSL	CR
Humeda et al. [8]	2022	67 / M	10.7	3.5	MCNS	Normal	All negative	FPE No EDD	PSL HD for 3 weeks	CR

LM, light microscopy; IF, immunofluorescence; EM, electron microscopy; N.A., not applicable; F, female; M, male; MCNS, minimal change nephrotic syndrome; MPG, mesangial proliferative glomerulonephritis; PGN, proliferative glomerulonephritis; MPGN, mesangial proliferative glomerulonephritis; MN, membranous nephritis; FPE; foot process effacement; EDD, electron-dense deposits; CR, complete remission; ICR, incomplete remission; PSL, prednisolone; mPSL, methyl prednisolone; CB, chlorambucil; CPA, cyclophosphamide; HD, hemodialysis; PD, peritoneal dialysis.

* Vascular sclerosis, chronic changes in renal tubules, patchy interstitial inflammatory infiltrate, and segmental sclerosis of glomerular mesangium.