

## The Potential for Bidirectional Promoter Activity of the Human PDGF-A Chain Gene

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### ABSTRACT

Platelet-derived growth factor (PDGF) is a heterodimeric glycoprotein consisting of A and B chains. A functional promoter had been identified in the 5' flanking region of the human PDGF-A chain gene. We found that the PDGF-A chain promoter region possesses the potential for bidirectional activity. This bidirectional promoter activity is influenced by the 5'-untranslated region (5'-UTR) and serum concentration. The 5'-UTR may regulate expression of the PDGF-A chain by transcription in the opposite direction.

**Key words:** PDGF-A, Bidirectional promoter

### INTRODUCTION

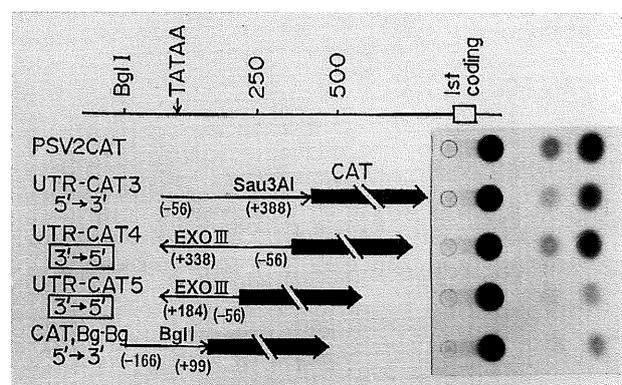
Platelet-derived growth factor (PDGF) is a heterodimeric glycoprotein composed of A and B chains, and is one of the major mitogens of cells derived from mesodermal tissue<sup>7)</sup>. The PDGF-A chain is expressed in many tumor cells<sup>4)</sup>, and abnormal expression of this gene is considered to be closely related to the process of carcinogenesis. The mechanism of regulation of expression of a PDGF-A chain gene is very complex. The 5' regulatory site has been clarified<sup>25)</sup>, but the mechanism of regulation remains to be elucidated. The PDGF-A chain gene possesses the 5'-untranslated region (UTR) of 845 base pair long<sup>25)</sup>, and its involvement in the regulation of expression of the gene has been suggested. In a present study, we found that the PDGF-A chain promoter possesses a bidirectional potential whose activity is influenced by the UTR.

### MATERIALS AND METHODS

**Plasmid construction:** Deletion mutants on the 3' side of the 5'-UTR were constructed using various restriction enzymes and exonuclease III. The initiation site on the 5' side was constructed by partial digestion with nuclease S1 and was 25 bp upstream of TATAA for UTR-CAT3, 5'-3', UTR-CAT4, 3'-5' and UTR-CAT5, 3'-5'. Plasmids shown in Fig. 1 were prepared by fusing pSVO-CAT containing only the structural gene of chloramphenicol acetyl transferase (CAT) and fragments of the 5'-UTR. Whether each fragment was correctly constructed was confirmed by se-

quencing. pSV2CAT included the promoter and enhancer sequences of SV40.

**DNA transfection and CAT assay:** About 24 hours before transfection, RD cells (a human embryonal rhabdomyosarcoma cell line) were seeded at  $5.5 \times 10^5$  cells per 100 mm Petri dish. Twenty micrograms of each plasmid was transfected by the CaPO<sub>4</sub> method. After 3 hours, the cells were treated with 15% (v/v) glycerol in 20 mM Hepes buffer for 3 minutes, washed, and incubated for 48 hours with 0.2%, 10% or 20% FCS. They were then harvested, and 100 µg of lysates was assayed for CAT activity.



**Fig. 1.** Plasmid construction and CAT assay UTR-CAT4, 3'-5', and UTR-CAT5, 3'-5', in which the fragment was inserted in the direction opposite to that of the CAT gene, showed CAT activity. The transcription start site corresponds to the +1.

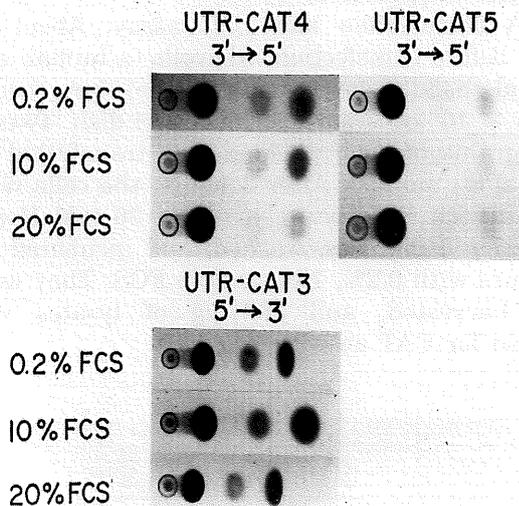
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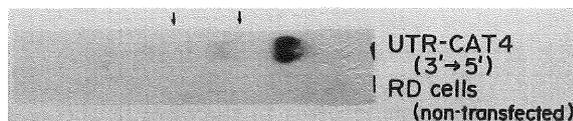
Northern blot analysis: Total cellular RNA was isolated for Northern blot<sup>17</sup>. The probe for the CAT gene was labeled with  $\alpha$ -<sup>32</sup>P dCTP by random primer labeling.

### RESULTS

As shown in Fig. 1, UTR-CAT4, 3'-5', and UTR-CAT5, 3'-5', in which the fragment of 5'-UTR is inserted in the direction opposite to that of the CAT gene, showed CAT activity. CAT activity in UTR-CAT4, 3'-5', possessing a long UTR, was greater than that in UTR-CAT5, 3'-5'. UTR-CAT4, 3'-5', showed a decrease in CAT activity due to stimulation with 20% fetal calf serum, while UTR-CAT5, 3'-5' and UTR-CAT3, 5'-3' were not affected by that concentration of serum (Fig. 2). In the RD cells which were transfected by UTR-CAT4, 3'-5', CAT mRNA was detected as shown in Fig. 3, suggesting that transcription of the CAT gene occurred with the fragment inserted in the direction opposite to that of the CAT gene.



**Fig. 2.** Effect of serum on CAT activity  
The CAT activity of UTR-CAT4, 3'-5', was decreased by treatment with 20% fetal calf serum.



**Fig. 3.** Northern hybridization  
CAT mRNA was observed in a transfectant of UTR-CAT4, 3'-5'.

### DISCUSSION

The present study revealed that the PDGF-A chain promoter region possesses the potential for bidirectional activity. There are several reports on bidirectional promoters: SV40<sup>2,9</sup>, dihydrofolate reductase gene<sup>21</sup>,  $\alpha$ 1(IV) and  $\alpha$ 2(IV) collagen genes<sup>5,19</sup>, proliferating cell nuclear antigen (PCNA) gene<sup>20</sup>, mitochondrial promoters<sup>6</sup>, c-Ha-ras gene<sup>16</sup>, hypoxanthine phosphoribosyl transferase gene and 3-phosphoglycerate kinase gene<sup>12</sup>, and Surf-1 and Surf-2 genes<sup>15</sup>. Although no actual transcript has been demonstrated, the potential for bidirectional promoter activity has been found in the following genes: insulin II gene<sup>8</sup>, 3-hydroxy-3-methylglutaryl coenzyme A (HMG CoA) reductase gene<sup>1</sup>, and HTF 9 gene<sup>14</sup>.

Among genes with a bidirectional promoter, SV40 early and late transcript and  $\alpha$ 1 (IV) and  $\alpha$ 2 (IV) collagen transcript are translated, and they are considered to be divergently transcribed genes. The expression of divergent genes is regulated by a common mechanism, and the products also show functional relationships: e.g., they are involved in a common metabolic pathway, like GAL-1 and GAL-10<sup>13</sup>, and associated polypeptides form protein complexes like  $\alpha$ 1(IV) and  $\alpha$ 2 (IV) collagen<sup>5,19</sup> and histone H2A and H2B<sup>24</sup>.

RNA produced from a bidirectional promoter, whose direction is opposite to that of the structural gene, may also be involved in gene regulation as antisense RNA. In prokaryotes, expression of plasmid Col E1<sup>27</sup>, transposon Tn 10<sup>22</sup> and membrane protein Omp C and Omp F genes<sup>18</sup> is regulated by the antisense RNA. In eukaryotes, the antisense RNA may block RNA elongation for the myc gene<sup>3</sup>. In the case of the PDGF-A chain, the promoter region forms an S1-hypersensitive site, showing a single-strand state<sup>28</sup> where an antisense RNA binding may occur. According to Franklin et al<sup>10</sup> paucimolecular PDGF-B chain mRNA may be the antisense RNA of the major c-sis transcript. With regard to the PDGF-A chain gene, however, no antisense RNA against the transcript has been reported.

The majority of divergently transcribed genes are house keeping genes. They are characterized by a defect in the TATAA box. When the TATAA box is deleted from the gene possessing it, divergent transcription occurs<sup>11</sup>. As a second characteristic, the part surrounding the promoter is G-C rich and a GC box is often present. Even if a TATAA box defect is present, divergent transcription does not occur unless the promoter region is G-C rich<sup>23</sup>. The origin of bidirectional transcription is in the methylation-free island<sup>14</sup>. The surrounding part of the PDGF-A chain promoter is very rich in G-C and it possesses GC boxes as previously reported, but a TATAA box is present. The present analysis revealed that bidirectional

promoter activity is influenced by the 5'-UTR. We have previously found that the 5'-UTR regulates the PDGF-A chain gene at the transcriptional and post-transcriptional levels<sup>26</sup>). The present study suggests that the 5'-UTR further regulates expression of the PDGF-A chain by regulating the transcription in the direction opposite to that of the 5'-UTR.

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## REFERENCES

1. **Abrams, J.M. and Schimke, R.T.** 1989. Chimeric 3-hydroxy-3-methylglutaryl coenzyme A reductase-dihydrofolate reductase gene display bidirectional expression and unidirectional regulation in stably transfected cells. *Mol. Cell. Biol.* **9**: 620-628.
2. **Baty, D., Barrera-Saldana, H.A., Everett, R.D., Vigneron, M. and Chambon, P.** 1984. Mutational dissection of the 21bp repeat region of the SV40 early promoter reveals that it contains overlapping elements of the early-early and late-early promoters. *Nucl. Acids Res.* **12**: 919-935.
3. **Bentley, D.L. and Groudine, M.** 1986. A block to elongation is largely responsible for decreased transcription of c-myc in differentiated HL60 cells. *Nature* **321**: 702-706.
4. **Betsholtz, C., Johnsson, A., Heldin, C.-H., Westermark, B., Lind, P., Urdea, M.S., Eddy, R., Shows, T.B., Philpott, K., Mellor, A.L., Knott, T.J. and Scott, J.** 1986. cDNA sequence and chromosomal localization of human platelet-derived growth factor A-chain and its expression in tumor cell lines. *Nature* **320**: 695-699.
5. **Burbelo, P.D., Martin, G.R. and Yamada, Y.** 1988.  $\alpha 1$  (IV) and  $\alpha 2$  (IV) collagen genes are regulated by a bidirectional promoter and shared enhancer. *Proc. Natl. Acad. Sci. USA* **85**: 9679-9682.
6. **Chang, D.D., Hixson, J.E. and Clayton, D.A.** 1986. Minor transcription initiation events indicate that both human mitochondrial promoters function bidirectionally. *Mol. Cell. Biol.* **6**: 294-301.
7. **Deuel, T.F.** 1987. Polypeptide growth factors : roles in normal and abnormal cell growth. *Ann. Rev. Cell. Biol.* **3**: 443-492.
8. **Efrat, S. and Hanahan, D.** 1987. Bidirectional activity of the rat insulin II 5'-flanking region in transgenic mice. *Mol. Cell. Biol.* **7**: 192-198.
9. **Everett, R.D., Baty, D. and Chambon, P.** 1983. The repeat G-C rich motifs upstream from the TATAA box are important elements of the SV40 early promoter. *Nucl. Acids Res.* **11**: 2447-2464.
10. **Franklin, G.C., Donovan, M., Adam, G.I.R., Holmgren, L., Pfeifer-Ohlsson, S. and Ohlsson, R.** 1991. Expression of the human PDGF-B gene is regulated by both positively and negatively acting cell type-specific regulatory elements located in the first intron. *EMBO J.* **10**: 1365-1373.
11. **Grichnik, J.M., French, B.A. and Schwartz, R. J.** 1988. The chicken skeletal alpha-actin promoter region partial dyad symmetry and a capacity to drive bidirectional transcription. *Mol. Cell. Biol.* **8**: 4587-4597.
12. **Johnson, P. and Friedmann, T.** 1990. Limited bidirectional activity of two housekeeping gene promoters: human HPRT and PGK. *Gene* **88**: 207-213.
13. **Johnston, M. and Davies, R.W.** 1984. Sequences that regulate the divergent GAL1-GAL10 promoter in *Saccharomyces cerevisiae*. *Mol. Cell. Biol.* **4**: 1440-1448.
14. **Lavia, P., Macleod, D. and Bird, A.** 1987. Coincident start sites for divergent transcripts at a randomly selected CpG-rich island of mouse. *EMBO J.* **6**: 2773-2779.
15. **Lennard, A.C. and Fried, M.** 1991. The bidirectional promoter of the divergently transcribed mouse Surf-1 and Surf-2 genes. *Mol. Cell. Biol.* **11**: 1281-1294.
16. **Lowndes, N.F., Paul, J., Wu, J. and Allan, M.** 1989. c-Ha-ras gene bidirectional promoter expressed in vitro: location and regulation. *Mol. Cell. Biol.* **9**: 3758-3770.
17. **Maniatis, T., Fritsch, E.F. and Sambrook, J.** 1982. *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor Lab., Cold Spring Harbor, NY.
18. **Mizuno, T., Chou, M.-Y. and Inoue, M.** 1984. A unique mechanism regulating gene expression: translational inhibition by a complementary RNA transcript (mic RNA). *Proc. Natl. Acad. Sci. USA* **81**: 1966-1970.
19. **Poschl, E., Pollner, R. and Kuhn, K.** 1988. The genes for the 1(IV) and 2(IV) chains of human basement membrane collagen type IV are arranged head-to-head and separated by a bidirectional promoter of unique structure. *EMBO J.* **7**: 2687-2695.
20. **Rizzo, M.G., Ottavio, L., Travali, S., Chang, C.-D., Kaminska, B. and Baserga, R.** 1990. The promoter of the human proliferating cell nuclear antigen (PCNA) gene is bidirectional. *Exp. Cell Res.* **188**: 286-293.
21. **Shimada, T., Fujii, H. and Lin, H.** 1989. A 165-base pair sequence between the dihydrofolate reductase gene and the divergently transcribed upstream gene is sufficient for bidirectional transcriptional activity. *J. Biol. Chem.* **264**: 20171-20174.
22. **Simons, R.W. and Kleckner, N.** 1983. Translational control of IS10 trans position. *Cell* **34**: 683-691.
23. **Smale, S.T. and Baltimore, D.** 1989. The initiator as a transcriptional control element. *Cell* **57**: 103-113.
24. **Sturm, R.A., Dalton, S. and Wells, J.R.E.** 1988. Conservation of histone H2A/H2B intergene regions: a role for the H2B specific element in divergent transcription. *Nucl. Acids Res.* **16**: 8571-8586.
25. **Takimoto, Y., Wang, Z.Y., Kobler, K. and Deuel, T.F.** 1991. Promoter region of the human platelet-derived growth factor A-chain gene. *Proc. Natl. Acad. Sci. USA* **88**: 1686-1690.
26. **Takimoto, Y. and Kuramoto, A.** 1994. Gene

regulation by the 5'-untranslated region of the platelet-derived growth factor A-chain. *Biochem. Biophys. Acta* **1222**: 511–514.

27. **Tomizawa, J., Itoh, T., Selzer, G. and Som, T.** 1981. Inhibition of Col E1 primer formation by a plasmid-specified small RNA. *Proc. Natl. Acad. Sci. USA* **78**: 1421–1425.
28. **Wang, Z.-Y., Lin, X.-H., Nobyuoshi, M., Qiu, Q.-Q. and Deuel, T.F.** 1992. Binding of single-stranded oligonucleotides to a non-B-form DNA structure results in loss of promoter activity of the platelet-derived growth factor A-chain gene. *J. Biol. Chem.* **267**: 13669–13674.