

Natural Occurrence and Chemical Synthesis of Bile Alcohols, Higher Bile Acids, and Short Side Chain Bile Acids

Mizuho UNE and Takahiko HOSHITA

*Institute of Pharmaceutical Sciences, Hiroshima University School of Medicine, 1-2-3 Kasumi,
Minami-ku, Hiroshima, 734, Japan*

Key words: *Bile alcohols, Higher bile acids, Short side chain bile acids*

Both bile acids and bile alcohols, a subclass of steroids, are made from cholesterol as its major end metabolites by the liver of vertebrates. Their conjugates are the major constituents of the bile and possess a special function as an aid to intestinal lipid digestion and absorption.

The most common naturally occurring bile acids in mammals, birds, most snakes, and various teleostean fishes, are cholic acid, chenodeoxycholic acid, deoxycholic acid, and lithocholic acid. In addition to these 3 α -, 7 α -, or 12 α -hydroxy derivatives of 5 β -cholan-24-oic acid, a number of 5 β -cholan-24-oic acids carrying a hydroxyl group at C-1, C-2, C-4, C-6, C-16, C-22, or C-23 position, a β -oriented hydroxyl group, or a keto group have been found in the bile, serum, urine and feces of these vertebrates. 5 α -Cholan-24-oic acid derivatives occur as a high proportion of the bile acids in some lizards and in lesser amounts in various vertebrates.

Biles of evolutionarily primitive vertebrates such as some lizards, crocodiles, alligators, turtles, tortoises, all amphibians, certain bony fishes, sharks, and rays contain, in place of the C₂₄ bile acid conjugates, other types of bile salts, sulfate esters of bile alcohols and unconjugated or taurine-conjugated bile acids with more than 24 carbon atoms (higher bile acids). Bile alcohols and higher bile acids have the C₂₄ bile acid type of nuclear structure and all or part of the cholesterol type side chain in their carbon skeleton. Although the distribution of bile alcohols and higher bile acids in nature was long considered to be confined to the primitive vertebrates, it is now recognized that these compounds are accumulated in patients with inherited diseases related to abnormal cholesterol metabolism. They are present in trace amounts even in healthy humans. Furthermore, a few 4- or 5-cholen-24-oic acid derivatives and a few C₂₂ and C₂₃ bile acids with the shortened side chain have been found in biological fluids of healthy and diseased humans.

Several excellent reviews about the naturally occurring bile acids and bile alcohols have appeared^{61, 74, 75, 101, 105, 183}. It should be men-

tioned, however, that there is no review covering all the known bile acids and bile alcohols including chemically synthetic derivatives. The present review is a detailed tabulation of bile alcohols, higher bile acids, and short side chain bile acids. Derivatives of 5 α - and 5 β -cholan-24-oic acids and of 4- and 5-cholen-24-oic acids are not included in this review. Natural occurrence and chemical synthesis of these C₂₄ bile acids are the subject of our forthcoming review.

Bile Alcohols

Bile alcohols are here defined as neutral steroid-alcohols biochemically related bile acids.

Tables 1-6 list all the naturally occurring bile alcohols of established structure and most chemically derived C₂₇-C₂₂ bile alcohols. Artifacts formed from native bile alcohol sulfates by alkaline hydrolysis, dehydro-derivatives having no hydroxyl group, esterified derivatives, and derivatives containing other elements than oxygen, e.g. nitrogen, sulfur, or halogens, are not included in the tables. Mono- and di-oxygenated cholestan derivatives, e.g. 7 α -hydroxycholesterol, are not listed in the tables because they are usually called sterols rather than bile alcohols. Similarly, polyoxygenated steroids found in marine invertebrates and shark-repelling and ichthyotoxic steroids obtained from some fishes are also excluded.

The trivial names of bile alcohols, isolated from the bile of vertebrates other than humans, include as a prefix part of the Latin name of the genus of the animal from which they were first isolated. When both isomers differing in the configuration at C-5 are known, the prefix 5 α or 5 β is then used with the original name. For bile alcohols with epimeric hydroxyl groups for which there is no accepted trivial name, the prefix "epi" is used preceded by a number and a hyphen, indicating the position at which the epimeric hydroxyl group is present, e.g. 3-epimyxinol. For deoxy derivatives, the prefix "deoxy" will be used, preceded by a number and a hyphen, indicating the place where the hydroxyl group is missing,

e.g. 26-deoxy- 5β -ranol. Bile alcohols found only in the biological fluids of humans and unnatural

bile alcohols have no trivial names; they are usually designated their systematic names.

Table 1. Hexa-oxygenated C₂₇ bile alcohols

No.	Systematic name (Trivial name)	Natural source ^a	Synthetic source ^b
0101	5 β -Cholestane-2 β , 3 α , 7 α , 12 α , 26,27-hexol (Arapaimol-B)	<i>Arapaima gigas</i> ⁷⁸⁾	— ^c
0102	5 β -Cholestane-3 α , 7 α , 12 α , 23, 24, 25-hexol	CTX-U ²²⁶⁾	—
0103	5 β -Cholestane-3 α , 7 α , 12 α , 24, 25, 26-hexol	Human-U ^{82, 83)} , -S ⁸⁴⁾ ; CTX-U ²⁵¹⁾ ; LD-U ⁸³⁾	0311 ⁸²⁾ ; 0312 ⁸²⁾
0104	(24R)-5 β -Cholestane-3 α , 7 α , 12 α , 24, 26, 27-hexol (Scymnol)	Sharks and rays ^{32, 111, 112, 121, 158)}	CA ³²⁾
0105	5 α -Cholestane-3 α , 7 α , 12 α , 25, 26, 27-hexol (5 α -Dermophol)	Amphibians ¹³⁶⁾	0325 ¹³⁷⁾
0106	5 β -Cholestane-3 α , 7 α , 12 α , 25, 26, 27-hexol (5 β -Dermophol)	Amphibians ¹³⁶⁾	0326 ¹³⁶⁾

a. All natural sources refer to bile, unless otherwise referred to specific source details, which are shown by the following abbreviations: U, urine; S, serum; F, feces; L, liver; AF, amniotic fluid; GC, gastric content; M, meconium; UCB, umbilical cord blood.

Diseases are shown by the following abbreviations: CTX, cerebrotendinous xanthomatosis; HS, hyper-sitosterolemia; HSDD, 3 α -hydroxysteroid dehydrogenase deficiency; LD, liver diseases; ZS, Zellweger syndrome; CHO, cholestasis; RF, Refsum disease; COA, coprostanic acidemia; THD, thiolase deficiency; β OD, β -oxidation deficiency; IO, intestinal obstruction; NALD, neonatal adrenoleukodystrophy; IBDA, intrahepatic bile duct anomalies; IMA, intestinal malabsorption; SH, subdural hematoma.

b. Bile acids and bile alcohols having no trivial names are shown as their compound numbers which are given in the first column of each tables.

Abbreviations for bile acids and bile alcohols (systematic name or compound number in parentheses): CA, cholic acid (3 α , 7 α , 12 α -trihydroxy-5 β -cholan-24-oic acid); DCA, deoxycholic acid (3 α , 12 α -dihydroxy-5 β -cholan-24-oic acid); CDCA, chenodeoxycholic acid (3 α , 7 α -dihydroxy-5 β -cholan-24-oic acid); UDCA, ursodeoxycholic acid (3 α , 7 β -dihydroxy-5 β -cholan-24-oic acid); HDCA, hyodeoxycholic acid (3 α , 6 α -dihydroxy-5 β -cholan-24-oic acid); LCA, lithocholic acid (3 α -hydroxy-5 β -cholan-24-oic acid); 22-OH-CDCA, 22R-hydroxychenodeoxycholic acid (formerly haemulcholic acid, (22R)-3 α , 7 α , 22-trihydroxy-5 β -cholan-24-oic acid)^{99, 139)}; 23-OH-CDCA, 23R-hydroxychenodeoxycholic acid (formerly phocaecholic acid, (23R)-3 α , 7 α , 23-trihydroxy-5 β -cholan-24-oic acid)²⁰⁸⁾; 6-OH-7-ODCA, 6 α -hydroxy-7-oxodeoxycholic acid (3 α , 6 α , 12 α -trihydroxy-7-oxo-5 β -cholan-24-oic acid)²³⁵⁾; ACA, allocholic acid (3 α , 7 α , 12 α -trihydroxy-5 α -cholan-24-oic acid)¹⁸⁵⁾; 7-EACA, 7-epiallocholic acid (3 α , 7 β , 12 α -trihydroxy-5 α -cholan-24-oic acid)⁸⁾; NCA, norcholic acid (1204); NHDCDA, norhyodeoxycholic acid (1213); NCDCA, norchenodeoxycholic acid (1214); NUDCA, norursodeoxycholic acid (1215); NDCA, nordeoxycholic acid (1216); NLCA, norlithocholic acid (1218); BNCA, bisnorcholic acid (1303); BNDCA, bisnordeoxycholic acid (1308); HoCA, homocholic acid (1107); HoCDCA, homochenodeoxycholic acid (1114); CAld, cholyl aldehyde (0613); CDCAld, chenodeoxycholyl aldehyde (0620); NCAld, norcholyl aldehyde (0624); BNCAld, bisnorcholyl aldehyde (0640); CY, 5 α -cyprinol (0222); DECY, 27-deoxy-5 α -cyprinol (0323); ANSC, anhydroscymnol (24, 26-epoxy-5 β -cholestane-3 α , 7 α , 12 α , 27-tetrol)⁶⁵⁾; ANCY, anhydro-5 α -cyprinol (26, 27-epoxy-5 α -cholestane-3 α , 7 α , 12 α -triol)⁹⁰⁾; DNCT, (26, 27-dinor-5 α -cholest-24-ene-3 α , 7 α , 12 α -triol)¹⁷⁹⁾; TADNC, (3 α , 7 α , 12 α -triformoxy-25-acetoxy-26, 27-dinor-5 β -cholestan-24-one)²¹⁴⁾.

c. not reported.

Table 2. Penta-oxygenated C₂₇ bile alcohols

No.	Systematic name (Trivial name)	Natural source	Synthetic source
0201	5β-Cholestane-2β, 3α, 7α, 12α, 26-pentol (Arapaimol-A)	<i>Arapaima gigas</i> ⁷⁸⁾ ; Frogs ¹⁴⁾	—
0202	5β-Cholestane-3α, 6α, 7β, 25, 26-pentol (ω-Trichechol)	Manatee ¹⁷³⁾	—
0203	5β-Cholestane-3α, 6β, 7α, 25, 26-pentol (α-Trichechol)	Manatee ¹⁷³⁾ ; Rat ⁸⁶⁾	1105 ²⁵⁶⁾
0204	5β-Cholestane-3α, 6β, 7β, 25, 26-pentol (β-Trichechol)	Manatee ¹⁷³⁾	—
0205	(22R)-5β-Cholestane-3α, 7α, 12α, 22, 25-pentol	CTX ²²⁶⁾ , -U ^{226, 251)}	BNCALd ¹³⁵⁾
0206	(22S)-5β-Cholestane-3α, 7α, 12α, 22, 25-pentol	—	BNCALd ¹³⁵⁾
0207	(22R, 25R)-5β-Cholestane-3α, 7α, 12α, 22, 26-pentol	—	0806 ²⁴⁷⁾
0208	(22S, 25R)-5β-Cholestane-3α, 7α, 12α, 22, 26-pentol	—	0806 ²⁴⁷⁾
0209	(23S)-5α-Cholestane-3α, 7α, 12α, 23, 25-pentol	CTX-U ¹⁴⁹⁾	—
0210	(23R)-5β-Cholestane-3α, 7α, 12α, 23, 25-pentol	CTX-U ²²⁶⁾	0306 ¹⁴¹⁾ ; 1104 ^{103, 146)}
0211	(23S)-5β-Cholestane-3α, 7α, 12α, 23, 25-pentol	CTX ^{104, 225, 226)} , -U ^{226, 251)} , -F ^{225, 226, 255)}	0307 ¹⁴¹⁾ ; 1104 ^{103, 146)}
0212	(23R)-5β-Cholestane-3α, 7α, 12α, 23, 26-pentol	—	0306 ²⁴⁷⁾
0213	(23S)-5β-Cholestane-3α, 7α, 12α, 23, 26-pentol	—	0307 ²⁴⁷⁾
0214	(24R)- and (24S)-5β-Cholestane-3α, 7α, 12α, 24, 25-pentols	Human ¹⁷²⁾ , -S ⁸⁴⁾ , -U ^{82, 83, 174)} , CTX ^{104, 221, 225, 226)} , -U ^{226, 251)} , -F ^{221, 225, 226, 255)} , HS-F, U ⁵⁵⁾ LD-U ⁸³⁾	0405 ^{44, 91)}
0215	Cholest-5-ene-3β, 7α, 12α, 24, 25-pentol	HSDD ¹⁰⁸⁾	—
0216	5α-Cholestane-3α, 7α, 12α, 24, 26-pentol (5α-Chimaerol)	White sucker ¹³⁾ ; Fishes ¹⁵⁾ ; Lungfish ⁶⁾	—
0217	5β-Cholestane-3α, 7α, 12α, 24, 26-pentol (5β-Chimaerol)	<i>Chimaera monstrosa</i> ³³⁾ ; Sharks and rays ^{32, 112, 202)} ; Rat ⁸⁶⁾ ; Human ¹⁷²⁾ , -S ⁸⁴⁾ , -U ^{82, 83)} , CHO-U ^{107, 120)} , LD-U ⁸³⁾	ANS ^{C42)}
0218	5α-Cholestane-3α, 7α, 12α, 25, 26-pentol (5α-Bufol)	Coelacanth ^{6, 142)} ; Lungfish ⁶⁾ ; Frogs ^{14, 240)} ; Newt ⁹⁸⁾ ; Rat ⁸⁶⁾	0403 ⁹⁸⁾
0219	5α-Cholestane-3β, 7α, 12α, 25, 26-pentol	Coelacanth ¹⁴²⁾	0404 ¹⁴²⁾
0220	5β-Cholestane-3α, 7α, 12α, 25, 26-pentol (5β-Bufol)	Toads and frogs ^{14, 97, 161, 166, 201,} ^{240, 257)} ; Rat ⁸⁶⁾ ; Human ¹⁷²⁾ , -S ^{84,} ⁸⁵⁾ , -U ^{82, 83, 174, 177)} ; CTX-U ²²⁶ , -F ⁵²⁾ ; CHO-U ^{107, 120)} ; HS-U, F ⁵⁵⁾ ; LD-U ^{83, 177)}	HoCA ¹⁴⁶⁾ ; 0406 ⁹¹⁾
0221	Cholest-5-ene-3β, 7α, 12α, 25, 26-pentol	HSDD ¹⁰⁸⁾	—
0222	5α-Cholestane-3α, 7α, 12α, 26, 27-pentol (5α-Cyprinol)	Carp and related fishes ^{9, 15, 93, 94,} ²¹⁶⁾ ; White sucker ¹³⁾ ; Coelacanth ^{6,} ¹⁴²⁾ ; Lungfishes ⁶⁾ ; Giant salamander ⁵⁾ ; Frogs ^{14, 161, 197, 240)}	—
0223	5α-Cholestane-3β, 7α, 12α, 26, 27-pentol (Latimerol)	Coelacanth ^{6, 10, 142)}	—
0224	5β-Cholestane-3α, 7α, 12α, 26, 27-pentol (5β-Cyprinol)	Fishes ^{15, 43, 76, 95)} ; Toads and frogs ^{14, 131, 161, 240)} ; Human ¹⁷²⁾ , -U ¹⁷⁴⁾ ; CHO-U ^{106, 107)}	CA ^{76, 92)}

For abbreviations, see Table 1.

Table 3. Tetra-oxygenated C₂₇ bile alcohols

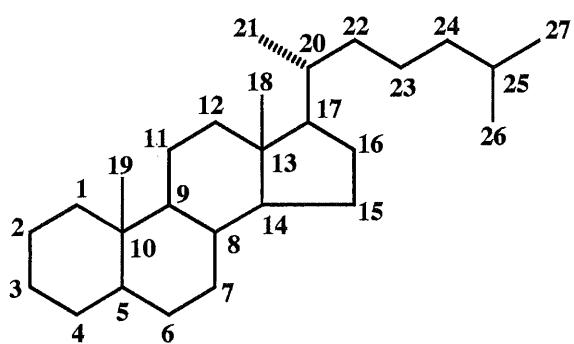
No.	Systematic name (Trivial name)	Natural source	Synthetic source
0301	5β-Cholestane-3α, 6β, 7α, 26-tetrol	Manatee ¹⁷³⁾	—
0302	(22R)-5β-Cholestane-3α, 7α, 12α, 22-tetrol	—	BNCAlD ¹⁶⁵⁾
0303	(22S)-5β-Cholestane-3α, 7α, 12α, 22-tetrol	—	BNCAlD ¹⁶⁵⁾
0304	(23R)-5β-Cholestane-3α, 7α, 12α, 23-tetrol	CTX ^{104, 226)} , -U ^{226, 251)} , -F ^{140, 226, 255)}	0307 ¹⁴¹⁾ , 0308 ¹⁶⁵⁾
0305	(23S)-5β-Cholestane-3α, 7α, 12α, 23-tetrol	—	0308 ¹⁶⁵⁾
0306	(23R)-5β-Cholest-25-ene-3α, 7α, 12α, 23-tetrol	—	NCAlD ¹⁴¹⁾
0307	(23S)-5β-Cholest-25-ene-3α, 7α, 12α, 23-tetrol	—	NCAlD ¹⁴¹⁾
0308	3α, 7α, 12α-Trihydroxy-5β-cholestane-23-one	—	NCA ¹⁶⁵⁾
0309	5α-Cholestane-3α, 7α, 12α, 24-tetrol	Fishes ¹⁵⁾ ; Lungfish ⁶⁾ ; Rabbit ¹⁹³⁾ ; Human ¹⁷²⁾ ; CTX ²²⁶⁾	—
0310	(24R)- and (24S)-5β-Cholestane-3α, 7α, 12α, 24-tetrols	Rabbit ¹⁹³⁾ ; Human ¹⁷²⁾ ; CTX ^{104, 226)} , -U ²²⁶⁾ , -F ^{226, 255)} ; HS-U, F ⁵⁵⁾	0313 ^{180, 227)} , 0809 ²⁴¹⁾ ; 0405 ⁴⁸⁾
0311	(24R)-5β-Cholest-25-ene-3α, 7α, 12α, 24-tetrol	—	CAld ⁸²⁾
0312	(24S)-5β-Cholest-25-ene-3α, 7α, 12α, 24-tetrol	—	CAld ⁸²⁾
0313	3α, 7α, 12α-Trihydroxy-5β-cholestane-24-one	—	CA ²²⁷⁾
0314	Cholest-5-ene-3β, 7α, 12α, 24-tetrol	HSDD ¹⁰⁸⁾	—
0315	5α-Cholestane-3α, 7α, 12α, 25-tetrol	CTX-U, F ²²⁶⁾	0403 ²²⁶⁾
0316	5β-Cholestane-3α, 7α, 12α, 25-tetrol	Alligator ²³⁷⁾ ; Rabbit ¹⁹³⁾ ; Human ¹⁷²⁾ , -S ¹⁵⁷⁾ , -L ²⁰⁰⁾ ; CTX ^{104, 200, 221, 226)} , -S ¹⁵⁷⁾ , -U ^{226, 251)} , -F ^{221, 226, 255)} , -L ²⁰⁰⁾ ; HS-U, F ⁵⁵⁾	HoCA ^{44, 50, 116, 146, 207)}
0317	5β-Cholestane-3β, 7α, 12α, 25-tetrol	—	0319 ⁵³⁾
0318	5β-Cholestane-3α, 7β, 12α, 25-tetrol	—	1110 ²²⁶⁾
0319	7α, 12α, 25-Trihydroxy-5β-cholestane-3-one	—	0316 ¹¹⁶⁾
0320	3α, 12α, 25-Trihydroxy-5β-cholestane-7-one	CTX ²²⁶⁾ , -F ²²⁶⁾	0316 ²²⁶⁾
0321	Cholest-5-ene-3β, 7α, 12α, 25-tetrol	HSDD ¹⁰⁸⁾	—
0322	7α, 12α, 25-Trihydroxycholest-4-en-3-one	—	0319 ¹¹⁶⁾
0323	5α-Cholestane-3α, 7α, 12α, 26-tetrol (27-Deoxy-5α-cyprinol)	Carp ⁹⁶⁾ ; Fishes ¹⁵⁾ ; Lungfishes ⁶⁾ ; Toads ^{97, 257)} ; Frogs ¹⁴⁾ ; Alligator ²³⁷⁾ ; Rat ⁸⁶⁾	ANCY ⁹⁰⁾
0324	(25R)- and (25S)-5β-cholestane-3α, 7α, 12α, 26-tetrols	Toads and frogs ^{14, 97, 240, 257)} ; Rat ⁸⁶⁾ ; Human ¹⁷²⁾ ; CHO-U ^{106, 107)}	0902 ^{13, 51, 63, 130)} ; 0406 ⁴⁸⁾
0325	5α-Cholest-25-ene-3α, 7α, 12α, 27-tetrol	—	0909 ¹³⁷⁾
0326	5β-Cholest-25-ene-3α, 7α, 12α, 27-tetrol	—	0910 ¹³⁶⁾
0327	3α, 7α, 12α-Trihydroxy-5β-cholestane-26-al	—	0324 ⁵¹⁾ ; CAld ²⁰³⁾
0328	7α, 12α, 26-Trihydroxy-5β-cholestane-3-one	—	0324 ⁵¹⁾
0329	7α, 12α-Dihydroxy-3-oxo-5β-cholestane-26-al	—	0324 ⁵¹⁾
0330	Cholest-5-ene-3β, 7α, 12α, 26-tetrol	HSDD ¹⁰⁸⁾	—
0331	5α-Cholestane-3α, 7α, 16α, 26-tetrol (3-Epimyxinol)	Hagfish ²³⁹⁾	—
0332	5α-Cholestane-3β, 7α, 16α, 26-tetrol (Myxinol)	Hagfishes ^{11, 73, 239)}	—
0333	5β-Cholestane-3α, 7α, 24, 25-tetrol	—	0414 ^{20, 164)}
0334	5β-Cholestane-3α, 7α, 24, 26-tetrol	Human ²⁴⁵⁾	0917 ²⁴⁵⁾
0335	5β-Cholestane-3α, 7α, 25, 26-tetrol	Manatee ¹⁷³⁾ ; Human ²⁴⁵⁾	0414 ¹⁶⁴⁾
0336	Cholest-5-ene-3β, 7α, 25, 26-tetrol	HSDD ¹⁰⁸⁾	—
0337	5α-Cholestane-3α, 7α, 26, 27-tetrol	—	CY ¹⁶¹⁾
0338	5β-Cholestane-3α, 7α, 26, 27-tetrol	Human ²⁴⁵⁾	1005 ²⁴⁵⁾

For abbreviations, see Table 1.

Table 4. Tri-oxygenated C₂₇ bile alcohols

No.	Systematic name (Trivial name)	Natural source	Synthetic source
0401	5 α -Cholestane-3 α , 7 α , 12 α -triol	—	0403 ¹⁰²⁾
0402	5 β -Cholestane-3 α , 7 α , 12 α -triol	Human-L ²⁰⁰⁾ , CTX-F ^{200, 226)} , -L ²⁰⁰⁾	CA ^{26, 127)} , 0313 ^{219, 227)}
0403	5 α -Cholest-25-ene-3 α , 7 α , 12 α -triol	—	DECY ⁹⁸⁾
0404	5 α -Cholest-25-ene-3 β , 7 α , 12 α -triol	—	0403 ¹⁴²⁾
0405	5 β -Cholest-24-ene-3 α , 7 α , 12 α -triol	—	0316 ^{44, 48, 91)}
0406	5 β -Cholest-25-ene-3 α , 7 α , 12 α -triol	CTX ²⁰⁰⁾	0316 ^{44, 48, 91)}
0407	7 α , 12 α -Dihydroxy-5 α -cholestane-3-one	—	0401 ¹⁰²⁾
0408	7 α , 12 α -Dihydroxy-5 β -cholestane-3-one	—	0402 ²⁷⁾
0409	3 α , 12 α -Dihydroxy-5 β -cholestane-7-one	—	0402 ⁶³⁾
0410	Cholest-4-ene-3 α , 7 α , 12 α -triol	—	0411 ¹⁰²⁾
0411	7 α , 12 α -Dihydroxycholest-4-en-3-one	—	0408 ^{27, 102)}
0412	5 β -Cholestane-3 α , 7 α , 24-triol	Human ¹⁷²⁾	0414 ⁴⁷⁾
0413	5 α -Cholestane-3 α , 7 α , 25-triol	—	1113 ⁸⁷⁾
0414	5 β -Cholestane-3 α , 7 α , 25-triol	Human ¹⁷²⁾	HoCDCA ^{40, 116)}
0415	5 β -Cholestane-3 β , 7 α , 25-triol	—	0414 ⁵³⁾
0416	7 α , 25-Dihydroxy-5 β -cholestane-3-one	—	0414 ¹¹⁶⁾
0417	5 α -Cholestane-3 α , 7 α , 26-triol (3-Epi-16-deoxymyxinol)	Hagfish ²³⁹⁾	DECY ²³⁹⁾
0418	5 β -Cholestane-3 α , 7 α , 26-triol	Human ¹⁷²⁾	0414 ⁴⁷⁾
0419	5 α -Cholestane-3 β , 7 α , 26-triol (16-Deoxymyxinol)	Hagfish ^{12, 239)}	Kryptogenin ¹⁹⁶⁾
0420	Cholest-5-ene-3 β , 7 α , 26-triol	HSDD ¹⁰⁸⁾	—
0421	7 α , 25-Dihydroxycholest-4-en-3-one	—	0416 ¹¹⁶⁾

For abbreviations, see Table 1.

**Fig. 1.** Cholestanone skeleton.

Bile alcohols with a cholestanone skeleton are numbered as in Fig. 1. If one of the methyl groups attached to C-25 is substituted, it is

assigned the lower number, C-26, as recommended by IUPAC-IUB Joint Commission on Biochemical Nomenclature¹⁹¹⁾. Hence, what is termed, mistakenly, as "5 β -cholestane-3 α , 7 α , 12 α , 27-tetrol" should be correctly called (25R)-5 β -cholestane-3 α , 7 α , 12 α , 26-tetrol (III).

The occurrence of bile alcohols in nature was first demonstrated in 1898 by Hammarsten who found that the bile of the northern shark *Scymnus borealis* contains the sulfate ester of a neutral steroid named scymnol as its major constituent⁶⁵⁾. The structure of scymnol eluded investigators for many years, and was verified by partial synthesis in 1962³²⁾. The configuration of 24-hydroxyl group was finally determined in 1991 by means of X-ray diffraction analyses¹¹²⁾. Scymnol sulfate has been found in all elasmobranchii (sharks and rays) as their major bile salt, but has not been located in other natural sources⁷⁵⁾.

Table 5. C₂₆ Bile alcohols

No.	Systematic name (Trivial name)	Natural source	Synthetic source
0501	24-Nor-5β-cholestane-3α, 7α, 12α, 23, 25-pentol	—	0509 ⁴⁵⁾
0502	24-Nor-5β-cholestane-3α, 7α, 12α, 23-tetrol	—	0509 ⁴⁹⁾
0503	24-Nor-5β-cholestane-3α, 7α, 12α, 25-tetrol	Rabbit ¹⁹³⁾ ; Human ¹⁷²⁾ ; CHO ¹³³⁾	CA ⁴⁵⁾
0504	24-Nor-5β-cholestane-3α, 7α, 12α, 26-tetrol	—	0510 ⁴⁹⁾
0505	(22R)-24-Nor-5β-cholestane-3α, 7α, 22, 25-tetrol	—	22-OH-CDCA ⁹⁹⁾
0506	24-Nor-5β-cholestane-3α, 7α, 23, 25-tetrol	—	0513 ¹⁶³⁾
0507	24-Nor-5β-cholestane-3α, 7α, 25, 26-tetrol	—	0514 ¹⁶³⁾
0508	24-Nor-5β-cholestane-3α, 7α, 12α-triol	—	0503 ^{45, 49)}
0509	24-Nor-5β-cholest-23-ene-3α, 7α, 12α-triol	—	0503 ^{45, 49)}
0510	24-Nor-5β-cholest-25-ene-3α, 7α, 12α-triol	—	0503 ^{45, 49)}
0511	24-Nor-5β-cholestane-3α, 7α, 25-triol	—	CDCA ¹⁶³⁾
0512	24-Nor-5β-cholestane-3α, 7β, 25-triol	—	UDCA ¹⁸²⁾
0513	24-Nor-5β-cholest-23-ene-3α, 7α-diol	—	0511 ¹⁶³⁾
0514	24-Nor-5β-cholest-25-ene-3α, 7α-diol	—	0511 ¹⁶³⁾
0515	27-Nor-5β-cholestane-3α, 7α, 12α, 24, 25, 26-hexol	Human ¹⁷²⁾ , -S ⁸⁴⁾ , -U ^{82, 83)} ; CHO-U ¹²⁰⁾ , LD-U ⁸³⁾	0523 ¹⁷²⁾
0516	27-Norcholestane-3α, 7α, 12α, 24, 25-pentol	Rat ⁸⁶⁾ ; Human ¹⁷²⁾ , -S ^{84, 85)} , -U ^{82, 83, 84, 119, 177)} ; CTX-U ²²⁶⁾ ; ZS ⁵⁷⁾ ; CHO-U ¹⁰⁷⁾ ; LD-U ^{83, 177)} ; HS-U, F ⁵⁵⁾	CA ⁵⁵⁾ ; 0529 ¹⁷⁴⁾
0517	27-Norcholest-5-ene-3β, 7α, 12α, 24, 25-pentol	HSDD ¹⁰⁸⁾	—
0518	27-Nor-5α-cholestane-3α, 7α, 12α, 24, 26-pentol (5α-Ranol)	Frogs ^{14, 69, 72, 197, 240)}	—
0519	(24R)-27-Nor-5β-cholestane-3α, 7α, 12α, 24, 26-pentol (5β-Ranol)	Frogs ^{14, 132, 161, 197, 240)}	1101 ^{132, 167)} , 0523 ¹⁴⁵⁾
0520	(24S)-27-Nor-5β-cholestane-3α, 7α, 12α, 24, 26-pentol (24-Epi-5β-ranol)	—	1101 ¹⁶⁷⁾
0521	(24R)- and (24S)-27-Nor-5α-cholestane-3α, 7α, 12α, 24-tetrols (24-Epi-26-deoxy-5α-ranol and 26-Deoxy-5α-ranol)	Frogs ^{14, 197, 240)}	0525 ¹⁹⁷⁾
0522	(24R)- and (24S)-27-Nor-5β-cholestane-3α, 7α, 12α, 24-tetrols (24-Epi-26-deoxy-5β-ranol and 26-Deoxy-5β-ranol)	Frogs ^{14, 197, 240)}	0526 ¹⁹⁷⁾ , 0523 ¹⁴⁵⁾
0523	(24R)- and (24S)-27-Nor-5β-cholest-25-ene-3α, 7α, 12α, 24-tetrols	Human ¹⁷²⁾	CAld ^{145, 172)}
0524	27-Nor-5β-cholestane-3α, 7α, 12α, 26-tetrol	—	1102 ¹³⁰⁾
0525	3α, 7α, 12α-Trihydroxy-27-nor-5α-cholestan-24-one	Bullfrog ¹⁹⁸⁾	ACA ¹⁹⁷⁾
0526	3α, 7α, 12α-Trihydroxy-27-nor-5β-cholestan-24-one	Bullfrog ¹⁹⁸⁾ ; Human ¹⁷²⁾ ; THD ³⁹⁾ ; βOD ³⁹⁾	CA ¹⁹⁷⁾
0527	3α, 7α, 12α-Trihydroxy-27-nor-5β-cholest-25-one	—	HoCA ²⁰¹⁾
0528	27-Nor-5β-cholestane-3α, 7α, 12α-triol	—	0526 ¹²⁶⁾
0529	27-Nor-5β-cholest-24-ene-3α, 7α, 12α-triol	—	CAld ¹⁷⁴⁾

For abbreviations, see Table 1.

Table 6. C₂₅, C₂₄, C₂₃, and C₂₂ Bile alcohols

No.	Systematic name (Trivial name)	Natural source	Synthetic source
0601	26, 27-Dinor-5 β -cholestane-3 α , 7 α , 12 α , 24, 25-pentol	CHO ¹³³⁾	TADNC ²⁵⁴⁾
0602	26, 27-Dinor-5 α -cholestane-3 α , 7 α , 12 α , 24-tetrol	Frogs ¹⁴⁾	—
0603	26, 27-Dinor-5 β -cholestane-3 α , 7 α , 12 α , 24-tetrol	Frogs ¹⁴⁾	0604 ¹³⁴⁾
0604	3 α , 7 α , 12 α -Trihydroxy-26, 27-dinor-5 β -cholestane-24-one	CHO ¹³³⁾	CA ¹²⁵⁾
0605	26, 27-Dinor-5 β -cholestane-3 α , 7 α , 12 α , 25-tetrol	—	HoCA ¹³⁰⁾
0606	26, 27-Dinor-5 β -cholestane-3 α , 7 α , 24, 25-tetrol	—	CDCA ⁸⁸⁾
0607	26, 27-Dinor-5 β -cholestane-3 α , 7 α , 12 α -triol	—	0604 ¹²⁵⁾
0608	26, 27-Dinor-5 β -cholestane-3 α , 24, 25-triol	—	LCA ¹⁸⁹⁾
0609	3 α , 6 α , 7 α , 12 α -Tetrahydroxy-5 β -cholan-24-al	—	6-OH-7-ODCA ²⁴⁸⁾
0610	5 α -Cholane-3 α , 7 α , 12 α , 24-tetrol (5 α -Petromyzonol)	Lamprey ⁷⁷⁾ ; Lungfishes ⁶⁾	ACA ⁷⁷⁾
0611	5 β -Cholane-3 α , 7 α , 12 α , 24-tetrol (5 β -Petromyzonol)	Lungfish ⁶⁾ ; Human ¹⁷²⁾	CA ^{150, 166, 250)}
0612	3 α , 7 α , 12 α -Trihydroxy-5 α -cholan-24-al	—	DNCT ¹⁹⁸⁾
0613	3 α , 7 α , 12 α -Trihydroxy-5 β -cholan-24-al (Cholyl aldehyde)	—	0601 ²⁵⁴⁾ ; CA ^{63, 241)}
0614	(23R)-5 β -Cholane-3 α , 7 α , 23, 24-tetrol	Human ¹⁷²⁾	23-OH-CDCA ¹⁷²⁾
0615	5 β -Cholane-3 α , 6 α , 24-triol	—	HDCA ¹⁵⁰⁾
0616	5 β -Cholane-3 α , 7 α , 12 α -triol	—	CA ¹²⁵⁾
0617	5 β -Cholane-3 α , 7 α , 24-triol	—	CDCA ¹⁴⁸⁾
0618	5 β -Cholane-3 α , 7 β , 24-triol	—	UDCA ¹⁴⁸⁾
0619	5 β -Cholane-3 α , 12 α , 24-triol	—	DCA ^{150, 250)}
0620	3 α , 7 α -Dihydroxy-5 β -cholan-24-al (Chenodeoxycholyl aldehyde)	—	0606 ⁸⁸⁾
0621	5 β -Cholane-3 α , 24-diol	—	LCA ^{150, 250)}
0622	3 α -Hydroxy-5 β -cholan-24-al (Lithocholyl aldehyde)	—	0608 ¹⁸⁹⁾
0623	24-Nor-5 β -cholane-3 α , 7 α , 12 α , 22, 23-pentol	—	0634 ¹³⁵⁾
0624	3 α , 7 α , 12 α -Trihydroxy-24-nor-5 β -cholan-23-al (Norcholyl aldehyde)	—	NCA ¹⁴¹⁾ ; CA ²²³⁾
0625	24-Nor-5 β -cholane-3 α , 7 α , 12 α , 23-tetrol	—	NCA ¹⁵⁰⁾
0626	24-Nor-5 β -cholane-3 α , 7 α , 22, 23-tetrol	—	0636 ¹³⁹⁾
0627	24-Nor-5 β -cholane-3 α , 7 β , 22, 23-tetrol	—	0637 ¹⁴³⁾
0628	24-Nor-5 β -cholane-3 α , 12 α , 22, 23-tetrol	—	0638 ¹⁴³⁾
0629	24-Nor-5 β -cholane-3 α , 6 α , 23-triol	—	NHDCA ¹⁵⁰⁾
0630	24-Nor-5 β -cholane-3 α , 7 α , 12 α -triol	—	NCA ¹⁵³⁾
0631	24-Nor-5 β -cholane-3 α , 7 α , 23-triol	—	NCDCA ¹⁵⁰⁾
0632	24-Nor-5 β -cholane-3 α , 7 β , 23-triol	—	NUDCA ¹⁵⁰⁾
0633	24-Nor-5 β -cholane-3 α , 12 α , 23-triol	—	NDCA ¹⁵⁰⁾
0634	24-Nor-5 β -chol-22-ene-3 α , 7 α , 12 α -triol	—	CA ^{36, 135)}
0635	24-Nor-5 β -chol-22-ene-3 α , 6 α -diol	—	HDCA ³⁶⁾
0636	24-Nor-5 β -chol-22-ene-3 α , 7 α -diol	—	CDCA ^{36, 139)}
0637	24-Nor-5 β -chol-22-ene-3 α , 7 β -diol	—	UDCA ³⁶⁾
0638	24-Nor-5 β -chol-22-ene-3 α , 12 α -diol	—	DCA ³⁶⁾
0639	3 α -Hydroxy-24-nor-5 β -cholan-23-al	—	LCA ^{212, 253)}
0640	3 α , 7 α , 12 α -Trihydroxy-23, 24-dinor-5 β -cholan-22-al (Bisnorcholyl aldehyde)	—	0623 ¹³⁵⁾
0641	3 α , 7 α -Dihydroxy-23, 24-dinor-5 β -cholan-22-al (Bisnorchenodeoxycholyl aldehyde)	—	0626 ¹³⁹⁾
0642	3 α , 7 β -Dihydroxy-23, 24-dinor-5 β -cholan-22-al (Bisnorursodeoxycholyl aldehyde)	—	0627 ¹⁴³⁾
0643	3 α , 12 α -Dihydroxy-23, 24-dinor-5 β -cholan-22-al (Bisnordeoxycholyl aldehyde)	—	0628 ¹⁴³⁾
0644	3 α -Hydroxy-23, 24-dinor-5 β -cholan-22-al	—	LCA ²⁵³⁾

For abbreviations, see Table 1.

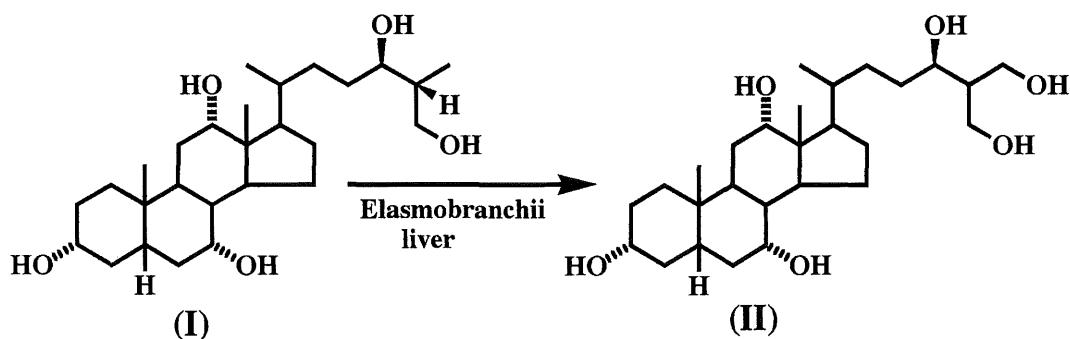


Fig. 2. Postulated structural relationship between 5β -chimaerol (I) and scymnol (II).

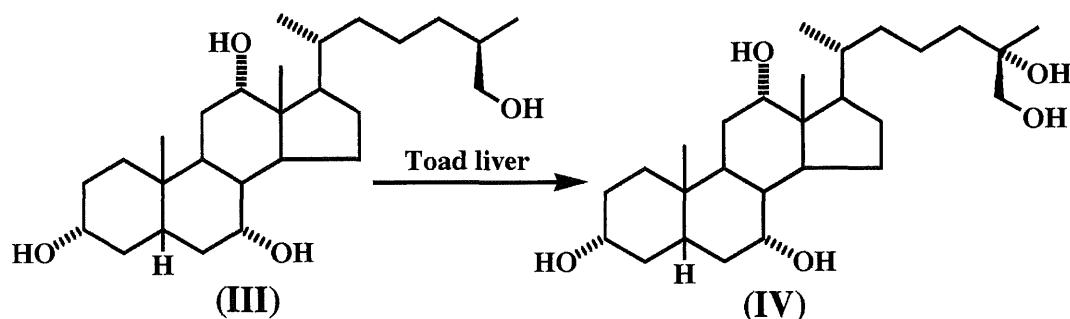


Fig. 3. Postulated structural relationship between (25R)- 5β -cholestane-3 α ,7 α ,12 α ,26-tetrol (III) and 5β -bufol (IV).

5β -Chimaerol sulfate is the chief bile salt of *Chimaera monstrosa*³³⁾ and a minor companion of scymnol sulfate in the bile of elasmobranchii^{112, 202)}. 5β -Chimaerol or its isomer with respect to C-24 and/or C-25 position has also been found in human bile¹⁷²⁾ and urine^{82, 83)}. The stereochemistry of the side chain of 5β -chimaerol remains unresolved, but the configuration at C-25 has been suggested as S on the basis of the data from molecular rotation contribution studies³³⁾. The structural relationship between scymnol and 5β -chimaerol (the latter is the 27-deoxy derivative of the former) as well as the co-existence of both bile alcohols in elasmobranchii bile and probably also in *Chimaera monstrosa* bile⁷⁴⁾ suggests that 5β -chimaerol (I) is the biosynthetic precursor of scymnol (II); hence the configuration at C-24 of 5β -chimaerol (I) should be the same; namely R, as that at C-24 of scymnol (II) (Fig. 2).

A principal bile alcohol of the white sucker *Catostomus commersoni* has the structure 5α -cholestane-3 α , 7 α , 12 α , 24, 26-pentol, whose configuration at C-24 and C-25 was assumed to be the same as those of 5β -chimaerol, and named 5α -chimaerol¹³⁾.

A number of 26-hydroxylated and 25, 26-dihydroxylated cholestan derivatives have been found not only in primitive vertebrates but also in mammals including humans. However, the configuration at C-25 of most of these bile alcohols is still undetermined. In biosynthetic experiments using radioactive mevalonate of 5β -bufol in the toad *Bufo vulgaris formosus*, it was shown that the terminal hydroxyl-bearing carbon atom (C-26) of 5β -bufol is derived from C-3' of mevalonate¹⁶²⁾. This indicates that in the biosynthesis of 5β -bufol the hydroxylation at the end methyl group of the cholesterol side chain is stereospecific, and the configuration at C-25 of 5β -cholestane-3 α , 7 α , 12 α , 26-tetrol, the direct precursor of 5β -bufol, is R. (25R)- 5β -Cholestan-3 α , 7 α , 12 α , 26-tetrol (III) has been recognized as an intermediate in the biosynthetic sequence between cholesterol and cholic acid in mammals²⁸⁾. Thus, until the formation of (25R)- 5β -cholestane-3 α , 7 α , 12 α , 26-tetrol (III), the pathway for the synthesis of 5β -bufol (IV) in the toad is the same as that for the synthesis of cholic acid in mammals. If 25-hydroxylation of (25R)- 5β -cholestane-3 α , 7 α , 12 α , 26-tetrol (III) takes place by the direct dis-

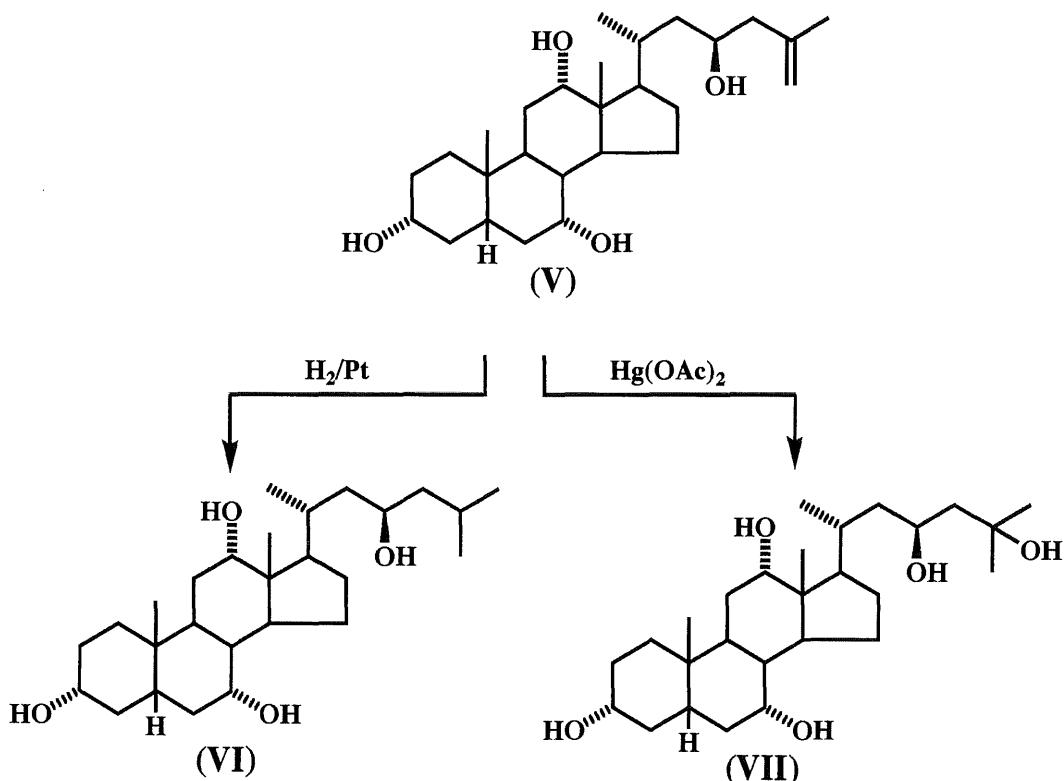


Fig. 4. Structural relationship of (23S)-5 β -cholest-25-ene-3 α ,7 α ,12 α ,23-tetrol (V), and (23R)-5 β -cholestane-3 α , 7 α , 12 α , 23-tetrol (VI) and (23S)-5 β -cholestane-3 α ,7 α ,12 α ,23,25-pentol (VII).

placement mechanism with retention of the configuration, as with hydroxylation of steroids by mixed function oxidases, the configuration at C-25 of 5 β -bufol (IV) could be assigned as S (Fig. 3).

Cerebrotendinous xanthomatosis (CTX) is a rare inherited disease caused by an inborn error of cholesterol catabolism and is characterized by accumulation of bile alcohols²²¹. The absolute configuration at C-23 of 5 β -cholestane-3 α , 7 α , 12 α , 23-tetrol (VI), one of the bile alcohols isolated from the bile, urine, and feces of patients with CTX, was determined as R by X-ray crystallography¹⁴⁰. The 23S-epimer has not yet been detected in nature.

One of two stereoisomers at C-23 of 5 β -cholestane-3 α , 7 α , 12 α , 23, 25-pentol is a chief bile alcohol of the urine from CTX patients^{226, 251}, and a second or a third bile alcohol of the bile and feces^{104, 225, 226}. The configuration at C-23 of the 23, 25-pentol (VII) was tentatively assigned as S [the same as that at C-23 of the naturally occurring (23R)-5 β -cholestane-3 α , 7 α , 12 α , 23-tetrol (VI)] on the basis of optical rotation differences¹⁰³. However, the opposite configuration at C-23 of the 23, 25-pentol (VII) was assigned by a circular dichroism study⁴⁶. Defi-

nite assignment of the 23S configuration of 5 β -cholestane-3 α ,7 α ,12 α ,23,25-pentol (VII) was made by the conversion of a key intermediate, (23S)-5 β -cholest-25-ene-3 α ,7 α ,12 α ,23-tetrol (V), to either the 23,25-pentol (VII) or the bile alcohol of known absolute configuration, (23R)-5 β -cholestane-3 α ,7 α ,12 α ,23-tetrol (VI) (Fig. 4)¹⁴¹. (23R)-5 β -Cholestane-3 α ,7 α ,12 α ,23,25-pentol was detected only in the urine of a CTX patient in a trace amount²²⁶.

Recently, a bile alcohol isolated from the urine of CTX patients was identified as 5 α -cholestane-3 α ,7 α ,12 α ,23,25-pentol¹⁴⁹. The configuration at C-23 of the 5 α -bile alcohol was deduced as S by the comparison of its ^{13}C -nuclear magnetic resonance spectrum with those of (23R)- and (23S)-5 β -cholestane-3 α ,7 α ,12 α ,23,25-pentols.

5 β -Ranol, a principal bile alcohol of the bullfrog, *Rana catesbeiana*, was identified as (24R)-27-nor-5 β -cholestane-3 α ,7 α ,12 α ,24,26-pentol (XI)^{145, 167}. However, the 24S-epimer (XII) of 5 β -ranol (XI) could not be detected in the bullfrog bile, though both stereoisomers at C-24 of 27-nor-5 β -cholestane-3 α ,7 α ,12 α ,24-tetrol, 26-deoxy-5 β -ranol (IX) and 24-epi-26-deoxy-5 β -ranol (X) were found in this bile¹⁹⁷. The bile alcohol pattern of the bullfrog bile is consistent with the results from the

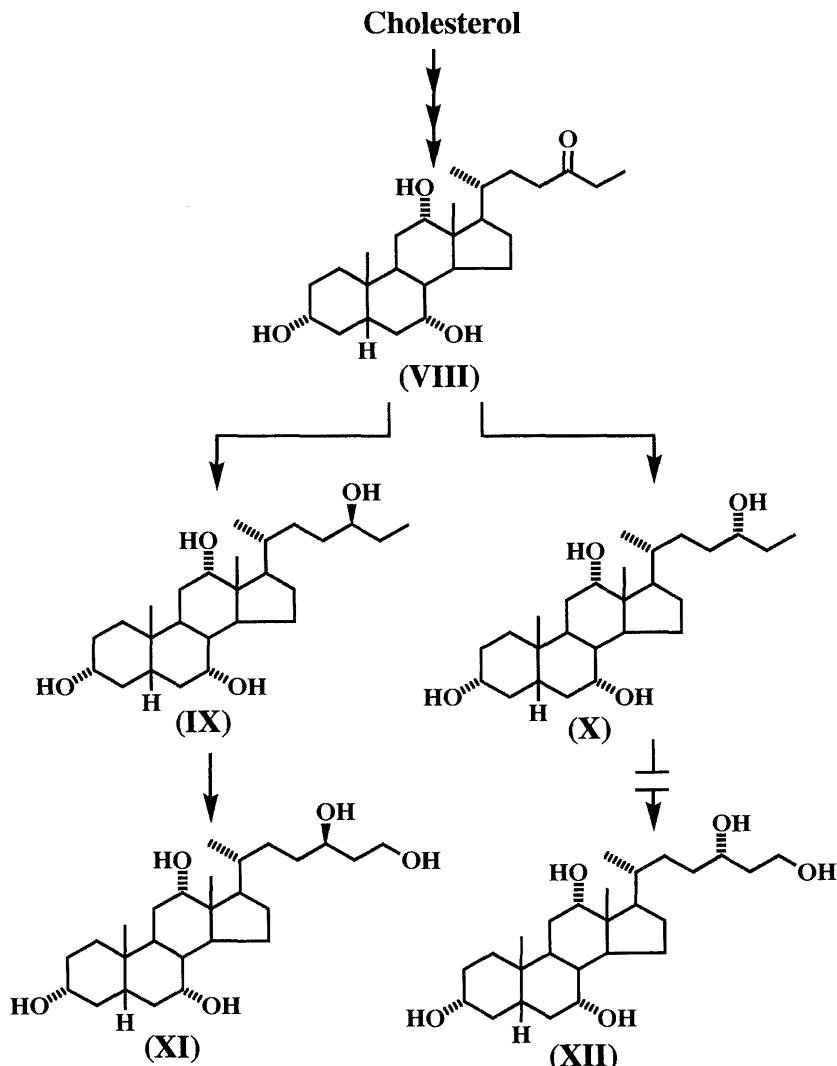


Fig. 5. Biosynthetic route of C₂₆ bile alcohols in bullfrog. VIII, 3 α ,7 α ,12 α -trihydroxy-27-nor-5 β -cholestan-24-one; IX, 26-deoxy-5 β -ranol [(24S)-27-nor-5 β -cholestane-3 α ,7 α ,12 α ,24-tetrol]; X, 24-epi-26-deoxy-5 β -ranol [(24R)-27-nor-5 β -cholestane-3 α ,7 α ,12 α ,24-tetrol]; XI, 5 β -ranol [(24R)-27-nor-5 β -cholestane-3 α ,7 α ,12 α ,24,26-pentol]; XII, 24-epi-5 β -ranol [(24S)-27-nor-5 β -cholestane-3 α ,7 α ,12 α ,24,26-pentol].

biosynthetic experiments of 5 β -ranol (XI), which showed that in the bullfrog 3 α , 7 α , 12 α -trihydroxy-27-nor-5 β -cholestan-24-one (VIII) formed from cholesterol is converted into both 26-deoxy-5 β -ranol (IX) and 24-epi-26-deoxy-5 β -ranol (X), and only 26-deoxy-5 β -ranol (IX) can be hydroxylated at C-26 to give 5 β -ranol (XI), but not 24-epi-26-deoxy-5 β -ranol (X) to 24-epi-5 β -ranol (XII) (Fig. 5)¹⁹⁹.

Higher Bile Acids

Higher bile acids are here defined as higher homologs with an extended side chain of C₂₄ bile

acids.

Tables 7–11 list all the naturally occurring higher bile acids of established structure and most chemically derived higher bile acids. Steroidal acids found in natural sources other than vertebrates, e.g. plants and invertebrates, chemically synthesized bile acids carrying no hydroxyl group, and esterified derivatives, are not included in the tables.

Higher bile acids are usually designated their systematic names, because they have no trivial names except with arapaimic acid and varanic acid.

Table 7. C₂₉ and C₂₈ Bile acids

No.	Systematic name	Natural source	Synthetic source
0701	3 α , 7 α , 12 α , 24-Tetrahydroxy-24-ethyl-5 β -cholestan-26-oic acid	—	0526 ¹⁸⁸⁾
0702	3 α , 7 α , 12 α -Trihydroxy-24-ethyl-5 β -cholestan-26-oic acid	—	0701 ¹⁸⁸⁾
0703	3 α , 7 α , 12 α -Trihydroxy-24-carboxymethyl-5 β -cholestan-26-oic acid	—	NCA ¹¹⁴⁾
0704	3 α , 7 α , 12 α -Trihydroxy-27-carboxymethyl-5 β -cholestan-26-oic acid	ZS ^{39, 59, 123, 205), -S^{38, 39, 59, 122, 123, 205, 220), -U^{39, 59, 205), RF-S^{38), COA^{205), -S^{114, 205), -U^{115, 205)}}}}}}}	CAld ²⁰⁶⁾
0705	3 α , 7 α , 12 α -Trihydroxy-24-ethyl-5 β -cholestane-26, 27-dioic acid	—	NCA ¹¹⁴⁾
0706	3 α , 7 α -Dihydroxy-24-ethyl-5 β -cholestan-26-oic acid	Sitosterol-fed monkey-F ¹⁵⁹⁾	—
0707	3 α -Hydroxy-24-ethyl-5 β -cholestan-26-oic acid	Sitosterol-fed monkey-F ¹⁵⁹⁾	—
0708	3 β -Hydroxy-24-ethylcholest-5-en-26-oic acid	Sitosterol-fed monkey-F ¹⁵⁹⁾	—
0709	3 α , 7 α , 12 α , 24-Tetrahydroxy-24-methyl-5 β -cholestan-26-oic acid	Toad ²⁴²⁾	0604 ⁸⁹⁾
0710	3 α , 7 α , 12 α -Trihydroxy-24-methyl-5 β -cholestan-26-oic acid	Bullfrog ¹⁹⁸⁾	0711 ⁸⁹⁾
0711	3 α , 7 α , 12 α -Trihydroxy-24-methyl-5 β -cholest-23-en-26-oic acid	—	0709 ⁸⁹⁾
0712	3 α , 7 α , 12 α -Trihydroxy-5 β -cholestane-24-carboxylic acid	—	0313 ¹⁰⁰⁾
0713	3 α , 7 α , 12 α -Trihydroxy-5 α -cholest-22-ene-24-carboxylic acid	Toad ²⁴²⁾	—
0714	3 α , 7 α , 12 α -Trihydroxy-5 β -cholest-22-ene-24-carboxylic acid	Toad ^{228, 242)}	—
0715	(25R)-3 α , 7 α , 12 α -Trihydroxy-5 β -cholestane-26-carboxylic acid	—	CA ³¹⁾
0716	(25S)-3 α , 7 α , 12 α -Trihydroxy-5 β -cholestane-26-carboxylic acid	—	CA ³¹⁾

For abbreviations, see Table 1.

Table 8. Tetra-oxygenated C₂₇ bile acids

No.	Systematic name (Trivial name)	Natural source	Synthetic source
0801	1 β , 3 α , 7 α , 12 α -Tetrahydroxy-5 β -cholestan-26-oic acid	Alligator ¹⁴⁷⁾ ; ZS-U ^{220, 246)}	—
0802	2 β , 3 α , 7 α , 12 α -Tetrahydroxy-5 β -cholestan-26-oic acid (Arapaimic acid)	Arapaima gigas ⁷⁸⁾	—
0803	3 α , 6 α , 7 α , 12 α -Tetrahydroxy-5 β -cholestan-26-oic acid	ZS-S ¹²²⁾ , -U ^{246, 248)}	0804 ²⁴⁸⁾
0804	3 α , 6 α , 7 α , 12 α -Tetrahydroxy-5 β -cholest-24-en-26-oic acid	—	0609 ²⁴⁸⁾
0805	(22R)-3 α , 7 α , 12 α , 22-Tetrahydroxy-5 β -cholestan-26-oic acid	ZS-U ^{246, 247)}	—
0806	(22S, 25R)-3 α , 7 α , 12 α , 22-Tetrahydroxy-5 β -cholestan-26-oic acid	Turtles ^{4, 62, 79, 252)}	—
0807	(23R)-3 α , 7 α , 12 α , 23-Tetrahydroxy-5 β -cholestan-26-oic acid	ZS-U ^{246, 247)}	—
0808	3 α , 7 α , 12 α , 24-Tetrahydroxy-5 α -cholestan-26-oic acid	Bullfrog ¹⁹⁸⁾	0612 ¹⁹⁸⁾
0809	(24R, 25R)-, (24R, 25S)-, (24S, 25R)-, and (24S, 25S)-3 α , 7 α , 12 α , 24-Tetrahydroxy-5 β -cholestan-26-oic acids (Varanic acid)	Toad ²⁴²⁾ ; Frogs ^{161, 198, 243)} ; Alligator ¹⁴⁷⁾ , Lizards ^{1, 69)} ; Human ¹⁸²⁾ ; ZS ^{39, 205)} , -S ^{38, 39, 122, 123, 220)} , -U ^{68, 123, 175, 181, 220, 246)} ; COA ²⁰⁵⁾ ; THD ³⁹⁾ , -S ³⁹⁾ ; β OD ³⁹⁾ , -S ³⁹⁾	CAld ^{23, 109, 152, 241)}
0810	3 α , 7 α , 12 α , 25-Tetrahydroxy-5 β -cholestan-26-oic acid	ZS ³⁹⁾ , -S ^{38, 39)} , -U ^{175, 220)} ; RF-U ¹⁷⁵⁾ , IO-GC ³⁷⁾ ; THD ³⁹⁾ , -S ³⁹⁾ ; β OD ³⁹⁾ , -S ³⁹⁾	CA ⁴¹⁾
0811	3 α , 7 α , 12 α , 26-Tetrahydroxy-5 α -cholestan-27-oic acid	Bullfrog ¹⁹⁸⁾	0909 ¹⁹⁸⁾
0812	3 α , 7 α , 12 α , 26-Tetrahydroxy-5 β -cholestan-27-oic acid	Toad and frogs ^{198, 240, 257)} ; Alligator ¹⁴⁷⁾ , Human ¹⁸²⁾ ; ZS ³⁹⁾ , -S ^{38, 39)} , -U ^{175, 246)} ; RF-S ³⁸⁾ , -U ¹⁷⁵⁾ ; THD ³⁹⁾ , -S ³⁹⁾ ; β OD ³⁹⁾ , -S ³⁹⁾	0910 ²⁴⁰⁾
0813	3 α , 7 α , 12 α , 26-Tetrahydroxy-5 β -cholest-23-en-27-oic acid	Toad ²⁵⁷⁾	—

For abbreviations, see Table 1.

Table 9. Tri-oxygenated C₂₇ bile acids

No.	Systematic name	Natural source	Synthetic source
0901	3 α , 7 α , 12 α -Trihydroxy-5 α -cholestane-26-oic acid	Coelacanth ¹⁴²⁾ ; Toads and frogs ^{161, 198, 240, 242, 257)} ; Alligator ¹⁴⁷⁾ ; Iguana ²⁰⁴⁾ ; Turtle*	DECY ¹¹⁷⁾
0902	(25R)- and (25S)-3 α , 7 α , 12 α -Trihydroxy-5 β -cholestane-26-oic acids	Toads and frogs ^{14, 154, 161, 171, 178, 198, 240, 242, 243, 257)} ; Alligator ^{22, 25, 70, 147)} ; -F ²³⁸⁾ ; Crocodile ^{70, 184)} ; Lizard ¹⁾ ; Turtle*; Kite ¹⁶⁰⁾ ; Baboon ²²²⁾ ; Human ^{24, 35, 182)} ; ZS ^{39, 59, 123, 187, 205), -S^{38, 39, 59, 64, 122, 123, 187, 205, 220, 249), -U^{68, 123, 181, 205, 220, 246), -F^{205), -AF²³¹⁾; RF-S^{38, 209)}; THD³⁹⁾, -S³⁹⁾; NALD-AF²³¹⁾; IBDA^{58, 67)}, -S, -U, -F⁶⁷⁾; COA²⁰⁵⁾, -S, -U, -F²⁰⁵⁾; βO³⁹⁾, -S³⁹⁾; IO-GC³⁷⁾}}}}	NCA ¹⁵⁵⁾ ; CA ³⁴⁾ ; HoCA ^{21, 22)} ; 0908 ⁶³⁾ ; 0715 ³¹⁾ ; 0716 ³¹⁾
0903	3 β , 7 α , 12 α -Trihydroxy-5 α -cholestane-26-oic acid	Coelacanth ¹⁴²⁾	0911 ¹⁴²⁾
0904	3 β , 7 α , 12 α -Trihydroxy-5 β -cholestane-26-oic acid	Alligator-F ²³⁸⁾	—
0905	3 α , 7 β , 12 α -Trihydroxy-5 β -cholestane-26-oic acid	Alligator ¹⁴⁷⁾	0914 ¹⁴⁷⁾
0906	3 α , 7 α , 12 α -Trihydroxy-5 α -cholest-23-en-26-oic acid	Toads ^{242, 257)}	—
0907	(25R)-3 α , 7 α , 12 α -Trihydroxy-5 β -cholest-23-en-26-oic acid	Toads ^{80, 81, 242, 257)} ; Lizard ¹⁾ ; THD ³⁹⁾	—
0908	(24E)- and (24Z)-3 α , 7 α , 12 α -Trihydroxy-5 β -cholest-24-en-26-oic acids	Lizard ¹⁾ ; Human ¹⁸²⁾ ; THD ³⁹⁾ , -S ³⁹⁾	CAld ^{63, 110, 241)}
0909	3 α , 7 α , 12 α -Trihydroxy-5 α -cholestane-26, 27-dioic acid	—	CY ^{137, 198)}
0910	3 α , 7 α , 12 α -Trihydroxy-5 β -cholestane-26, 27-dioic acid	—	CA ^{136, 240)}
0911	7 α , 12 α -Dihydroxy-3-oxo-5 α -cholestane-26-oic acid	Alligator ²³⁷⁾	DECY ¹⁴²⁾
0912	7 α , 12 α -Dihydroxy-3-oxo-5 β -cholestane-26-oic acid	Alligator ^{147, 237)}	0902 ¹⁴⁷⁾
0913	7 β , 12 α -Dihydroxy-3-oxo-5 β -cholestane-26-oic acid	Alligator ¹⁴⁷⁾	—
0914	3 α , 12 α -Dihydroxy-7-oxo-5 β -cholestane-26-oic acid	Alligator ^{147, 237)}	0902 ^{63, 147)}
0915	3 α , 7 α -Dihydroxy-12-oxo-5 α -cholestane-26-oic acid	—	0901 ¹¹⁷⁾
0916	3 β , 7 α , 12 α -Trihydroxycholest-5-en-26-oic acid	HSDD ¹⁰⁸⁾	—
0917	3 α , 7 α , 24-Trihydroxy-5 β -cholestane-26-oic acid	Lizard ¹⁾ ; ZS-S ^{122, 123)} , -U ¹²³⁾	CDCald ²⁴⁵⁾
0918	3 α , 12 α , 22-Trihydroxy-5 β -cholestane-26-oic acid	Turtle ⁷⁹⁾	0806*

*. Our unpublished observation. For abbreviations, see in Table 1.

Table 10. Di- and mono-oxygenated C₂₇ bile acids

No.	Systematic name	Natural source	Synthetic source
1001	3 α , 7 α -Dihydroxy-5 α -cholestan-26-oic acid	—	0915 ¹¹⁷⁾
1002	3 α , 7 α -Dihydroxy-5 β -cholestan-26-oic acid	Alligator ^{56, 147), -F²³⁸⁾; Kite^{160); Human^{66), ZS^{58, 59, 123, 187), -S^{38, 59, 122, 123, 187, 205, 220, 249), -U^{59, 68, 123, 181, 205, 246), -F^{205), -AF^{231), NALD-AF^{231), RF-S^{38), COA^{205), -S, -U, -F²⁰⁵⁾}}}}}}}}}}}	CDCA ^{34, 56, 66, 138); HoCDCA^{21), 0902^{25); 1004⁸⁸⁾}}}
1003	3 α , 7 α -Dihydroxy-5 β -cholest-23-en-26-oic acid	Lizard ¹⁾	—
1004	3 α , 7 α -Dihydroxy-5 β -cholest-24-en-26-oic acid	Lizard ¹⁾	CDCA ^{88, 110)}
1005	3 α , 7 α -Dihydroxy-5 β -cholestane-26, 27-dioic acid	—	CDCA ²⁴⁵⁾
1006	3 β , 7 α -Dihydroxy-5 α -cholestan-26-oic acid	—	Kryptogenin ¹⁹²⁾
1007	7 α -Hydroxy-3-oxo-5 α -cholestan-26-oic acid	—	Kryptogenin ¹⁹²⁾
1008	3 β , 7 α -Dihydroxycholest-5-en-26-oic acid	Human-S ^{16, 17), HSDD^{108), LD-S^{18); IMA-S¹⁷⁾}}}	—
1009	3 α , 7 β -Dihydroxy-5 β -cholestan-26-oic acid	Alligator ^{147), -F²³⁸⁾}	1010 ¹⁴⁷⁾
1010	3 α , 7 β -Dihydroxy-5 β -cholest-24-en-26-oic acid	—	UDCA ¹⁴⁷⁾
1011	3 β , 7 β -Dihydroxy-5 β -cholestan-26-oic acid	Alligator-F ²³⁸⁾	—
1012	7 α -Hydroxy-3-oxocholest-4-en-26-oic acid	Human-S ^{16, 17), LD-S^{18); IMA-S^{17); SH-S¹⁹⁴⁾}}}	—
1013	3 α , 12 α -Dihydroxy-5 β -cholestan-26-oic acid	Alligator ^{147, 237), -F²³⁸⁾}	DCA ^{34, 56), 1014¹⁴⁷⁾}
1014	3 α , 12 α -Dihydroxy-5 β -cholest-24-en-26-oic acid	—	DCA ^{110, 147)}
1015	3 β , 12 α -Dihydroxy-5 β -cholestan-26-oic acid	Alligator-F ²³⁸⁾	—
1016	3 α , 7 α -Dihydroxy-24-methyl-27-nor-5 β -cholestan-26-oic acid	—	CDCA ¹³⁸⁾
1017	3 α -Hydroxy-5 β -cholestan-26-oic acid	Alligator-F ^{238); ZS-S^{205); COA-S^{114, 205)}}}	LCA ^{34); 1019¹⁸⁹⁾}
1018	3 β -Hydroxy-5 β -cholestan-26-oic acid	Alligator-F ²³⁸⁾	
1019	3 α -Hydroxy-5 β -cholest-24-en-26-oic acid	—	LCA ^{110, 189)}
1020	3 β -Hydroxycholest-5-en-26-oic acid	Human-S ^{16, 17), ZS^{205), -S^{205); IMA-S^{17); COA^{205), -S^{114, 205); LD-S¹⁸⁾}}}}}}	—

For abbreviations, see Table 1.

Table 11. C₂₆ and C₂₅ Bile acids

No.	Systematic name (Trivial name)	Natural source	Synthetic source
1101	(24R)- and (24S)-3 α , 7 α , 12 α , 24-Tetrahydroxy-27-nor-5 β -cholestan-26-oic acids	—	CAld ¹⁶⁷⁾
1102	3 α , 7 α , 12 α -Trihydroxy-27-nor-5 β -cholestan-26-oic acid	—	CA ^{31, 129)} ; NCA ¹²⁸⁾
1103	3 α , 7 α , 12 α -Trihydroxy-24-nor-5 β -cholestan-26-oic acid	—	CA ⁵⁴⁾
1104	3 α , 7 α , 12 α , 23-Tetrahydroxy-26, 27-dinor-5 β -cholestan-25-oic acid	—	NCAld ^{103, 146)}
1105	3 α , 6 β , 7 α -Trihydroxy-26, 27-dinor-5 β -cholestan-25-oic acid	—	1122 ²⁵⁶⁾
1106	3 α , 7 α , 12 α -Trihydroxy-26, 27-dinor-5 α -cholestan-25-oic acid (Allohomocholic acid)	Bullfrog*	ANCY ⁹⁰⁾
1107	3 α , 7 α , 12 α -Trihydroxy-26, 27-dinor-5 β -cholestan-25-oic acid (Homocholic acid)	Bullfrog*	CA ^{31, 126, 207)}
1108	3 β , 7 α , 12 α -Trihydroxy-26, 27-dinor-5 β -cholestan-25-oic acid	—	1111 ²¹³⁾
1109	3 α , 7 β , 12 α -Trihydroxy-26, 27-dinor-5 α -cholestan-25-oic acid	—	7-EACA ⁹⁾
1110	3 α , 7 β , 12 α -Trihydroxy-26, 27-dinor-5 β -cholestan-25-oic acid	—	1112 ¹⁶⁸⁾
1111	7 α , 12 α -Dihydroxy-3-oxo-26, 27-dinor-5 β -cholestan-25-oic acid	—	HoCA ²¹³⁾
1112	3 α , 12 α -Dihydroxy-7-oxo-26, 27-dinor-5 β -cholestan-25-oic acid	—	HoCA ¹⁶⁸⁾
1113	3 α , 7 α -Dihydroxy-26, 27-dinor-5 α -cholestan-25-oic acid	—	1117 ⁸⁷⁾
1114	3 α , 7 α -Dihydroxy-26, 27-dinor-5 β -cholestan-25-oic acid (Homochenodeoxycholic acid)	—	CDCA ⁴⁰⁾
1115	3 α , 7 β -Dihydroxy-26, 27-dinor-5 β -cholestan-25-oic acid (Homoursodeoxycholic acid)	—	UDCA ¹⁶⁸⁾
1116	3 α , 7 β -Dihydroxy-26, 27-dinor-5 β -cholest-11-en-25-oic acid	—	1110 ¹⁶⁸⁾
1117	7 α -Hydroxy-3-oxo-26, 27-dinor-5 α -cholestan-25-oic acid	—	HoCDCA ⁸⁷⁾
1118	7 α -Hydroxy-3-oxo-26, 27-dinor-5 β -cholestan-25-oic acid	—	HoCDCA ^{87, 116)}
1119	7 α -Hydroxy-3-oxo-26, 27-dinorchest-4-en-25-oic acid	—	1118 ¹¹⁶⁾
1120	3 α -Hydroxy-7-oxo-26, 27-dinor-5 β -cholestan-25-oic acid	—	HoCDCA ¹⁶⁹⁾
1121	3 α -Hydroxy-26, 27-dinor-5 β -cholestan-25-oic acid (Homolithocholic acid)	—	1120 ¹⁶⁹⁾
1122	3 α -Hydroxy-26, 27-dinor-5 β -cholest-6-en-25-oic acid	—	CDCA ²⁵⁶⁾

*. Our unpublished observation. For abbreviations, see Table 1.

The occurrence of higher bile acids in nature was first demonstrated in 1934 by Shimizu and Oda who isolated a major bile acid from the bile of the toad, *Bufo vulgaris formosus*, to which they assigned the formula C₂₈H₄₆O₅, and named it "trihydroxybufosterochenic acid"²²⁸⁾. Although

the nuclear structure (cholic acid type) and location of the double bond (between C-22 and C-23) of trihydroxybufosterochenic acid (XIII) were elucidated in 1936 from the fact that it gave bisnorcholic acid (XIV) upon ozonolysis (Fig. 6)²²⁹⁾, the structure of the terminal part of the side

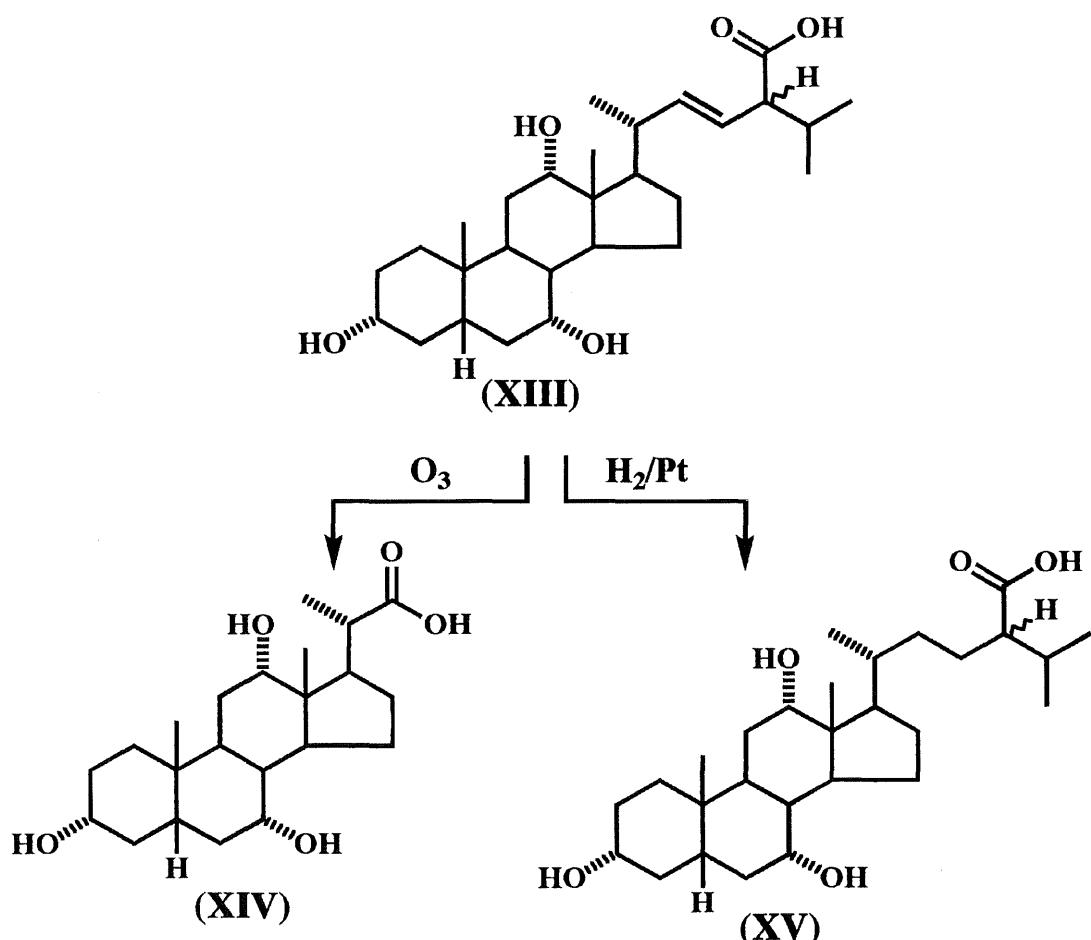


Fig. 6. Structural determination of "trihydroxybufosterochenolic acid". XIII, $3\alpha, 7\alpha, 12\alpha$ -trihydroxy- 5β -cholest-22-ene-24-carboxylic acid; XIV, bisnorcholic acid; XV, $3\alpha, 7\alpha, 12\alpha$ -trihydroxy- 5β -cholestane-24-carboxylic acid;

chain had not been determined at that time. The structure of trihydroxybufosterochenolic acid was finally described in 1967 by Hoshita et al who synthesized $3\alpha, 7\alpha, 12\alpha$ -trihydroxy- 5β -cholestane-24-carboxylic acid (XV), which was identical with the hydrogenated derivative of trihydroxybufosterochenolic acid (XIII); hence the major bile acid of the toad was characterized as $3\alpha, 7\alpha, 12\alpha$ -trihydroxy- 5β -cholest-22-ene-24-carboxylic acid (XIII) (Fig. 6)¹⁰⁰. The stereochemistry of the side chain is still not yet established.

The natural occurrence of $3\alpha, 7\alpha, 12\alpha$ -trihydroxy- 5β -cholestane-26-oic acid was first demonstrated in 1939 in the bile of the bullfrog, *Rana catesbeiana*¹⁷¹, and later in various species of amphibians^{14, 154, 161, 178, 240, 242, 243, 257}, all crocodilians examined⁷⁵, the kite, *Milvus lineatus lineatus*¹⁶⁰, and even in healthy and diseased humans^{35, 58}. This C₂₇ homolog of cholic acid has an asymmetric carbon atom at C-25; thus there are two stereoisomers. Most of the above-mentioned vertebrates have only (25R)- $3\alpha, 7\alpha, 12\alpha$ -trihydroxy- 5β -cholestane-26-oic acid (XVIII).

However, the bullfrog bile and the crocodile bile contain both 25R and 25S isomers^{178, 184}. The occurrence of both 25R and 25S isomers in the same species at once arouses the suspicion that one is an artifact formed from the other. This is especially possible in the case of the crocodile in which the bile acids occur as taurine conjugates. Alkaline conditions necessary to hydrolyze the taurine-conjugated trihydroxy- 5β -cholestanoates might cause racemization at C-25. Une et al clearly showed that in the bullfrog both (25R)- and (25S)- $3\alpha, 7\alpha, 12\alpha$ -trihydroxy- 5β -cholestane-26-oic acids are authentic metabolites in this species²⁴³. Since all bile acids of the bullfrog bile occur in unconjugated form, they could be obtained without use of drastic procedures such as alkaline hydrolysis. High-performance liquid chromatographic analysis of the bile acid mixture of the bullfrog revealed that both 25R and 25S isomers occur in a ratio of about 20:1.

Patients with the cerebrohepatorenal syndrome of Zellweger accumulate $3\alpha, 7\alpha, 12\alpha$ -trihydroxy- 5β -cholestane-26-oic acid and other higher

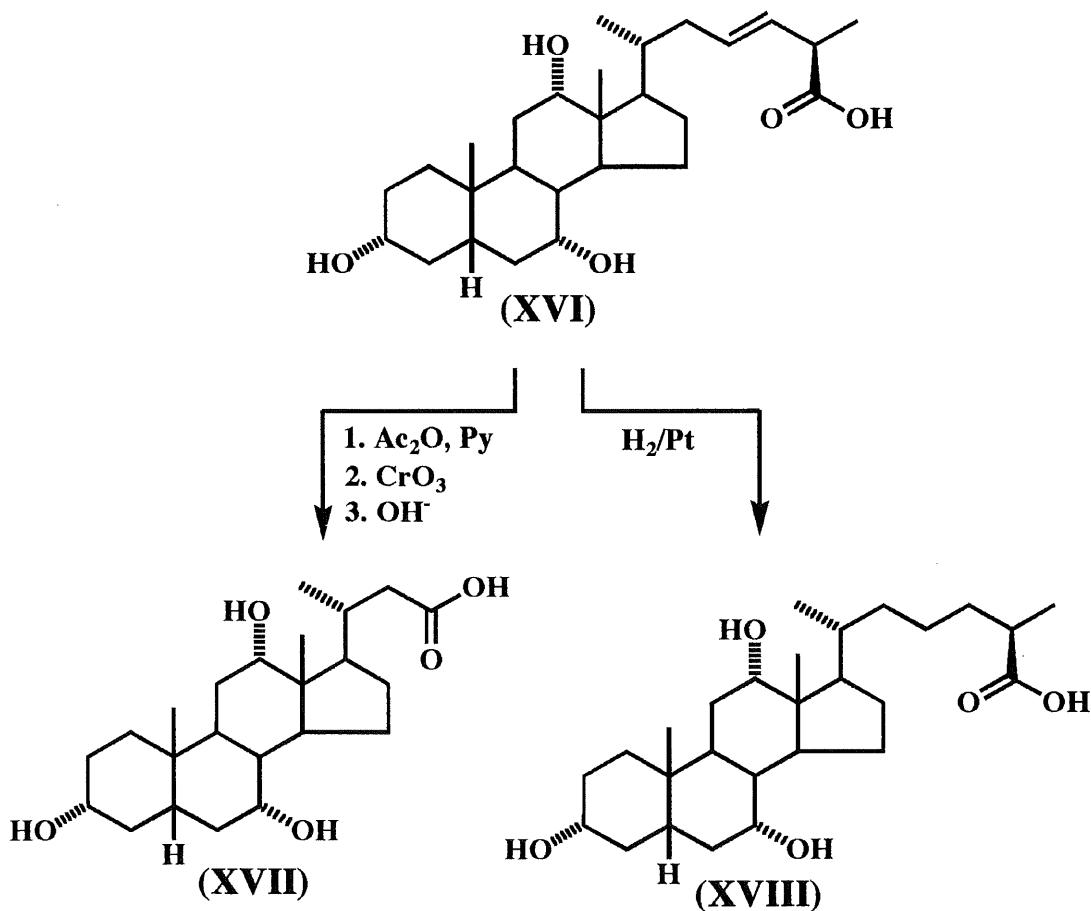


Fig. 7. Structural determination of the second major bile acid of *Bufo vulgaris formosus*. XVI, (25R)-3 α ,7 α ,12 α -trihydroxy-5 β -cholest-23-en-26-oic acid; XVII, norcholic acid; XVIII, (25R)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-oic acid.

bile acids in their bile, serum, and urine as unusual metabolites of cholesterol^{68, 123}. Une et al have reported that 3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-oic acid excreted into the urine from an infant with Zellweger syndrome as the unconjugated form consisted of a mixture of the 25R and 25S isomers in the ratio of about 7:3²⁴⁶. Human liver is thought to synthesize only the 25R isomer (XVIII) of 3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-oic acid as the biosynthetic precursor of cholic acid in normal conditions²⁸. The formation of (25S)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-oic acid in Zellweger syndrome may reflect the operation of an unusual pathway of cholesterol catabolism in this unusual condition.

Okuda et al isolated a higher bile acid from the bile of the iguana, *Iguana iguana*, as a minor companion of allocholic acid, the major bile acid of this animal²⁰⁴. The structure of the minor bile acid of the iguana was deduced as the C₂₇ homolog of allocholic acid, 3 α ,7 α ,12 α -trihydroxy-5 α -cholestan-26-oic acid by lithium aluminum hydride reduction to 5 α -cholestane-3 α ,7 α ,12 α ,26-tetrol²⁰⁴. The 5 α -C₂₇ bile acid was later pre-

pared from 5 α -cholestane-3 α ,7 α ,12 α ,26-tetrol¹¹⁷, and also found in the bile of some species of amphibians^{161,198,240,242}. The stereochemistry at C-25 of the naturally occurring 3 α ,7 α ,12 α -trihydroxy-5 α -cholestan-26-oic acid remained unknown. Kanemitsu isolated a minor bile acid from the bile of the turtle, *Amyda japonica*, and named "heterocholic acid"¹¹⁸. Our own observation has revealed that heterocholic acid is the 2:1 mixture of 3 α , 7 α , 12 α -trihydroxy-5 α - and 5 β -cholestan-26-oic acids.

A second major bile acid (XVI) from the bile of the toad, *Bufo vulgaris formosus*, was characterized as (25R)-3 α ,7 α ,12 α -trihydroxy-5 β -cholest-23-en-26-oic acid from the fact that the second major bile acid could be converted to norcholic acid (XVII) by oxidation and to (25R)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-oic acid (XVIII) by hydrogenation (Fig. 7)^{80,81}. This unsaturated C₂₇ bile acid also occurs in the bile of another species of the toad, *Bufo marinus*, as a major bile acid²⁵⁷, and in the bile of the monitor lizard, *Varanus monitor*, as a minor constituent¹.

In 1936, Yamasaki and Yuuki isolated a major

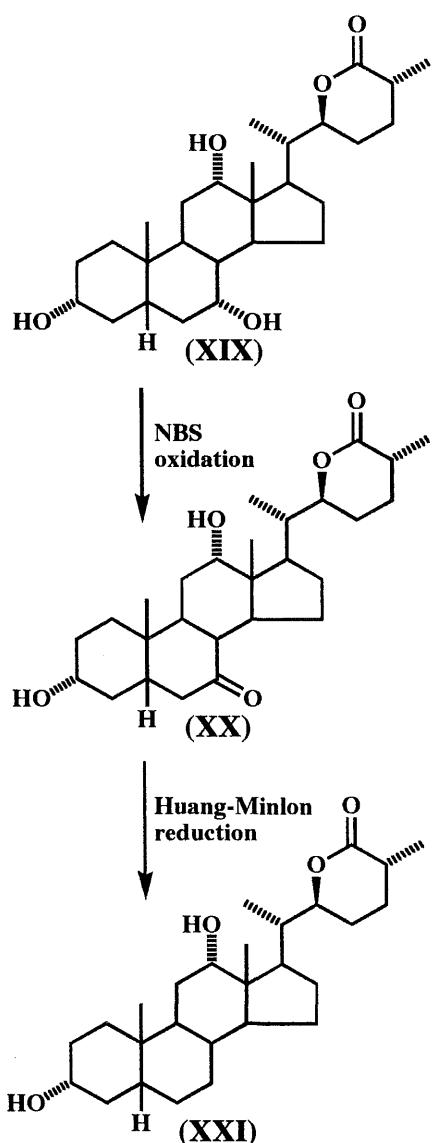


Fig. 8. Chemical conversion of the major bile acid of the turtle into a minor bile acid. XIX, (22S, 25R)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestano-26,22-lactone; XX, (22S,25R)-3 α ,12 α -dihydroxy-7-oxo-5 β -cholestano-26,22-lactone; XXI, (22S,25R)-3 α ,12 α -dihydroxy-5 β -cholestano-26,22-lactone. NBS, *N*-Bromo-succinimide.

bile acid of the turtle, *Amyda japonica*, as the lactone form, and named it "tetrahydroxysterolanic lactone"²⁵². The structure of tetrahydroxysterolanic lactone was characterized as (22S,25R)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestano-26,22-lactone (XIX) by ^1H -nuclear magnetic resonance spectrometry comparison with the reference compounds, four possible stereoisomers with respect to the C-22 and C-25 positions of 6 β -methyl-3 α ,5 α -cholestano-26,22-lactones⁶². Thus, the native higher bile acid of the turtle should be formulated as (22S,25R)-3 α ,7 α ,12 α ,22-tetrahydroxy-5 β -cholestano-26-oic acid,

which was found in all turtles and tortoises examined as their major biliary constituent but not in any other vertebrate, and is now recognized as the characteristic component of the bile of the chelonians⁷⁵.

3 α ,12 α ,22-Trihydroxy-5 β -cholestano-26-oic acid was found in the bile of the green turtle, *Chelonia mydas* along with (22S, 25R)-3 α ,7 α ,12 α ,22-tetrahydroxy-5 β -cholestano-26-oic acid⁷⁹. Our own observation has also revealed that 3 α ,12 α ,22-trihydroxy-5 β -cholestano-26-oic acid is a minor companion of (22S,25R)-3 α ,7 α ,12 α ,22-tetrahydroxy-5 β -cholestano-26-oic acid in the bile of the turtle, *Amyda japonica*. The stereochemistry of the side chain of 3 α ,12 α ,22-trihydroxy-5 β -cholestano-26-oic acid is believed to be the same as that of (22S,25R)-3 α ,7 α ,12 α ,22-tetrahydroxy-5 β -cholestano-26-oic acid because the lactone (XXI) of the former can be derived from the lactone (XIX) of the latter by the selective oxidation of 7 α -hydroxyl group followed by Huang-Minlon reduction of the resultant 7-oxo group to the methylene group (our unpublished observation) (Fig. 8).

Une et al have found two novel taurine-conjugated higher bile acids in urine from a patient with Zellweger syndrome^{246,247}. These higher bile acids were obtained as the lactone form after alkaline hydrolysis followed by the extraction with ether of the acidified hydrolysate. The two steroidal lactones (XXIV, XXV) were treated with lithium aluminum hydride and the resultant reduction products (XXVI, XXVII) were identified as (22R,25R)-5 β -cholestane-3 α ,7 α ,12 α ,22, 26-pentol and (23R)-5 β -cholestane-3 α ,7 α ,12 α ,23, 26-pentol by comparison with authentic compounds. These results indicate that the two native higher bile acids (XXII, XXIII) are (22R,25R)-3 α ,7 α ,12 α ,22- and (23R)-3 α ,7 α ,12 α ,23-tetrahydroxy-5 β -cholestano-26-oic acids, respectively (Fig. 9)²⁴⁷.

Varanic acid was isolated from the lizard, *Varanus niloticus*, from which the bile acid received its name, and characterized as a 3 α ,7 α ,12 α ,24-tetrahydroxy-5 β -cholestano-26-oic acid^{41,69,109}. Varanic acid or its diastereoisomers at C-24 and/or C-25 was found in the bile of several species of amphibians^{242,243} and in the biological fluids of healthy and diseased humans^{175,205}. Une et al synthesized all four stereoisomers at C-24 and C-25 of 3 α ,7 α ,12 α ,24-tetrahydroxy-5 β -cholestano-26-oic acid²⁴¹. Comparison with the synthetic varanic acids of known absolute configuration revealed that the varanic acid of the frog, *Bombina orientalis*, has the 24R, 25S configuration²⁴³. It is, however, still necessary to continue study of the stereochemistry of varanic acids; since Kinoshita et al also examined the stereochemistry of all four isomers at C-24 and C-25 of varanic acid and claimed the 24R, 25R configuration for the *Bombina* varanic acid¹⁵².

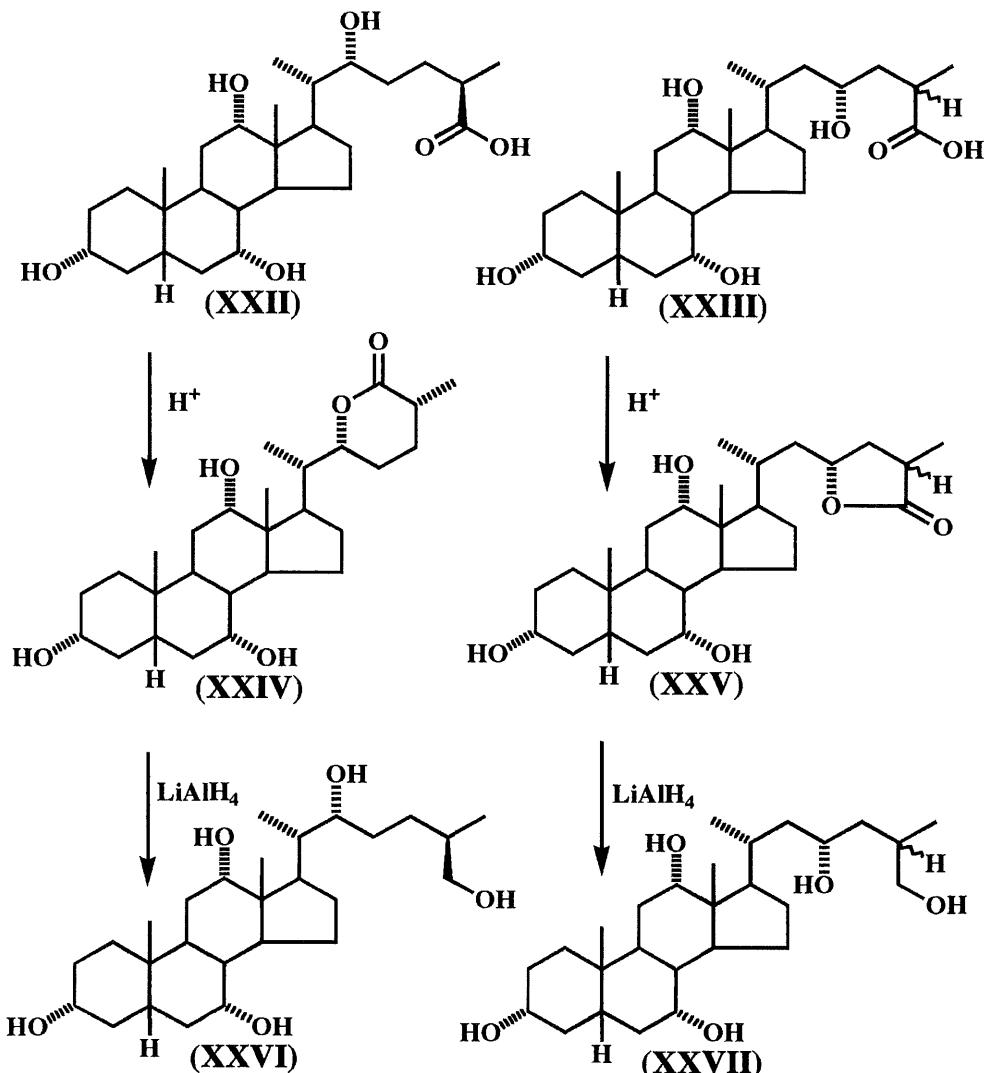


Fig. 9. Structural determination of two novel higher bile acids found in urine from patients with Zellweger syndrome. XXII, (22R,25R)-3 α ,7 α ,12 α ,22-tetrahydroxy-5 β -cholestane-26-oic acid; XXIII, (23R)-3 α ,7 α ,12 α , 23-tetrahydroxy-5 β -cholestane-26-oic acid; XXIV, (22R,25R)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestano-26, 22-lactone; XXV, (23R)-3 α , 7 α ,12 α -trihydroxy-5 β -cholestano-26,23-lactone; XXVI, (22R,25R)-5 β -cholestane-3 α ,7 α ,12 α ,22,26-pentol; XXVII, (23R)-5 β -cholestane-3 α ,7 α ,12 α ,23,26-pentol.

According to current concepts, the major pathway for the biosynthesis of cholic acid (XXXI) in mammals involves a 3 α ,7 α ,12 α -trihydroxy-5 β -cholest-24-en-26-oic acid and a 3 α ,7 α ,12 α ,24-tetrahydroxy-5 β -cholestane-26-oic acid as the intermediates^{28,244}. By direct comparison with the compounds of known absolute configuration, these intermediary higher bile acids were identified as (24E)-3 α ,7 α ,12 α -trihydroxy-5 β -cholest-24-en-26-oic acid (XXIX) and the 3 α ,7 α ,12 α ,24-tetrahydroxy-5 β -cholestane-26-oic acid assigned by us to have the 24R, 25S configuration (Fig. 10)²⁴⁴.

3 α ,7 α ,12 α -Trihydroxy-5 β -cholest-24-en-26-oic acid has been detected in the bile of the lizard, *Varanus monitor*, as its minor constituent¹⁾ and in the bile of healthy and diseased humans^{39,182}.

The stereochemistry of the Δ^{24} -double bond of the C₂₇ bile acid found in healthy human bile was determined as Z¹⁸², while a patient with thiolase deficiency contained both the 24E and 24Z isomers³⁹.

An unusual C₂₉-dicarboxylic bile acid, 3 α ,7 α ,12 α -trihydroxy-27-carboxymethyl-5 β -cholestane-26-oic acid (3 α ,7 α ,12 α -trihydroxy-27a,27b-dihydro-5 β -cholestane-26,27b-dioic acid) (XXXII) was found in the serum of patients with Zellweger syndrome (Fig. 11)²⁰⁵. The structure has recently been confirmed by partial synthesis²⁰⁶.

Short Side Chain Bile Acids

Lester et al have claimed that the term "bile acids" should be applied to the steroids with a

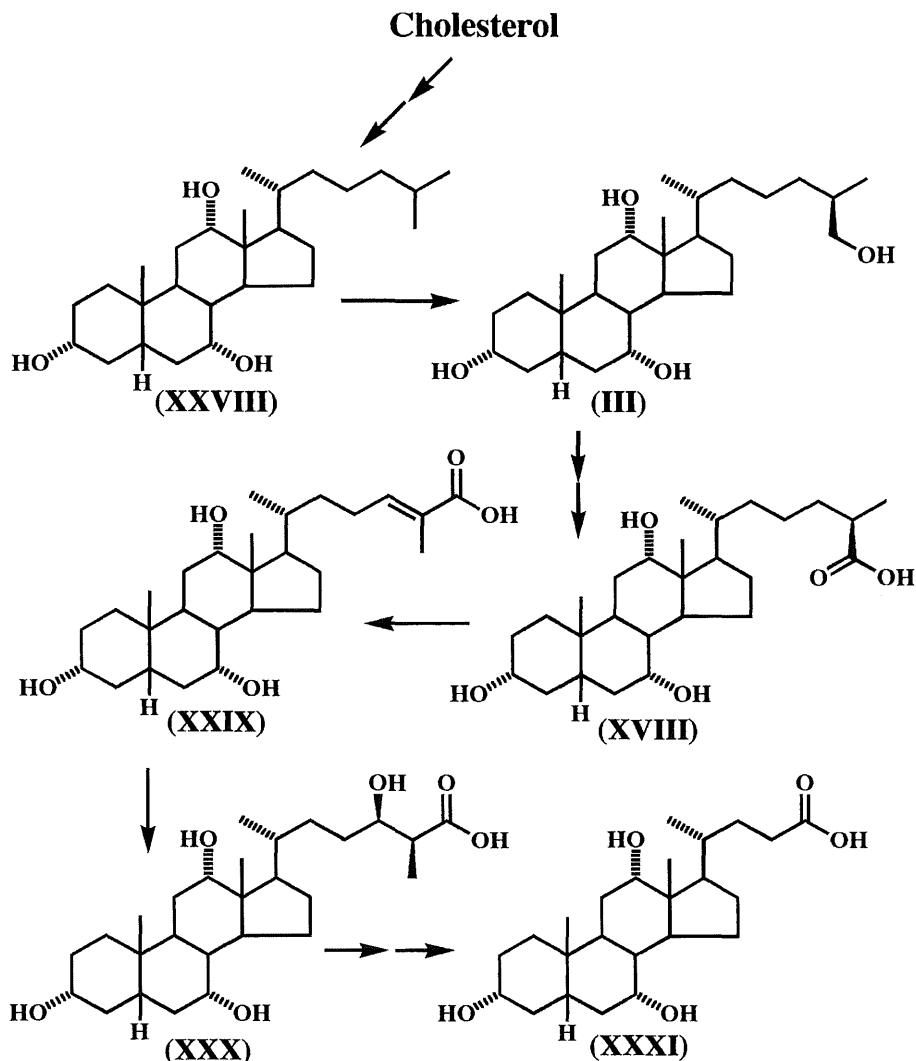


Fig. 10. Postulated pathway for the biosynthesis of cholic acid. XXVIII, 5 β -cholestane-3 α ,7 α ,12 α -triol; III, (25R)-5 β -cholestane-3 α ,7 α ,12 α ,26-tetrol; XVIII, (25R)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestane-26-oic acid; XXIX, (24E)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-24-en-26-oic acid; XXX, (24R,25S)-3 α ,7 α ,12 α ,24-tetrahydroxy-5 β -cholestane-26-oic acid; XXXI, cholic acid.

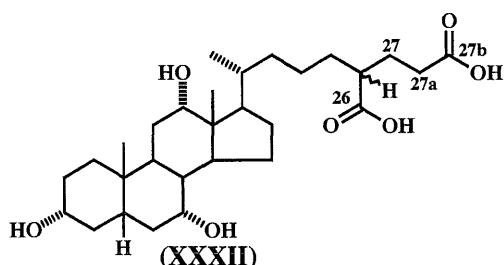


Fig. 11. 3 α ,7 α ,12 α -Trihydroxy-27-carboxymethyl-5 β -cholestane-26-oic acid (XXXII).

side chain at C-17 ending in a carboxylic acid group¹⁷⁶). However, etianic acid derivatives and pregnanoic acid derivatives are not termed “bile acids” in this review, since these C₂₀ and C₂₁ steroidal acids are biochemically related to steroid hormones rather than common C₂₄ bile acids. Thus, short side chain bile acids are here defined as C₂₃ and C₂₂ steroidal acids with one and two less carbon atoms in the side chain than common C₂₄ bile acids.

The prefixes “nor” and “dinor” are used for systematic names of C₂₃ and C₂₂ bile acids, respectively, while the prefix “bisnor” is used only for the trivial name of C₂₂ bile acids.

Tables 12 and 13 list most of the naturally occurring and chemically derived C₂₃ and C₂₂ bile acids with the shortened side chain. Specifically

Table 12. C₂₃ Bile acids

No.	Systematic name (Trivial name)	Natural source	Synthetic source
1201	1 β , 3 α , 7 α , 12 α -Tetrahydroxy-24-nor-5 β -cholan-23-oic acid	CTX-U ¹⁷⁰⁾	—
1202	2 β , 3 α , 7 α , 12 α -Tetrahydroxy-24-nor-5 β -cholan-23-oic acid	CTX-U ¹⁷⁰⁾	—
1203	3 α , 7 α , 12 α -Trihydroxy-24-nor-5 α -cholan-23-oic acid (Allonorholic acid)	Human ¹⁸²⁾ ; CTX ^{144, 182)} , -U ^{144, 170)}	1207 ²²⁴⁾
1204	3 α , 7 α , 12 α -Trihydroxy-24-nor-5 β -cholan-23-oic acid (Norcholic acid)	Human ¹⁸²⁾ , -U ²⁾ , -M ¹⁹⁾ , -AF ²³⁰⁾ , -UCB ²³⁰⁾ ; CTX ^{144, 182)} , -U ^{144, 156, 170)} ; LD-U ^{2, 7, 29, 230)} , -S ^{30, 230)} ; IMA-U ³⁾ ; CHO-U ²³⁶⁾	CA ^{190, 217, 223, 224, 229, 233)}
1205	3 β , 7 α , 12 α -Trihydroxy-24-nor-5 α -cholan-23-oic acid	—	1207 ²²⁴⁾
1206	3 β , 7 α , 12 α -Trihydroxy-24-nor-5 β -cholan-23-oic acid	—	1208 ²²⁴⁾
1207	7 α , 12 α -Dihydroxy-3-oxo-24-nor-5 α -cholan-23-oic acid	—	NCA ²²⁴⁾
1208	7 α , 12 α -Dihydroxy-3-oxo-24-nor-5 β -cholan-23-oic acid	—	NCA ^{195, 224)}
1209	7 α , 12 α -Dihydroxy-3-oxo-24-norchol-4-en-23-oic acid	—	NCA ²²⁴⁾
1210	3 α , 12 α -Dihydroxy-7-oxo-24-nor-5 β -cholan-23-oic acid	CTX ¹⁸²⁾ , -U ¹⁷⁰⁾	NCA ¹⁹⁵⁾
1211	3 α , 7 α -Dihydroxy-12-oxo-24-nor-5 β -cholan-23-oic acid	CTX-U ¹⁷⁰⁾	NCA ²³³⁾
1212	3 α -Hydroxy-7, 12-dioxo-24-nor-5 β -cholan-23-oic acid	—	NCA ²³³⁾
1213	3 α , 6 α -Dihydroxy-24-nor-5 β -cholan-23-oic acid (Norhyodeoxycholic acid)	—	HDCA ^{151, 186, 217)}
1214	3 α , 7 α -Dihydroxy-24-nor-5 β -cholan-23-oic acid (Norchenodeoxycholic acid)	Human ¹⁸²⁾	CDCA ^{71, 113, 217)}
1215	3 α , 7 β -Dihydroxy-24-nor-5 β -cholan-23-oic acid (Norursodeoxycholic acid)	Human ¹⁸²⁾ ; CTX-U ¹⁵⁶⁾	UDCA ^{156, 217);} 0512 ¹⁸²⁾
1216	3 α , 12 α -Dihydroxy-24-nor-5 β -cholan-23-oic acid (Nordeoxycholic acid)	—	DCA ^{217);} 1210 ¹⁹⁵⁾
1217	3 α -Hydroxy-12-oxo-24-nor-5 β -cholan-23-oic acid	—	NDCA ²¹⁸⁾
1218	3 α -Hydroxy-24-nor-5 β -cholan-23-oic acid (Norlithocholic acid)	—	LCA ^{212, 215, 217);} 1217 ²¹⁸⁾
1219	3 β -Hydroxy-24-nor-5 β -cholan-23-oic acid	—	NLCA ²¹²⁾
1220	3 β -Hydroxy-24-norchol-5-en-23-oic acid	—	1313 ²³²⁾

For abbreviations, see Table 1.

Table 13. C₂₂ Bile acids

No.	Systematic name (Trivial name)	Natural source	Synthetic source
1301	1 β , 3 α , 7 α , 12 α -Tetrahydroxy-23, 24-dinor-5 β -cholan-22-oic acid	CTX-U ¹⁷⁰⁾	—
1302	2 β , 3 α , 7 α , 12 α -Tetrahydroxy-23, 24-dinor-5 β -cholan-22-oic acid	CTX-U ¹⁷⁰⁾	—
1303	3 α , 7 α , 12 α -Trihydroxy-23, 24-dinor-5 β -cholan-22-oic acid (Bisnorcholic acid)	CTX ¹⁸²⁾ , -U ¹⁷⁰⁾	NCA ^{190, 229, 234)}
1304	3 α , 12 α -Dihydroxy-7-oxo-23, 24-dinor-5 β -cholan-22-oic acid	CTX-U ¹⁷⁰⁾	BNCA ¹⁷⁰⁾
1305	3 α , 7 α -Dihydroxy-12-oxo-23, 24-dinor-5 β -cholan-22-oic acid	CTX-U ¹⁷⁰⁾	BNCA ^{170, 234)}
1306	3 α , 6 α -Dihydroxy-23, 24-dinor-5 β -cholan-22-oic acid (Bisnorhyodeoxycholic acid)	—	NHDCA ^{151, 186)}
1307	3 α , 7 α -Dihydroxy-23, 24-dinor-5 β -cholan-22-oic acid (Bisnorchenodeoxycholic acid)	—	NCDCA ^{113), 1305⁹⁹⁾}
1308	3 α , 12 α -Dihydroxy-23, 24-dinor-5 β -cholan-22-oic acid (Bisnordeoxycholic acid)	—	NDCA ¹²⁴⁾
1309	3 α -Hydroxy-12-oxo-23, 24-dinor-5 β -cholan-22-oic acid	—	BNDCA ²¹⁸⁾
1310	3 α -Hydroxy-23, 24-dinor-5 α -cholan-22-oic acid or 3 β -Hydroxy-23, 24-dinor-5 β -cholan-22-oic acid	Human-S ²¹¹⁾	1313 ²¹⁰⁾
1311	3 α -Hydroxy-23, 24-dinor-5 β -cholan-22-oic acid (Bisnorlithocholic acid)	Human-M ²¹⁰⁾	1313 ²¹⁰⁾
1312	3 β -Hydroxy-23, 24-dinor-5 α -cholan-22-oic acid	Human-S ²¹¹⁾	1313 ²¹⁰⁾
1313	3 β -Hydroxy-23, 24-dinorchol-5-en-22-oic acid	Human-S ²¹¹⁾ , -M ²¹⁰⁾	Stigmasterol ⁶⁰⁾

For abbreviations, see Table 1.

excluded are 11-oxygenated derivatives and $\Delta^{20(22)}$ -norcholeenoic acids, which had some passing importance in the manufacture of steroid hormones. It has been known that some microorganisms degraded cholesterol and C₂₄ bile acids to bisnorcholanoic acid derivatives. However, these microbial metabolites are not included in the tables.

The first recognition of the natural occurrence of bile acids with the shortened side chain came from the studies, in 1977, of Alme et al who found a trace amount of unconjugated norcholic acid (XVII) in urine from healthy and liver diseased humans²⁾. In healthy humans, the C₂₃ bile acid (XVII) is thought to be formed from the corresponding C₂₄ bile acid, cholic acid (XXXI), by shortening of the side chain by one carbon atom

(α -oxidation) (Fig. 12). Matoba et al¹⁸²⁾ and Kuramoto et al¹⁷⁰⁾ have found a relatively large amount of C₂₃ and C₂₂ bile acids in the bile and urine from patients with CTX, respectively. It seems unlikely that these short side chain bile acids are derived from common C₂₄ bile acids by α - and β -oxidations, since the production of C₂₄ bile acids is below normal in this disease. We postulate, therefore, that the increased formation of the short side chain bile acids in CTX is ascribed to the degradation of some 22- and 23-hydroxylated bile alcohols (e.g. VI) which are synthesized unusually in patients with CTX (Fig. 12).

(Received February 28, 1994)

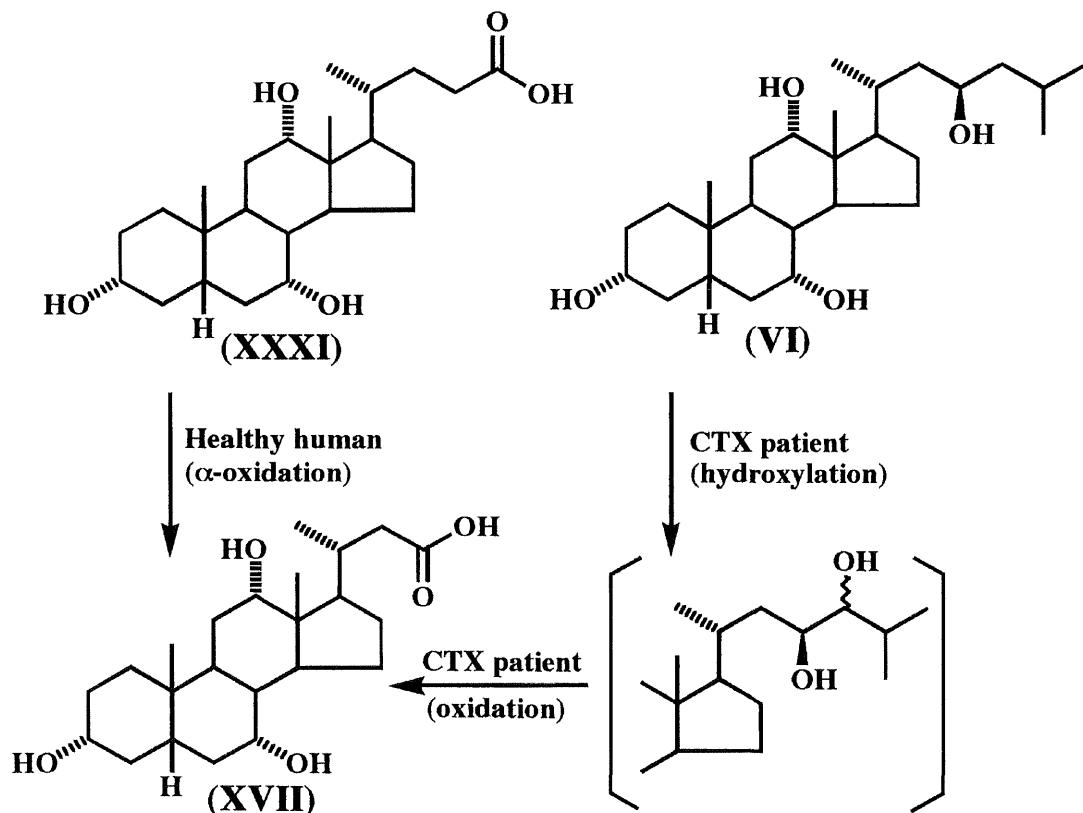


Fig. 12. Postulated pathways for the formation of norcholic acid in healthy persons and in patients with CTX. XXXI, cholic acid; XVII, norcholic acid; VI, (23R)-5 β -cholestane-3 α ,7 α ,12 α ,23-tetrol.

REFERENCES

- Ali, S.S., Stephenson, E. and Elliott, W.H. 1982. Bile acids. LXVII. The major bile acids of *Varanus monitor*. *J. Lipid Res.* **23**: 947–954.
- Alme, B., Bremmelgaard, A., Sjovall, J. and Thomassen, P. 1977. Analysis of metabolic profiles of bile acids in urine using a lipophilic anion exchanger and computerized gas-liquid chromatography-mass spectrometry. *J. Lipid Res.* **18**: 339–362.
- Alme, B., Norden, A. and Sjovall, J. 1978. Glucuronides of unconjugated 6-hydroxylated bile acids in urine of a patient with malabsorption. *Clin. Chim. Acta* **86**: 251–259.
- Amimoto, K., Hoshita, T. and Kazuno, T. 1965. Studies on the bile of the turtle. *J. Biochem.* **57**: 565–570.
- Amimoto, K. 1966. Bile salts of the salamander, *Megalobatrachus japonicus*. *J. Biochem.* **59**: 340–343.
- Amos, B., Anderson, I.G., Haslewood, G.A.D. and Tokes, L. 1977. Bile salts of the lungfishes *Lepidosiren*, *Neoceratodus* and *Protopterus* and those of the coelacanth *Latimeria chalumnae* Smith. *Biochem. J.* **161**: 201–204.
- Amuro, Y., Hayashi, E., Endo, T., Higashino, K. and Kishimoto, S. 1983. Unusual trihydroxylated bile acids in urine of patients with liver cirrhosis. *Clin. Chim. Acta* **127**: 61–67.
- Anderson, I.G. and Haslewood, G.A.D. 1962. Comparative studies of 'bile salts'. 15. The natural occurrence and preparation of allocholic acid. *Biochem. J.* **85**: 236–242.
- Anderson, I.G., Briggs, T. and Haslewood, G.A.D. 1964. Comparative studies of 'bile salts'. 18. The chemistry of cyprinol. *Biochem. J.* **90**: 303–308.
- Anderson, I.G. and Haslewood, G.A.D. 1964. Comparative studies of 'bile salts'. 20. Bile salts of the coelacanth, *Latimeria chalumnae* Smith. *Biochem. J.* **93**: 34–39.
- Anderson, I.G., Haslewood, G.A.D., Cross, A.D. and Tokes, L. 1967. New evidence for the structure of myxinol. *Biochem. J.* **104**: 1061–1063.
- Anderson, I.G. and Haslewood, G.A.D. 1969. Comparative studies of 'bile salts'. 16-Deoxymyxinol, a second bile alcohol from hagfish. *Biochem. J.* **112**: 763–765.
- Anderson, I.G. and Haslewood, G.A.D. 1970. Comparative studies of 'bile salts'. 5 α -Chimaerol, a new bile alcohol from the white sucker *Catostomus commersoni* Lacepede. *Biochem. J.* **116**: 581–587.
- Anderson, I.G., Haslewood, G.A.D., Oldham, R.S., Amos, B. and Tokes, L. 1974. A more detailed study of bile salt evolution, including techniques for small-scale identification and their application to amphibian biles. *Biochem. J.* **141**: 485–494.
- Anderson, I.G., Banister, K.E., Haslewood,

- G.A.D., Cho, D. and Tokes, L.** 1980. Bile salts of fishes collected on the Zaire river expedition (1974–5): their chemical nature and its possible significance. *Zool. J. Linn. Soc.* **68**: 41–51.
16. **Axelson, M., Mork, B. and Sjovall, J.** 1988. Occurrence of 3 β -hydroxy-5-cholestenoic acid, 3 β ,7 α -dihydroxy-5-cholestenoic acid, and 7 α -hydroxy-3-oxo-4-cholestenoic acid as normal constituents in human blood. *J. Lipid Res.* **29**: 629–641.
17. **Axelson, M., Mork, B., Aly, A., Walldius, G. and Sjovall, J.** 1989. Concentrations of cholestenoic acids in plasma from patients with reduced intestinal reabsorption of bile acids. *J. Lipid Res.* **30**: 1883–1887.
18. **Axelson, M., Mork, B., Aly, A., Wisen, O. and Sjovall, J.** 1989. Concentrations of cholestenoic acids in plasma from patients with liver disease. *J. Lipid Res.* **30**: 1877–1882.
19. **Back, P. and Walter, K.** 1980. Developmental pattern of bile acid metabolism as revealed by bile acid analysis of meconium. *Gastroenterology* **78**: 671–676.
20. **Batta, A.K., Dayal, B., Tint, G.S., Shefer, S., Toome, V., Salen, G. and Mosbach, E.H.** 1978. Preparation and characterization of (24R and 24S)-5 β -cholestane-3 α , 7 α , 24, 25-tetrols and (24R and 24S)-5 β -cholestane-3 α , 24, 25-triols. *Steroids* **31**: 99–111.
21. **Batta, A.K., Salen, G., Tint, G.S. and Shefer, S.** 1979. Improved synthesis of 5 β -cholestane-26-oic acids. *Steroids* **33**: 589–594.
22. **Batta, A.K., Salen, G., Blount, J.F. and Shefer, S.** 1979. Configuration at C-25 in 3 α , 7 α , 12 α -tri-hydroxy-5 β -cholestane-26-oic acid by X-ray crystallography. *J. Lipid Res.* **20**: 935–940.
23. **Batta, A.K., Tint, G.S., Dayal, B., Shefer, S. and Salen, G.** 1982. Improved synthesis of 3 α , 7 α , 12 α , 24 ξ -tetrahydroxy-5 β -cholestane-26-oic acid. *Steroids* **39**: 693–702.
24. **Batta, A.K., Salen, G., Shefer, S., Dayal, B. and Tint, G.S.** 1983. Configuration at C-25 in 3 α , 7 α , 12 α -tri-hydroxy-5 β -cholestane-26-oic acid isolated from human bile. *J. Lipid Res.* **24**: 94–96.
25. **Batta, A.K., Mirchandani, R., Salen, G. and Shefer, S.** 1992. Synthesis of 3 α , 7 α -dihydroxy-5 β -cholestane-26-oic acid from 3 α , 7 α , 12 α -tri-hydroxy-5 β -cholestane-26-oic acid: configuration in the bile of *Alligator mississippiensis*. *Steroids* **57**: 162–166.
26. **Bergstrom, S., Paabo, K. and Rumpf, J.A.** 1954. Synthesis and metabolism of 3 α , 7 α , 12 α [4- 14 C] coprostanone. *Acta Chem. Scand.* **8**: 1109–1110.
27. **Berseus, O., Danielsson, H. and Kallner, A.** 1965. Synthesis and metabolism of cholesta-4-ene-7 α ,12 α -diol-3-one and 5 β -cholestane-7 α ,12 α -diol-3-one. *J. Biol. Chem.* **240**: 2396–2401.
28. **Bjorkhem, I.** 1992. Mechanism of degradation of the steroid side chain in the formation of bile acids. *J. Lipid Res.* **33**: 455–471.
29. **Bremmelgaard, A. and Sjovall, J.** 1979. Bile acid profiles in urine of patients with liver diseases. *Eur. J. Clin. Invest.* **9**: 341–348.
30. **Bremmelgaard, A. and Alme, B.** 1980. Analysis of plasma bile acid profiles in patients with liver diseases associated with cholestasis. *Scand. J. Gastroent.* **15**: 593–600.
31. **Bridgwater, R.J.** 1956. Partial synthesis of the two 3 α :7 α :12 α -trihydroxycoprostanic acids and of similar bile acids with extended side chains. *Biochem. J.* **64**: 593–599.
32. **Bridgwater, R.J., Briggs, T. and Haslewood, G.A.D.** 1962. Comparative studies of 'bile salts'. 14. Isolation from shark bile and partial synthesis of scymnol. *Biochem. J.* **82**: 285–290.
33. **Bridgwater, R.J., Haslewood, G.A.D. and Watt, J.R.** 1963. Comparative studies of 'bile salts'. 17. A bile alcohol from *Chimaera monstrosa*. *Biochem. J.* **87**: 28–31.
34. **Briggs, T.** 1970. Partial synthesis of 25D- and 25L-cholestanoic acids from some common bile acids. *J. Org. Chem.* **35**: 1431–1434.
35. **Carey, Jr., J.B. and Haslewood, G.A.D.** 1963. Crystallization of trihydroxycoprostanic acid from human bile. *J. Biol. Chem.* **238**: PC855–856.
36. **Carlson, G.L., Belobaba, D.T.E., Hofmann, A.F. and Wedmid, Y.** 1977. 24-Nor-5 β -chol-22-enes derived from the major bile acids by oxidative decarboxylation. *Steroids* **30**: 787–793.
37. **Clayton, P.T., Muller, D.P.R. and Lawson, A.M.** 1982. The bile acid composition of gastric contents from neonates with high intestinal obstruction. *Biochem. J.* **206**: 489–498.
38. **Clayton, P.T., Lake, B.D., Hall, N.A., Shortland, D.B., Carruthers, R.A. and Lawson, A.M.** 1987. Plasma bile acids in patients with peroxisomal dysfunction syndromes: analysis by capillary gas chromatography-mass spectrometry. *Eur. J. Pediat.* **146**: 166–173.
39. **Clayton, P.T., Patel, E., Lawson, A.M., Carruthers, R.A. and Collins, J.** 1990. Bile acid profiles in peroxisomal 3-oxoacyl-Coenzyme A thiolase deficiency. *J. Clin. Invest.* **85**: 1267–1273.
40. **Cohen, B.I., Tint, G.S., Kuramoto, T. and Mosbach, E.H.** 1975. New bile alcohols—Synthesis of 5 β -cholestane-3 α , 7 α , 25-triol and 5 β -cholestane-3 α , 7 α , 25–24(14 C)-triol. *Steroids* **25**: 365–378.
41. **Collings, B.G. and Haslewood, G.A.D.** 1966. The chemical nature of varanic acid. *Biochem. J.* **99**: 50p.
42. **Cross, A.D.** 1961. Scymnol sulfate and anhydroscymnol. *J. Chem. Soc.* 2817–2821.
43. **Cross, A.D., Landis, P.W. and Murphy, J.W.** 1965. Steroids CCLXXVII. Spectra and stereochemistry XIX. A comparative study of the NMR spectra of some steroids in five solvents. The structure of 'Acipenserol-A'. *Steroids* **5**: 655–662.
44. **Dayal, B., Shefer, S., Tint, G.S., Salen, G. and Mosbach, E.H.** 1976. Synthesis of 5 β -cholestane-3 α , 7 α , 12 α , 25-tetrol and 5 β -cholestane-3 α , 7 α , 12 α , 24 ξ , 25-pentol. *J. Lipid Res.* **17**: 74–77.
45. **Dayal, B., Shefer, S., Tint, G.S., Salen, G. and Mosbach, E.H.** 1976. C₂₆-Analogs of naturally occurring C₂₇ bile alcohols. *J. Lipid Res.* **17**: 478–484.
46. **Dayal, B., Salen, G., Tint, G.S., Toome, V., Shefer, S. and Mosbach, E.H.** 1978. Absolute configuration of pentahydroxy bile alcohols excreted by patients with cerebrotendinous xanthomatosis: a circular dichroism study. *J. Lipid Res.*

- Res. **19**: 187–190.
47. **Dayal, B., Batta, A.K., Shefer, S., Tint, G.S., Salen, G. and Mosbach, E.H.** 1978. Preparation of 24(R)- and 24(S)-5β-cholestane-3α, 7α, 24-triols and 25(R)- and 25(S)-5β-cholestane-3α, 7α, 26-triols by a hydroboration procedure. *J. Lipid Res.* **19**: 191–196.
 48. **Dayal, B., Batta, A.K., Shefer, S., Tint, G.S. and Salen, G.** 1978. Synthesis of biological precursors of cholic acid. *Steroids* **32**: 337–344.
 49. **Dayal, B., Tint, G.S. and Salen, G.** 1979. C₂₆-Analogs of naturally occurring bile alcohols-II. Preparation of 24-nor-5β-cholestane-3α, 7α, 12α, 23-tetrols (23R and 23S) and 24-nor-5β-cholestane-3α, 7α, 12α, 26-tetrols (25R and 25S) by a hydroboration procedure. *Steroids* **34**: 581–588.
 50. **Dayal, B., Bagan, E., Speck, J. and Salen, G.** 1980. A facile synthesis of 5β-cholestane-3α, 7α, 12α, 25-tetrol. *Steroids* **35**: 439–444.
 51. **Dayal, B., Tint, G.S., Batta, A.K., Shefer, S. and Salen, G.** 1981. Synthesis of biological precursors of cholic acid II. *Steroids* **37**: 205–211.
 52. **Dayal, B., Tint, G.S., Greeley, D.N., Williams, T.H. and Salen, G.** 1983. Identification of 5β-cholestane-3α, 7α, 12α, 25, 26-pentol in cerebrotendinous xanthomatosis. *Steroids* **42**: 441–448.
 53. **Dayal, B., Greeley, D.N., Williams, T.H., Tint, G.S. and Salen, G.** 1984. Stereospecific synthesis of 3β-hydroxylated bile alcohols. *J. Lipid Res.* **25**: 646–650.
 54. **Dayal, B., Salen, G., Tint, G.S., Batta, A.K. and Shefer, S.** 1984. Synthesis of the putative metabolites of plant sterols: (24R)- and (24S)-24-methyl-5β-cholestane-3α, 7α, 12α, 25-tetrols and 24-ethyl-5β-cholestane-3α, 7α, 12α, 24ξ-tetrol. *J. Lipid Res.* **25**: 865–870.
 55. **Dayal, B., Tint, G.S., Toome, V., Batta, A.K., Shefer, S. and Salen, G.** 1985. Synthesis and structure of 26 (or 27)-nor-5β-cholestane-3α, 7α, 12α, 24S, 25ξ-pentol isolated from the urine and feces of a patient with sitosterolemia and xanthomatosis. *J. Lipid Res.* **26**: 298–305.
 56. **Deen, P.D.G. and Aplin, R.T.** 1966. Mass spectrometric studies on bile acids: The differentiation between chenodeoxycholic acid and deoxycholic acid and the identification of 3α, 7α-dihydroxy-5β-cholestanoic acid in alligator bile. *Steroids* **8**: 565–579.
 57. **Deleze, G., Bjorkhem, I. and Karlaganis, G.** 1986. Bile acids and bile alcohols in two patients with Zellweger (cerebro-hepato-renal) syndrome. *J. Pediat. Gastroenterol. Nutr.* **5**: 701–710.
 58. **Eyssen, H., Parmentier, G., Compernolle, F., Boon, J. and Eggermont, E.** 1972. Trihydroxycoprostanic acid in the duodenal fluid of two children with intrahepatic bile duct anomalies. *Biochim. Biophys. Acta* **273**: 212–221.
 59. **Eyssen, H., Eggermont, E., van Eldere, J., Jaeken, J., Parmentier, G. and Janssen, G.** 1985. Bile acid abnormalities and the diagnosis of cerebro-hepato-renal syndrome (Zellweger syndrome). *Acta Pediat. Scand.* **74**: 539–544.
 60. **Fernholz, E.** 1933. Über die Konstitution des Stigmasterins. *Liebigs Ann. Chem.* **507**: 128–138.
 61. **Fieser, L.F. and Fieser, M.** 1959. Bile acids and alcohols. p. 421–443. In *Steroids*, Reinhold Publishing Corporation, New York.
 62. **Fujimoto, Y., Iwadate, H., Ikekawa, N., Kihira, K. and Hoshita, T.** 1985. Structure of the steroid lactone isolated from turtle bile: (22S, 25R)-3α, 7α, 12α-trihydroxy-5β-cholestano-26, 22-lactone. *J. Chem. Soc. Perkin Trans. I.* 2701–2704.
 63. **Gengenbacher, T., Gerok, W., Giese, U. and Kurz, G.** 1990. Synthesis and applicability of photolabile 7, 7-azo analogues of natural bile salt precursors. *J. Lipid Res.* **31**: 315–327.
 64. **Gustafsson, J., Sisfontes, L. and Bjorkhem, I.** 1987. Diagnosis of Zellweger syndrome by analysis of bile acids and plasmalogens in stored dried blood collected at neonatal screening. *J. Pediat.* **111**: 264–267.
 65. **Hammarsten, O.** 1898. Ueber eine neue Gruppe gepaarter Gallensaure. *Z. Physiol. Chem.* **24**: 322–350.
 66. **Hanson, R.F. and Williams, G.** 1971. The isolation and identification of 3α, 7α-dihydroxy-5β-cholestean-26-oic acid from human bile. *Biochem. J.* **121**: 863–864.
 67. **Hanson, R.F., Isenberg, J.N., Williams, G.C., Hachey, D., Szczepanik, P., Klein, P.D. and Sharp, H.L.** 1975. The metabolism of 3α, 7α, 12α-trihydroxy-5β-cholestean-26-oic acid in two siblings with cholestasis due to intrahepatic bile duct anomalies. *J. Clin. Invest.* **56**: 577–587.
 68. **Hanson, R.F., Szczepanik-Van Leeuwen, P., Williams, G.C., Grabowski, G. and Sharp, H.L.** 1979. Defects of bile acid synthesis in Zellweger's syndrome. *Science* **203**: 1107–1108.
 69. **Haslewood, G.A.D. and Wootton, V.** 1950. Comparative studies of 'bile salts'. 1. Preliminary survey. *Biochem. J.* **47**: 584–597.
 70. **Haslewood, G.A.D.** 1952. Comparative studies of 'bile salts'. 5. Bile salts of Crocodylidae. *Biochem. J.* **52**: 583–587.
 71. **Haslewood, G.A.D.** 1961. Comparative studies of 'bile salts'. 13. Bile acids of the leopard seal, *Hydrurga leptonyx*, and of two snakes of the genus *Bitis*. *Biochem. J.* **78**: 352–359.
 72. **Haslewood, G.A.D.** 1964. Comparative studies of 'bile salts'. 19. The chemistry of ranol. *Biochem. J.* **90**: 309–313.
 73. **Haslewood, G.A.D.** 1966. Comparative studies of 'bile salts'. Myxinol disulfate, the principal bile salt of hagfish (*Mixinidae*). *Biochem. J.* **100**: 233–237.
 74. **Haslewood, G.A.D.** 1967. Bile salt evolution. *J. Lipid Res.* **8**: 535–550.
 75. **Haslewood, G.A.D.** 1967. Bile salts. p. 1–116., Methuen Co. Ltd., London.
 76. **Haslewood, G.A.D. and Tammer, A.R.** 1968. Comparative studies of bile salts. Bile salts of sturgeons (Acipenseridae) and of the paddlefish *Polyodon spathula*: A new partial synthesis of 5β-cyprinol. *Biochem. J.* **108**: 263–268.
 77. **Haslewood, G.A.D. and Tokes, L.** 1969. Comparative studies of bile salts. Bile salts of the lamprey *Petromyzon marinus* L. *Biochem. J.* **114**: 179–184.
 78. **Haslewood, G.A.D. and Tokes, L.** 1972. Comparative studies of bile salts. A new type of bile

- salt from *Arapaima gigas* (Cuvier) (Family Osteoglossidae). Biochem. J. **126**: 1161–1170.
79. Haslewood, G.A.D., Ikawa, S., Tokes, L. and Wong, D. 1978. Bile salts of the green turtle *Chelonia mydas* (L.). Biochem. J. **171**: 409–412.
 80. Hayakawa, S. 1953. Studies on bile salts of toad (*Bufo vulgaris japonicus*). X. Separation of Δ^{23} -3(α), 7(α), 12(α)-trihydroxycoprostanic acid. Proc. Japan Acad. **29**: 279–284.
 81. Hayakawa, S. 1953. Studies on bile salts of toad (*Bufo vulgaris japonicus*). XI. Formation of norcholic acid from Δ^{23} -3(α), 7(α), 12(α)-trihydroxycoprostanic acid $C_{27}H_{44}O_5$ and stem acid $C_{27}H_{46}O_2$. Proc. Japan Acad. **29**: 285–288.
 82. Hiraoka, T., Kihira, K., Kajiyama, G., Kuramoto, T. and Hoshita, T. 1987. Identification of 5 β -cholestane-3 α , 7 α , 12 α , 24, 25, 26-hexol in human urine. J. Lipid Res. **28**: 895–899.
 83. Hiraoka, T., Kihira, K., Kohda, T., Kosaka, D., Kajiyama, G. and Hoshita, T. 1987. Urinary bile alcohols in liver dysfunction. Clin. Chim. Acta **169**: 127–132.
 84. Hiraoka, T., Kihira, K., Kosaka, D., Kohda, T., Hoshita, T. and Kajiyama, G. 1988. Identification of bile alcohols in serum from healthy humans. Steroids **51**: 543–550.
 85. Hiraoka, T., Kosaka, D., Kajiyama, G., Kohda, T., Funakura, T., Yamauchi, T., Kihira, K. and Hoshita, T. 1988. Measurement of serum bile alcohol levels in liver dysfunction, using isotope dilution-mass spectrometry. Scand. J. Gastroenterol. **23**: 821–826.
 86. Hiraoka, T., Kohda, T., Kosaka, D., Yamauchi, T., Kihira, K., Kuramoto, T., Hoshita, T. and Kajiyama, G. 1989. Identification of bile alcohols in rat bile. J. Lipid Res. **30**: 1889–1893.
 87. Hiremath, S.V. and Elliott, W.H. 1981. Bile acids. LXIV. Synthesis of 5 α -cholestane-3 α , 7 α , 25-triol and esters of new 5 α -bile acids. Steroids **38**: 465–475.
 88. Hoshita, N. and Okuda, K. 1967. Partial synthesis of sterobilic acids related to chenodeoxycholic acid. J. Biochem. **62**: 655–657.
 89. Hoshita, T. 1959. The partial synthesis of trihydroxy-24-methylcoprostanic acid. J. Biochem. **46**: 507–511.
 90. Hoshita, T. 1962. Isolation of a new bile sterol, 3 α , 7 α , 12 α -trihydroxy-26, 27-epoxycholestane from carp bile. J. Biochem. **52**: 125–130.
 91. Hoshita, T. 1962. Syntheses of 3 α , 7 α , 12 α , 25 ξ , 26- and 3 α , 7 α , 12 α , 24 ξ , 25-pentahydroxycoprostanes. J. Biochem. **52**: 176–179.
 92. Hoshita, T., Kouchi, M. and Kazuno, T. 1963. Synthesis of 3 α , 7 α , 12 α , 26, 27-pentahydroxycoprostanone. J. Biochem. **53**: 291–294.
 93. Hoshita, T., Nagayoshi, S. and Kazuno, T. 1963. Studies on the bile of carp. J. Biochem. **54**: 369–374.
 94. Hoshita, T., Nagayoshi, S., Kouchi, M. and Kazuno, T. 1964. Studies on the bile of the family Cyprinidae. J. Biochem. **56**: 177–181.
 95. Hoshita, T., Yukawa, M. and Kazuno, T. 1964. The isolation of a new bile alcohol, 5 β -cholestane-3 α , 7 α , 12 α , 26, 27-pentol from the bile of *Conger myriaster*. Steroids **4**: 569–574.
 96. Hoshita, T., Sasaki, T. and Kazuno, T. 1965. Isolation of a bile alcohol, 5 α -cholestane-3 α , 7 α , 12 α , 26-tetrol from carp bile. Steroids **5**: 241–247.
 97. Hoshita, T., Sasaki, T., Tanaka, Y., Betsuki, S. and Kazuno, T. 1965. Biosynthesis of bile acids and bile alcohols in toad. J. Biochem. **57**: 751–757.
 98. Hoshita, T., Hirofuji, S., Nakagawa, T. and Kazuno, T. 1967. Studies on the bile salts of the newt and synthesis of 5 α -cholestane-3 α , 7 α , 12 α , 25, 26-pentol (5 α -bufol). J. Biochem. **62**: 62–66.
 99. Hoshita, T., Hirofuji, S., Sasaki, T. and Kazuno, T. 1967. Isolation of a new bile acid, haemulcholic acid from the bile of *Parapristipoma trilineatum*. J. Biochem. **61**: 136–141.
 100. Hoshita, T., Okuda, K. and Kazuno, T. 1967. Synthesis of 3 α , 7 α , 12 α -trihydroxy-5 β -cholestane-24-carboxylic acid and the chemical structure of trihydroxybufosterochenic acid isolated from toad bile. J. Biochem. **61**: 756–759.
 101. Hoshita, T. and Kazuno, T. 1968. Chemistry and metabolism of bile alcohols and higher bile acids. p. 207–254. In Paoletti, R. and Kritchevsky, D. (eds.), Advances in Lipid Research. Vol 6., Academic Press, Inc., New York.
 102. Hoshita, T., Shefer, S. and Mosbach, E.H. 1968. Conversion of 7 α , 12 α -dihydroxycholest-4-en-3-one to 5 α -cholestane-3 α , 7 α , 12 α -triol by iguana liver microsomes. J. Lipid Res. **9**: 237–243.
 103. Hoshita, T., Yasuhara, M., Kihira, K. and Kuramoto, T. 1976. Identification of (23S)-5 β -cholestane-3 α , 7 α , 12 α , 23, 25-pentol in cerebrotendinous xanthomatosis. Steroids **27**: 657–664.
 104. Hoshita, T., Yasuhara, M., Une, M., Kibe, A., Itoga, E., Kito, S. and Kuramoto, T. 1980. Occurrence of bile alcohol glucuronides in bile of patients with cerebrotendinous xanthomatosis. J. Lipid Res. **21**: 1015–1021.
 105. Hoshita, T. 1985. Bile alcohols and primitive bile acids. p. 279–302. In Danielsson, H. and Sjovall, J. (eds.), Sterols and bile acids. New comprehensive biochemistry. Vol 12, Elsevier, Amsterdam.
 106. Ichimiya, H., Yanagisawa, J. and Nakayama, F. 1984. Significance of bile alcohol in urine of a patient with cholestasis: Identification of 5 β -cholestane-3 α , 7 α , 12 α , 26, 27-pentol (5 β -cyprinol) and 5 β -cholestane-3 α , 7 α , 12 α , 26-tetrol (27-deoxy-5 β -cypri-
 - rinol). Chem. Pharm. Bull. **32**: 2874–2877.
 107. Ichimiya, H., Yanagisawa, J. and Nakayama, F. 1987. Altered metabolism of bile alcohol and bile acid in complete extrahepatic cholestasis: qualitative and quantitative aspects. J. Lipid Res. **28**: 1028–1037.
 108. Ichimiya, H., Egestad, B., Nazer, H., Baginski, E.S., Clayton, P.T. and Sjovall, J. 1991. Bile acids and bile alcohols in a child with hepatic 3 β -hydroxy- Δ^5 -C₂₇-steroid dehydrogenase deficiency: effects of chenodeoxycholic acid treatment. J. Lipid Res. **32**: 829–841.
 109. Inai, Y., Tanaka, Y., Betsuki, S. and Kazuno, T. 1964. Synthesis of 3 α , 7 α , 12 α , 24 ξ -tetrahydroxycoprostanic acid. J. Biochem. **56**: 591–593.
 110. Iqbal, M.N., Patrick, P.H. and Elliott, W.H. 1991. Bile acids. LXXXI. Synthesis and structural assignment of E/Z isomers of substituted methyl

- hydroxy-5 β -cholest-24-en-26-oates. *Steroids* **56**: 505–512.
111. **Ishida, H., Yamaguchi, T., Natsuyama, R., Tsuji, K. and Kosuge, T.** 1988. Study on the bile salt sodium scymnol sulfate, from *Rhizopriionodon acutus*. *Chem. Pharm. Bull.* **36**: 4408–4413.
112. **Ishida, H., Kinoshita, S., Natsuyama, R., Nukaya, H., Tsuji, K., Kosuge, T. and Yamaguchi, K.** 1991. Study on the bile salt, sodium scymnol sulfate, from *Rhizopriionodon acutus*. II. The structures of scymnol, anhydroscymnol and sodium scymnol sulfate. *Chem. Pharm. Bull.* **39**: 3153–3156.
113. **Ishihara, T.** 1937. Über den systematischen Abbau der Chenodesoxycholsäure. *J. Biochem.* **27**: 265–277.
114. **Janssen, G. and Parmentier, G.** 1981. A further study of the bile acids in infants with coprostanic acidemia. *Steroids* **37**: 81–89.
115. **Janssen, G., Toppet, S. and Parmentier, G.** 1982. Structure of the side chain of the C₂₉ dicarboxylic acid occurring in infants with coprostanic acidemia. *J. Lipid Res.* **23**: 456–465.
116. **Joyce, M.J., Hiremath, S.V., Mattamal, M.B. and Elliott, W.H.** 1984. Bile acids. LXXIII. Synthesis of analogs of 7 α -hydroxy-4-cholest-3-one as substrates for hepatic steroid 12 α -hydroxylase. *Steroids* **44**: 95–101.
117. **Kamat, S.Y. and Elliott, W.H.** 1972. Bile acids XXXVI. Synthesis of 5 α -cholest-26-oic acids. *Steroids* **20**: 279–294.
118. **Kanemitsu, T.** 1942. Über eine isomere Cholsäure, Heterocholsäure aus der Alligatorschildkröten-galle. *J. Biochem.* **35**: 409–412.
119. **Karlaganis, G., Alme, B., Karlaganis, V. and Sjovall, J.** 1981. Bile alcohol glucuronides in urine. Identification of 27-nor-5 β -cholestane-3 α , 7 α , 12 α , 24 ξ , 25 ξ -pentol in man. *J. Steroid Biochem.* **14**: 341–345.
120. **Karlaganis, G., Karlaganis, V. and Sjovall, J.** 1984. Identification of 27-nor-5 β -cholestane-3 α , 7 α , 12 α , 24 ξ , 25 ξ , 26-hexol and partial characterization of the bile alcohol profile in urine. *J. Lipid Res.* **25**: 693–702.
121. **Karlaganis, G., Bradley, S.E., Boyer, J.L., Batta, A.K., Salen, G., Egestad, B. and Sjovall, J.** 1989. A bile alcohol sulfate as a major component in the bile of the small skate (*Raja erinacea*). *J. Lipid Res.* **30**: 317–322.
122. **Kase, B.F., Bjorkhem, I., Haga, P. and Pedersen, J.I.** 1985. Defective peroxisomal cleavage of the C₂₇-steroid side chain in the cerebro-hepato-renal syndrome of Zellweger. *J. Clin. Invest.* **75**: 427–435.
123. **Kase, B.F., Pedersen, J.I., Strandvik, B. and Bjorkhem, I.** 1985. In vivo and in vitro studies on formation of bile acids in patients with Zellweger syndrome. *J. Clin. Invest.* **76**: 2393–2402.
124. **Kazuno, T. and Shimizu, T.** 1939. Über den Abbau der Desoxycholsäure zu Dioxyketon C₁₉H₃₂O₃ und 22-Methyl-3,12-diketonorchole(20). *J. Biochem.* **29**: 421–433.
125. **Kazuno, T., Moori, A., Sasaki, K., Kuroda, M. and Mizuguchi, M.** 1952. The synthesis of cholan-, trihydroxycholane and trihydroxyhomocholane. Proc. Japan Acad. **28**: 416–423.
126. **Kazuno, T., Moori, A., Sasaki, K. and Mizuguchi, M.** 1952. The synthesis of trihydroxybisnorsterocholane and homocholic acid. Proc. Japan Acad. **28**: 424–428.
127. **Kazuno, T. and Mori, A.** 1954. The synthesis of trihydroxycoprostane. Proc. Japan Acad. **30**: 486–487.
128. **Kazuno, T., Komatsubara, T. and Baba, T.** 1954. The partial synthesis of bisnorsterocholic acid. Proc. Japan Acad. **30**: 987–990.
129. **Kazuno, T., Mori, A. and Goto, T.** 1955. The partial synthesis of bisnorsterocholic acid by Kolbe electrolytic reaction. *J. Biochem.* **42**: 77–80.
130. **Kazuno, T., Masui, T. and Hoshita, T.** 1961. Isolation of a new bile sterol, 3 α , 7 α , 12 α , 26-tetrahydroxy- Δ^{23} -bishomocholene, from bullfrog bile. *J. Biochem.* **50**: 12–19.
131. **Kazuno, T., Betsuki, S., Tanaka, Y. and Hoshita, T.** 1965. Studies on bile of *Rana nigromaculata*. *J. Biochem.* **58**: 243–247.
132. **Kazuno, T., Masui, T. and Okuda, K.** 1965. The isolation and the chemistry of a new bile alcohol, 3 α , 7 α , 12 α , 24 ξ , 26-pentahydroxybishomocholane from *Rana catesbeiana* bile. *J. Biochem.* **57**: 75–80.
133. **Kibe, A., Nakai, S., Kuramoto, T. and Hoshita, T.** 1980. Occurrence of bile alcohols in the bile of a patient with cholestasis. *J. Lipid Res.* **21**: 594–599.
134. **Kibe, A., Fukura, M., Kihira, K., Kuramoto, T. and Hoshita, T.** 1981. Metabolism of bile alcohols, 24-nor-5 β -cholestane-3 α , 7 α , 12 α , 25-tetrol and 3 α , 7 α , 12 α -trihydroxy-26, 27-dinor-5 β -cholest-24-one, in rats. *J. Biochem.* **89**: 369–377.
135. **Kihira, K., Kuramoto, T. and Hoshita, T.** 1976. New bile alcohols—Synthesis of (22R)- and (22S)-5 β -cholestane-3 α , 7 α , 12 α , 22, 25-pentols. *Steroids* **27**: 383–393.
136. **Kihira, K., Yasuhara, M., Kuramoto, T. and Hoshita, T.** 1977. New bile alcohols, 5 α - and 5 β -dermophols from amphibians. *Tetrahedron Lett.* 687–690.
137. **Kihira, K., Une, M., Kuramoto, T. and Hoshita, T.** 1979. Synthesis of 5 α -cholestane-3 α , 7 α , 12 α , 25, 26, 27-hexol. *Hiroshima J. Med. Sci.* **28**: 167–172.
138. **Kihira, K., Batta, A.K., Mosbach, E.H. and Salen, G.** 1979. Reverse cross-coupling in the synthesis of 3 α , 7 α -dihydroxy-5 β -cholestanoic acid. *J. Lipid Res.* **20**: 421–427.
139. **Kihira, K., Morioka, Y. and Hoshita, T.** 1981. Synthesis of (22R) and (22S)-3 α , 7 α , 22-trihydroxy-5 β -cholan-24-oic acids and structure of haemulcholic acid, a unique bile acid isolated from fish bile. *J. Lipid Res.* **22**: 1181–1187.
140. **Kihira, K., Ohira, S., Kuramoto, M., Kuramoto, J., Nakayama, M. and Hoshita, T.** 1982. Configuration at C-23 in 5 β -cholestane-3 α , 7 α , 12 α , 23-tetrol excreted by patients with cerebrotendinous xanthomatosis. *Chem. Pharm. Bull.* **30**: 3040–3041.
141. **Kihira, K., Kubota, A. and Hoshita, T.** 1984. Absolute configuration at a carbon 23 of 5 β -cholestane-3 α , 7 α , 12 α , 23,25-pentol excreted by patients with cerebrotendinous xanthomatosis. *J. Lipid Res.* **25**: 871–875.
142. **Kihira, K., Akashi, Y., Kuroki, S., Yanagisawa,**

- J., Nakayama, F. and Hoshita, T.** 1984. Bile salts of the coelacanth, *Latimeria chalumnae*. *J. Lipid Res.* **25:** 1330–1336.
- 143.**Kihira, K. and Hoshita, T.** 1985. Synthesis of α,β -unsaturated C₂₄-bile acids. *Steroids* **46:** 767–774.
- 144.**Kihira, K., Shimazu, K., Kuwabara, M., Yoshii, M., Takeuchi, H., Nakano, I., Ozawa, S., Onuki, M., Hatta, Y. and Hoshita, T.** 1986. Bile acid profiles in bile, urine, and feces of a patient with cerebrotendinous xanthomatosis. *Steroids* **48:** 109–119.
- 145.**Kihira, K., Noma, Y., Tsuda, K., Watanabe, T., Yamamoto, Y., Une, M. and Hoshita, T.** 1986. Absolute configuration at C-24 of 5 β -ranol, a principal bile alcohol of the bullfrog. *J. Lipid Res.* **27:** 393–397.
- 146.**Kihira, K., Kosaka, D., Une, M., Hiraoka, T., Kajiyama, G. and Hoshita, T.** 1987. Syntheses of deuterium labeled bile alcohols. *J. Label. Comp. Radiopharm.* **24:** 1421–1428.
- 147.**Kihira, K., Okamoto, A. and Hoshita, T.** 1987. Identification of new C₂₇ and C₂₄ bile acids in the bile of *Alligator mississippiensis*. *J. Biochem.* **101:** 1377–1384.
- 148.**Kihira, K., Yoshii, M., Okamoto, A., Ikawa, S., Ishii, H. and Hoshita, T.** 1990. Synthesis of new bile salt analogues, sodium 3 α , 7 α -dihydroxy-5 β -cholane-24-sulfonate and sodium 3 α , 7 β -dihydroxy-5 β -cholane-24-sulfonate. *J. Lipid Res.* **31:** 1323–1326.
- 149.**Kihira, K., Fukuda, K., Kuramoto, T., Kuriyama, M., Fujiyama, J., Osame, M. and Hoshita, T.** 1991. Identification of (23S)-5 α -cholestane-3 α , 7 α , 12 α , 23, 25-pentol in urine of patients with cerebrotendinous xanthomatosis. *Steroids* **56:** 464–468.
- 150.**Kihira, K., Mikami, T., Ikawa, S., Okamoto, A., Yoshii, M., Miki, S., Mosbach, E.H. and Hoshita, T.** 1992. Synthesis of sulfonate analogs of bile acids. *Steroids* **57:** 193–198.
- 151.**Kimura, T. and Sugiyama, G.** 1939. Über den Abbau der Hyodesoxycholsäure zur Bisnorhydesoxycholsäure und 6-Oxypregnanol-3-on-20. *J. Biochem.* **29:** 409–419.
- 152.**Kinoshita, T., Miyata, M., Ismail, S.M., Fujimoto, Y., Kakinuma, K., Ikekawa, N. and Morisaki, M.** 1988. Synthesis and determination of stereochemistry of four diastereoisomers at the C-24 and C-25 positions of 3 α , 7 α , 12 α , 24-tetrahydroxy-5 β -cholestan-26-oic acid. *Chem. Pharm. Bull.* **36:** 134–141.
- 153.**Komatsubara, T.** 1954. The synthesis of trihydroxynorcholesterol. *Proc. Japan Acad.* **30:** 488–491.
- 154.**Komatsubara, T.** 1954. On the bile acid of *Rana nigromaculata nigromaculata*. *Proc. Japan Acad.* **30:** 614–617.
- 155.**Komatsubara, T.** 1954. The synthesis of 3(α), 7(α), 12(α)-trihydroxycoprostanic acid. *Proc. Japan Acad.* **30:** 618–621.
- 156.**Koopman, B.J., Wolthers, B.G., van der Molen, J.C., Nagel, G.T. and Kruizinga, W.** 1987. Abnormal urinary bile acids in a patient suffering from cerebrotendinous xanthomatosis during oral administration of ursodeoxycholic acid. *Biochim. Biophys. Acta* **917:** 238–246.
- 157.**Kosaka, D., Hiraoka, T., Kohda, T., Kajiyama, G., Yamauchi, T., Kihira, K., Kuramoto, T. and Hoshita, T.** 1991. Stable isotope dilution assay for 5 β -cholestane-3 α , 7 α , 12 α , 25-tetrol and 5 β -cholestane-3 α , 7 α , 12 α , 23, 25-pentol in human serum using [26, 27-D₆] labeled internal standards; a highly accurate approach to the serological diagnosis of cerebrotendinous xanthomatosis. *Clin. Chim. Acta* **199:** 83–89.
- 158.**Kouchi, M.** 1964. Studies on the bile of shark, *Mustelus manazo*. *Hiroshima J. Med. Sci.* **13:** 341–350.
- 159.**Kritchevsky, D., Davidson, L.M., Mosbach, E.H. and Cohen, B.I.** 1981. Identification of acidic steroids in feces of monkeys fed β -sitosterol. *Lipids* **16:** 77–78.
- 160.**Kuramoto, T. and Hoshita, T.** 1972. The identification of C₂₇-bile acids in kite bile. *J. Biochem.* **72:** 199–201.
- 161.**Kuramoto, T., Kikuchi, H., Sanemori, H. and Hoshita, T.** 1973. Bile salts of anura. *Chem. Pharm. Bull.* **21:** 952–959.
- 162.**Kuramoto, T., Itakura, S. and Hoshita, T.** 1974. Studies on the conversion of mevalonate into bile acids and bile alcohols in toad and the stereospecific hydroxylation at carbon atom 26 during bile alcohol biogenesis. *J. Biochem.* **75:** 853–859.
- 163.**Kuramoto, T., Cohen, B.I. and Mosbach, E.H.** 1977. New bile alcohols II: Synthesis and mass spectra of C₂₆ bile alcohols. *J. Am. Oil Chem. Soc.* **54:** 578–581.
- 164.**Kuramoto, T., Cohen, B.I., Rothschild, M.A., Donor, D.A. and Mosbach, E.H.** 1978. Chemical synthesis of 5 β -cholestane-3 α , 7 α , 24, 25-tetrol and its metabolism in the perfused rabbit liver. *J. Biol. Chem.* **253:** 4688–4692.
- 165.**Kuramoto, T., Matsumoto, N. and Hoshita, T.** 1978. Syntheses of 22- and 23-hydroxylated bile alcohols. *Chem. Pharm. Bull.* **26:** 2788–2792.
- 166.**Kuramoto, T., Kihira, K., Matsumoto, N. and Hoshita, T.** 1981. Determination of the sulfated position in 5 β -bufol sulfate by a carbon-13 nuclear magnetic resonance study. *Chem. Pharm. Bull.* **29:** 1136–1139.
- 167.**Kuramoto, T., Noma, Y. and Hoshita, T.** 1983. Synthesis of (24R)- and (24S)-27-nor-5 β -cholestane-3 α , 7 α , 12 α , 24, 26-pentols. *Chem. Pharm. Bull.* **31:** 1330–1334.
- 168.**Kuramoto, T., Kawamoto, K., Moriwaki, S. and Hoshita, T.** 1984. Synthesis of homoursodeoxycholic acid and [11, 12-³H]homoursodeoxycholic acid. *Steroids* **44:** 549–559.
- 169.**Kuramoto, T., Moriwaki, S., Kawamoto, K. and Hoshita, T.** 1987. Intestinal absorption and metabolism of homoursodeoxycholic acid in rats. *J. Pharmacobi-Dyn.* **10:** 309–316.
- 170.**Kuramoto, T., Furukawa, Y., Nishina, T., Sugimoto, T., Mahara, R., Tohma, M., Kihira, K. and Hoshita, T.** 1990. Identification of short side chain bile acids in urine of patients with cerebrotendinous xanthomatosis. *J. Lipid Res.* **31:** 1895–1902.

171. **Kurauti, Y. and Kazuno, T.** 1939. Tetraoxycholan, Trioxycholen und Trioxy-bis-norsterocholansäure aus der Galle von *Rana catesbeiana* Shaw. Z. Physiol. Chem. **262**: 53–60.
172. **Kuroki, S., Shimazu, K., Kuwabara, M., Une, M., Kihira, K., Kuramoto, T. and Hoshita, T.** 1985. Identification of bile alcohols in human bile. J. Lipid Res. **26**: 230–240.
173. **Kuroki, S., Schteingart, C.D., Hagey, L.R., Cohen, B.I., Mosbach, E.H., Rossi, S.S., Hofmann, A.F., Matoba, N., Une, M., Hoshita, T. and Odell, D.K.** 1988. Bile salts of the West Indian manatee, *Trichechus manatus latirostris*: novel bile alcohol sulfates and absence of bile acids. J. Lipid Res. **29**: 509–522.
174. **Kuwabara, M., Ushiroguchi, T., Kihira, K., Kuramoto, T. and Hoshita, T.** 1984. Identification of bile alcohols in urine from healthy humans. J. Lipid Res. **25**: 361–368.
175. **Lawson, A.M., Madigan, M.J., Shortland, D. and Clayton, P.T.** 1986. Rapid diagnosis of Zellweger syndrome and infantile Refsum's disease by fast atom bombardment-mass spectrometry of urine bile salts. Clin. Chim. Acta **161**: 221–231.
176. **Lester, R., Pyrek, J.S., Little, J.M. and Adcock, E.W.** 1983. What is meant by the term "bile acid"? Am. J. Physiol. **244**: G107–G110.
177. **Ludwig-Kohn, H., Henning, H.V., Sziedat, A., Matthaei, D., Spitteler, G., Reiner, J. and Egger, H.J.** 1983. The identification of urinary bile alcohols by gas chromatography-mass spectrometry in patients with liver disease and in healthy individuals. Eur. J. Clin. Invest. **13**: 91–98.
178. **Mabuti, H.** 1941. Über Trioxy-bisnor-sterocholansäure $C_{26}H_{44}O_5$ aus der Galle von *Rana catesbeiana* Shaw. J. Biochem. **33**: 117–130.
179. **Masui, T.** 1963. Isolation of 3α , 7α , 12α -trihydroxy- Δ^{24} -homo- 5α -cholene from bullfrog bile. J. Biochem. **54**: 41–46.
180. **Masui, T. and Staple, E.** 1967. The separation of the stereo-isomers of bile steroids, 5β -cholestane- 3α , 7α , 12α , 24α -tetrol and 5β -cholestane- 3α , 7α , 12α , 24β -tetrol, by thin layer chromatography. Steroids **9**: 443–450.
181. **Mathis, R.K., Watkins, J.B., Szczepanik-Van Leeuwen, P. and Lott, I.T.** 1980. Liver in the cerebro-hepato-renal syndrome: Defective bile acid synthesis and abnormal mitochondria. Gastroenterology **79**: 1311–1317.
182. **Matoba, N., Une, M. and Hoshita, T.** 1986. Identification of unconjugated bile acids in human bile. J. Lipid Res. **27**: 1154–1162.
183. **Matschiner, J.T.** 1971. Naturally occurring bile acids and alcohols and their origins. p. 11–46. In Nair, P.P. and Kritchevsky, D. (eds.), The bile acids. Vol. 1, Plenum Press, New York.
184. **Mendelsohn, D. and Mendersohn, L.** 1969. The catabolism of cholesterol in vitro. Formation of 3α , 7α , 12α -trihydroxy- 5β -cholestanoic acid from cholesterol by rat liver. Biochem. J. **114**: 1–3.
185. **Mitra, M.N. and Elliott, W.H.** 1968. Bile acids. XXIII. A new direct synthesis of allcholic acid and its 3β isomer. J. Org. Chem. **33**: 175–181.
186. **Moffett, R.B., Stafford, J.E., Linsk, J. and Hoehn, W.M.** 1946. Degradation of hyodesoxy-
- cholic acid. J. Am. Chem. Soc. **68**: 1857–1860.
187. **Monnens, L., Bakkeren, J., Parmentier, G., Janssen, G., van Haelst, U., Trijbels, F. and Eysen, H.** 1980. Disturbances in bile acid metabolism of infants with the Zellweger (Cerebro-hepato-renal) syndrome. Eur. J. Pediat. **133**: 31–35.
188. **Morimoto, K.** 1964. The synthesis of trihydroxy- 24 -ethyl-coprostanic acid and chromatography of stero-bile acids. J. Biochem. **55**: 410–414.
189. **Morimoto, K., Kurata, Y., Hoshita, N. and Okuda, K.** 1968. Partial synthesis of stero-bile acids related to lithocholic acid. Hiroshima J. Med. Sci. **17**: 1–6.
190. **Morsman, H., Steiger, M. and Reichstein, T.** 1937. Abbau der Cholsäure zu 3 , 7 , 12 -Trioxy-pregnan-20-on. Helv. Chim. Acta **20**: 3–16.
191. **Moss, G.P.** 1989. IUPAC-IUB Joint Commission on Biochemical Nomenclature (JCBN). The nomenclature of steroids. Recommendations 1989. Eur. J. Biochem. **186**: 429–458.
192. **Mui, M.M., Kamat, S.Y. and Elliott, W.H.** 1974. Bile acids XLII. Preparation of 5α -cholestane-26-oic acids from kryptogenin. Steroids **24**: 239–250.
193. **Murata, M., Kuramoto, T. and Hoshita, T.** 1978. Identification of bile alcohols in normal rabbit bile. Steroids **31**: 319–332.
194. **Nagata, K., Takakura, K., Asano, T., Seyama, Y., Hirota, H., Shigematsu, N., Shima, I., Kasama, T. and Shimizu, T.** 1992. Identification of 7α -hydroxy- 3 -oxo- 4 -cholestenoic acid in chronic subdural hematoma. Biochim. Biophys. Acta **1126**: 229–236.
195. **Nakatomi, F., Kihira, K., Kuramoto, T. and Hoshita, T.** 1985. Intestinal absorption and metabolism of norcholic acid in rats. J. Pharmacobio-Dyn. **8**: 557–563.
196. **Noll, B.W., Doisy, Jr., E.A. and Elliott, W.H.** 1973. Bile acids. XXXIX. Metabolism of 5α -cholestane- 3β , 26 -diol and 5α -cholestane- 3β , 7α , 26 -triol in the rat with a bile fistula. J. Lipid Res. **14**: 391–399.
197. **Noma, Y., Nama, Y., Kihira, K., Yasuhara, M., Kuramoto, T. and Hoshita, T.** 1976. Isolation of new C_{26} bile alcohols from bullfrog bile. Chem. Pharm. Bull. **24**: 2686–2691.
198. **Noma, Y., Une, M., Kihira, K., Yasuda, M., Kuramoto, T. and Hoshita, T.** 1980. Bile acids and bile alcohols of bullfrog. J. Lipid Res. **21**: 339–346.
199. **Noma, Y., Kihira, K., Kuramoto, T. and Hoshita, T.** 1988. Metabolism of C_{26} bile alcohols in the bullfrog, *Rana Catesbeiana*. Steroids **51**: 385–394.
200. **Oftebro, H., Bjorkhem, I., Skrede, S., Schreiner, A. and Pedersen, J.I.** 1980. Cerebrotendinous xanthomatosis. A defect in mitochondrial 26 -hydroxylation required for normal biosynthesis of cholic acid. J. Clin. Invest. **65**: 1418–1430.
201. **Okuda, K., Hoshita, T. and Kazuno, T.** 1962. Isolation of a new bile sterol, 3α , 7α , 12α , 25ζ , 26 -pentahydroxycoprostanone from toad bile. J. Biochem. **51**: 48–55.
202. **Okuda, K., Enomoto, S., Morimoto, K. and**

- Kazuno, T.** 1962. The isolation of a new bile sterol, 3 α , 7 α , 12 α -trihydroxy-24, 27-epoxycoprostanone, from sting-ray bile. *J. Biochem.* **51**: 441–442.
203. **Okuda, K. and Danielsson, H.** 1965. Synthesis and metabolism of 5 β -cholestane-3 α , 7 α , 12 α -triol-26-al. *Acta Chem. Scand.* **19**: 2160–2165.
204. **Okuda, K., Horning, M.G. and Horning, E.C.** 1972. Isolation of a new bile acid, 3 α , 7 α , 12 α -trihydroxy-5 α -cholestan-26-oic acid, from lizard bile. *J. Biochem.* **71**: 885–890.
205. **Parmentier, G.G., Janssen, G.A., Eggermont, E.A. and Eyssen, H.J.** 1979. C₂₇ bile acids in infants with coprostanic acidemia and occurrence of a 3 α , 7 α , 12 α -trihydroxy-5 β -C₂₉ dicarboxylic bile acid as a major component in their serum. *Eur. J. Biochem.* **102**: 173–183.
206. **Parmentier, G.G., Busson, R.H., Janssen, G.A., Mannaerts, G.P. and Eyssen, H.J.** 1993. Synthesis and 29-¹⁴C-labeling of 3 α , 7 α , 12 α -trihydroxy-27-carboxymethyl-5 β -cholestan-26-oic acid. A bile acid occurring in peroxisomal diseases. *Steroids* **58**: 351–356.
207. **Pearlman, W.H.** 1947. The preparation of C-27 steroids from bile acids. I. Coprostanetetrol-3(α), 7(α), 12(α), 25. *J. Am. Chem. Soc.* **69**: 1475–1476.
208. **Pellicciari, R., Natalini, B., Roda, A., Machado, M.I.L. and Maranozzi, M.** 1989. Preparation and physicochemical properties of natural (23R)-3 α , 7 α , 23- and (23R)-3 α , 12 α , 23-trihydroxylated bile acids and their (23S)-epimers. *J. Chem. Soc. Perkin Trans. I.* 1289–1296.
209. **Poulos, A. and Whiting, M.J.** 1985. Identification of 3 α , 7 α , 12 α -trihydroxy-5 β -cholestan-26-oic acid, an intermediate in cholic acid synthesis, in the plasma of patients with infantile Refsum's disease. *J. Inher. Metab. Dis.* **8**: 13–17.
210. **Pyrek, J.S., Sterzycki, R., Lester, R. and Adcock, E.** 1982. Constituents of human meconium: II. Identification of steroid acids with 21 and 22 carbon atoms. *Lipids* **17**: 241–249.
211. **Pyrek, J.S., Little, J.M. and Lester, R.** 1984. Detection of 3-hydroxy-etianic and 3-hydroxybis-norcholeanoic acids in human serum. *J. Lipid Res.* **25**: 1324–1329.
212. **Radominska-Pyre, A., Huynh, T., Lester, R. and Pyrek, J.St.** 1986. Preparation and characterization of 3-monohydroxylated bile acids of different side chain length and configuration at C-3. Novel approach to the synthesis of 24-norlithocholic acid. *J. Lipid Res.* **27**: 102–113.
213. **Riva, S., Bovara, R., Zetta, L., Pasta, P., Ottolina, G. and Carrea, G.** 1988. Enzymatic α/β inversion of C-3 hydroxyl of bile acids and study of the effects of organic solvents on reaction rates. *J. Org. Chem.* **53**: 88–92.
214. **Ruzicka, L., Plattner, Pl.A. and Heusser, H.** 1944. Über β' -[3 α , 7 α , 12 β -Trioxy-nor-cholanyl-(23)]- $\Delta^{\alpha',\beta'}$ -butenolid, ein Homologes der digitaloiden Aglucone. *Helv. Chim. Acta* **27**: 186–194.
215. **Sarel, S. and Yanuka, Y.** 1959. Preparation and degradation of 3 α -hydroxycholanic acid. *J. Org. Chem.* **24**: 2018–2019.
216. **Sasaki, T.** 1966. Comparative studies on the bile salts of fishes by thin layer chromatography. *J. Biochem.* **60**: 56–62.
217. **Schteingart, C.D. and Hofmann, A.F.** 1988. Synthesis of 24-nor-5 β -cholan-23-oic acid derivatives: a convenient and efficient one-carbon degradation of the side chain of natural bile acids. *J. Lipid Res.* **29**: 1387–1395.
218. **Schwenk, E., Riegel, B., Moffett, R.B. and Stahl, E.** 1943. The preparation of the homologs of 3-hydroxy-12-ketocholanic acid. *J. Am. Chem. Soc.* **65**: 549–551.
219. **Seno, H.** 1954. The partial synthesis of Trihydroxy-norsterocholane. *Proc. Japan Acad.* **30**: 887–890.
220. **Setchell, K.D.R., Bragetti, P., Zimmerman, N., Daugherty, C., Pelli, M.A., Vaccaro, R., Gentili, G., Distrutti, E., Dozzini, G., Morelli, A. and Clerici, C.** 1992. Oral bile acid treatment and the patient with Zellweger syndrome. *Hepatology* **15**: 198–207.
221. **Setoguchi, T., Salen, G., Tint, G.S. and Mosbach, E.H.** 1974. A biochemical abnormality in cerebrotendinous xanthomatosis. Implication of bile acid biosynthesis associated with incomplete degradation of the cholesterol side chain. *J. Clin. Invest.* **53**: 1393–1401.
222. **Shah, P.P., Staple, E., Shapiro, I.L. and Kritchevsky, D.** 1969. Isolation of 3 α , 7 α , 12 α -trihydroxycoprostanic acid from baboon bile. *Lipids* **4**: 82–83.
223. **Shalon, Y. and Elliott, W.H.** 1973. Improved aldehyde synthesis: Preparation of 3 α , 7 α , 12 α -triacetoxy-5 β -cholan-23-al with ruthenium tetroxide in neutral medium. *Syn. Commun.* **3**: 287–291.
224. **Shalon, Y. and Elliott, W.H.** 1976. Bile acids. LII. The synthesis of 24-nor-5 α -cholic acid and its 3 β -isomer. *Steroids* **28**: 655–667.
225. **Shefer, S., Dayal, B., Tint, G.S., Salen, G. and Mosbach, E.H.** 1975. Identification of pentahydroxy bile alcohols in cerebrotendinous xanthomatosis: characterization of 5 β -cholestane-3 α , 7 α , 12 α , 24 ξ , 25-pentol and 5 β -cholestane-3 α , 7 α , 12 α , 23 ξ , 25-pentol. *J. Lipid Res.* **16**: 280–286.
226. **Shimazu, K., Kuwabara, M., Yoshii, M., Kihira, K., Takeuchi, H., Nakano, I., Ozawa, S., Onuki, M., Hatta, Y. and Hoshita, T.** 1986. Bile alcohol profiles in bile, urine, and feces of a patient with cerebrotendinous xanthomatosis. *J. Biochem.* **99**: 477–483.
227. **Shimizu, K., Noda, F. and Yamasaki, K.** 1958. Preparation of 3 α , 7 α , 12 α -trihydroxy-24 α -oxocoprostanone and 3 α , 7 α , 12 α , 24 ξ -tetrahydroxycoprostanone. *J. Biochem.* **45**: 625–627.
228. **Shimizu, T. and Oda, T.** 1934. Untersuchung der Krotengalle. II. Trioxy-bufo-sterocholensaure C₂₈H₄₆O₅ aus Wintergalle. *Z. Physiol. Chem.* **227**: 74–83.
229. **Shimizu, T. and Kazuno, T.** 1936. Über die Konstitution der Trioxy-bufosterocholensaure und den systematischen Abbau der Cholsaure. V. Z. Physiol. Chem. **244**: 167–172.
230. **Shoda, J., Mahara, R., Osuga, T., Tohma, M., Ohnishi, S., Miyazaki, H., Tanaka, N. and Matsuzaki, Y.** 1988. Similarity of unusual bile acids in human umbilical cord blood and amniotic fluid from newborns and in sera and urine from

- adult patients with cholestatic liver diseases. 1988. *J. Lipid Res.* **29**: 847–858.
231. **Stellaard, F., Kleijer, W.J., Wanders, R.J.A., Schutgens, R.B.H. and Jakobs, C.** 1991. Bile acids in amniotic fluid: promising metabolites for the prenatal diagnosis of peroxisomal disorders. *J. Inher. Metab. Dis.* **14**: 353–356.
232. **Svoboda, J.A., Thompson, M.J. and Robbins, W.E.** 1968. 3 β -Hydroxy-24-norchol-5-en-23-oic acid--A new inhibitor of the Δ^{24} -sterol reductase enzyme system(s) in the tobacco hornworm, *Manduca sexta* (Johannson). *Steroids* **12**: 559–570.
233. **Takamori, M.** 1953. The synthesis of the derivatives of norcholic acid. *Hiroshima Igaku* (in Japanese) **6**: 320–322.
234. **Takamori, M.** 1953. The synthesis of the derivatives of bisnorcholic acid. *Hiroshima Igaku* (in Japanese) **6**: 323–325.
235. **Takeda, K., Komeno, T. and Igarashi, K.** 1954. Bile acids and steroids. VI. On the saponification of 7-oxo-6-bromocholanic acid. *Pharm. Bull.* **2**: 352–358.
236. **Thomassen, P.A.** 1979. Urinary bile acids during development of recurrent cholestasis of pregnancy. *Eur. J. Clin. Invest.* **9**: 417–423.
237. **Tint, G.S., Dayal, B., Batta, A.K., Shefer, S., Joosten, T., McNease, L. and Salen, G.** 1980. Biliary bile acids, bile alcohols, and sterols of *Alligator mississippiensis*. *J. Lipid Res.* **21**: 110–117.
238. **Tint, G.S., Dayal, B., Batta, A.K., Shefer, S., Joosten, T., McNease, L. and Salen, G.** 1981. The fecal bile acids and sterols of *Alligator mississippiensis*. *Gastroenterology* **80**: 114–119.
239. **Une, M., Kihira, K., Kuramoto, T. and Hoshita, T.** 1978. Two new bile alcohols, 3-epimyxinol and 3-epi-16-deoxymyxinol from the hagfish, *Heptatretus burgeri*. *Tetrahedron Lett.* 2527–2530.
240. **Une, M., Matsumoto, N., Kihira, K., Yasuhara, M., Kuramoto, T. and Hoshita, T.** 1980. Bile salts of frogs: a new higher bile acid, 3 α , 7 α , 12 α , 26-tetrahydroxy-5 β -cholestanoic acid from the bile of *Rana planctyi*. *J. Lipid Res.* **21**: 269–276.
241. **Une, M., Nagai, F., Kihira, K., Kuramoto, T. and Hoshita, T.** 1983. Synthesis of four diastereoisomers at carbons 24 and 25 of 3 α , 7 α , 12 α , 24-tetrahydroxy-5 β -cholestane-26-oic acid, intermediates of bile acid biosynthesis. *J. Lipid Res.* **24**: 924–929.
242. **Une, M., Kuramoto, T. and Hoshita, T.** 1983. The minor bile acids of the toad, *Bufo vulgaris formosus*. *J. Lipid Res.* **24**: 1468–1474.
243. **Une, M., Nagai, F. and Hoshita, T.** 1983. High-performance liquid chromatographic separation of higher bile acids. *J. Chromatogr.* **257**: 411–415.
244. **Une, M., Morigami, I., Kihira, K. and Hoshita, T.** 1984. Stereospecific formation of (24E)-3 α , 7 α , 12 α -trihydroxy-5 β -cholest-24-en-26-oic acid and (24R, 25S)-3 α , 7 α , 12 α , 24-tetrahydroxy-5 β -cholestane-26-oic acid from either (25R)- or (25S)-3 α , 7 α , 12 α -trihydroxy-5 β -cholestane-26-oic acid by rat liver homogenate. *J. Biochem.* **96**: 1103–1107.
245. **Une, M., Shinonaga, Y., Matoba, N., Kuroki, S., Kihira, K. and Hoshita, T.** 1986. Identification of new bile alcohols, 5 β -cholestane-3 α , 7 α , 24, 26-tetrol, 5 β -cholestane-3 α , 7 α , 25, 26-tetrol, and 5 β -cholestane-3 α , 7 α , 26, 27-tetrol in human gallbladder bile. *J. Lipid Res.* **27**: 1318–1323.
246. **Une, M., Tazawa, Y., Tada, K. and Hoshita, T.** 1987. Occurrence of both (25R)- and (25S)-3 α , 7 α , 12 α -trihydroxy-5 β -cholestanoic acids in urine from an infant with Zellweger's syndrome. *J. Biochem.* **102**: 1525–1530.
247. **Une, M., Tsujimura, K., Kihira, K. and Hoshita, T.** 1989. Identification of (22R)-3 α , 7 α , 12 α , 22- and (23R)-3 α , 7 α , 12 α , 23-tetrahydroxy-5 β -cholestanoic acids in urine from a patient with Zellweger's syndrome. *J. Lipid Res.* **30**: 541–547.
248. **Une, M., Kisaka, N., Yoshii, M. and Hoshita, T.** 1989. Identification of 3 α , 6 α , 7 α , 12 α -tetrahydroxy-5 β -cholestanoic acid in Zellweger's syndrome. *J. Biochem.* **106**: 501–504.
249. **Wanders, R.J.A., Casteels, M., Mannaerts, G.P., Van Roermund, C.W.T., Schutgens, R.B.H., Kozich, V., Zeman, J. and Hynek, J.** 1991. Accumulation and impaired in vivo metabolism of di- and trihydroxycholestanoic acid in two patients. *Clin. Chim. Acta* **202**: 123–132.
250. **Wessely, F. and Svoboda, W.** 1951. Über die Reduktion Gallensauren und ihrer Amide mit Lithiumaluminimumhydrid. *Monatshr. Chem.* **82**: 437–442.
251. **Wolthers, B.G., Volmer, M., van der Molen, J., Koopman, B.J., de Jager, A.E.J. and Waterreus, R.J.** 1983. Diagnosis of cerebrotendinous xanthomatosis (CTX) and effect of chenodeoxycholic acid therapy by analysis of urine using capillary gas chromatography. *Clin. Chim. Acta* **131**: 53–65.
252. **Yamasaki, K. and Yuuki, M.** 1936. Über die Gallensaure der Alligatorschildkroten. *Z. Physiol. Chem.* **244**: 173–180.
253. **Yanuka, Y., Katz, R. and Sarel, S.** 1968. Bile acid chemistry. III. Stepwise side-chain shortening by way of sodium per-iodate oxidation of α -hydroxy bile acids into corresponding aldehydes. *Tetrahedron Lett.* 1725–1728.
254. **Yashima, H.** 1963. Synthesis and metabolism of cholic aldehyde. *J. Biochem.* **54**: 47–50.
255. **Yasuhara, M., Kuramoto, T. and Hoshita, T.** 1978. Identification of 5 β -cholestane-3 α , 7 α , 12 α , 23 β -tetrol, 5 β -cholestane-3 α , 7 α , 12 α , 24 α -tetrol, and 5 β -cholestane-3 α , 7 α , 12 α , 24 β -tetrol in cerebrotendinous xanthomatosis. *Steroids* **31**: 333–345.
256. **Yoshii, M., Une, M., Kihira, K., Kuramoto, T. and Hoshita, T.** 1989. Synthesis of 5 β -cholestane-3 α , 6 β , 7 α , 25, 26-pentol and identification of a novel bile alcohol, α -trichechol, present in the West Indian manatee bile. *Chem. Pharm. Bull.* **37**: 1852–1854.
257. **Yoshii, M., Une, M., Kihira, K., Kuramoto, T., Akizawa, T., Yoshioka, M., Butler, Jr. V.P. and Hoshita, T.** 1994. Bile salts of the toad, *Bufo marinus*, characterization of a new unsaturated higher bile acid, 3 α , 7 α , 12 α , 26-tetrahydroxy-5 β -cholest-23-en-27-oic acid. *J. Lipid Res.* **35**: in press.