

Immunohistochemical Localization of Apolipoprotein E in Renal Amyloidosis

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ABSTRACT

To clarify the role of apolipoprotein E (apo E) in the formation of amyloid deposits, we examined specimens from 11 patients with renal amyloidosis who underwent renal biopsy by an immunohistochemical method using a monoclonal antibody (murine IgG₁). Apo E was distributed in the amyloid deposits of all patients in a pattern similar to that obtained with Congo red staining. Strong positive staining for apo E was found on the amyloid deposits in the glomeruli. These results suggest that apo E is a common constituent of amyloid fibrils and that it may be a useful marker for immunohistochemical studies of systemic amyloidosis including renal amyloidosis.

Key words: *Apolipoprotein E, Pathological molecular chaperones, Amyloid deposits*

Amyloidosis is a systemic disease which shows various symptoms because of the distribution of amyloid in various tissues^{3,5,6}. Of the organs affected, the kidney is injured the most frequently which is why this condition is often called renal amyloidosis. Nephrotic syndrome is the most frequent condition clinically and the prognosis is generally poor because it ultimately leads to renal failure^{4,9}.

Recently, it was reported that amyloid deposits stained positively for apolipoprotein E (apo E) and that the staining pattern was similar to that obtained with Congo red staining in tissues from patients with systemic amyloidosis. It was also suggested that fragments of apo E are a common constituent of amyloid fibrils of diverse origin, and that it may be a new marker for immunohistochemical studies of systemic amyloidosis¹⁶.

In this study, we describe 11 patients with renal amyloidosis who were diagnosed in our hospital and examined immunohistochemically using a monoclonal antibody (anti-apo E antibody).

MATERIALS AND METHODS

Tissue samples and classifications:

Eleven patients who underwent renal biopsy at our department between 1983 and 1994 were included in this study. These patients were initially diagnosed with renal amyloidosis by Congo red staining. There were 2 males and 9 females with a mean age of 56.5 ± 10.2 years. Of these 11 patients, the first symptoms were proteinuria and edema in 8 patients, edema in 2, and renal dysfunction in 1, while 9 patients out of the 11

showed nephrotic syndrome. Cases 1-6 were of primary amyloidosis and cases 7-11 were of secondary amyloidosis (Table 1). Chemical types of amyloid were immunohistochemically defined using IgG antibody against amyloid A (AA) component (Biomedica Corp., USA).

Immunohistochemistry:

Apo E was stained as follows: paraffin-embedded sections were deparaffinized, and antigen was unmasked by treatment with 0.01% trypsin diluted with 0.06 M phosphate-buffered saline (pH 7.2) for 30 minutes at room temperature and incubated with 3% hydrogen peroxide in methanol for 20 minutes at room temperature to block intrinsic peroxidase activity. They were incubated in a primary antibody (murine IgG₁, MON 5029, SANBIO BV, Holland) overnight at 4°C and stained immunohistochemically using an avidin-peroxidase/biotin staining kit (Vector Laboratories, USA). The immunoproducts were visualized by the application of diaminobenzidine, and the nuclei were stained with methyl green.

RESULTS

Clinical findings at biopsy:

Total daily urinary protein was 0.3-9.4 (mean \pm SD; 5.5 ± 3.2) g/day, serum total protein was 4.7-7.4 (5.5 ± 0.7) g/dl, and serum albumin was 1.4-3.9 (2.5 ± 0.9) g/dl. Serum creatinine was 0.49-3.27 (1.43 ± 1.06) mg/dl, and creatinine clearance was 14.8-93.4 (51.9 ± 28.1) ml/min (Table 2). Based on these findings, almost all patients with amyloidosis had nephrotic syndrome

Table 1. Background characteristics of patients with renal amyloidosis

Case	Name	Age	Sex	First symptoms	Nephrotic syndrome	Diagnosis
1	S.M.	55	female	proteinuria, edema	+	primary
2	H.H.	59	female	edema	-	primary
3	T.Y.	67	female	proteinuria, edema	+	primary
4	M.N.	58	female	proteinuria, edema	+	primary
5	K.M.	49	male	proteinuria, edema	+	primary
6	K.I.	69	female	edema	+	primary
7	H.Y.	32	male	proteinuria, edema	+	secondary (RA)
8	C.O.	66	female	proteinuria, edema	+	secondary (RA)
9	T.O.	53	female	renal dysfunction	-	secondary (RA)
10	O.M.	59	female	proteinuria, edema	+	secondary (RA)
11	S.H.	55	female	proteinuria, edema	+	secondary (RA)

Table 2. Clinical findings at biopsy

Case	Name	TUP (g/day)	TP (g/dl)	Alb. (g/dl)	Cr (mg/dl)	Ccr (ml/min)
1	S.M.	5.8	5.2	2.7	3.27	15.2
2	H.H.	2.1	5.9	3.8	0.90	42.9
3	T.Y.	5.9	4.7	1.4	0.49	53.2
4	M.N.	9.4	5.4	2.5	0.70	66.9
5	K.M.	7.2	4.9	2.2	1.39	93.4
6	K.I.	9.0	4.9	2.1	0.66	50.6
7	H.Y.	7.4	5.1	2.9	3.20	84.5
8	C.O.	8.4	5.3	2.8	0.80	56.3
9	T.O.	0.3	7.4	3.9	1.10	14.8
10	O.M.	1.2	5.5	3.0	0.62	78.6
11	S.H.	3.6	6.3	2.6	2.54	15.5

TUP, total urinary protein; TP, total protein; alb., albumin; Cr, creatinine; Ccr, creatinine clearance.

and exhibited renal dysfunction.

Distribution of amyloid deposits by Congo red staining:

Congo red staining showed that amyloid deposits were distributed in the glomeruli of all patients, and 10 of 11 patients were positive in the vascular walls. Moreover, the epithelial cells in the tubules and interstitial area were positive in all patients except for cases 1 and 3.

Chemical types of amyloid using by AA peptides:

Five patients showed positive staining for AA peptides and 6 were negative. The five patients positive for AA peptides were all diagnosed with rheumatic arthritis (RA) associated with secondary amyloidosis.

Distribution of apo E:

Apo E was distributed in the amyloid deposits of all patients in a pattern similar to that obtained with Congo red staining. Strong positive staining for apo E was found on the amyloid deposits in the glomeruli (Fig. 1).

DISCUSSION

Apo E is a plasma protein of 299 amino acids with a relative molecular mass of 34,000, which is involved in cholesterol transport¹⁴. Apo E has a wide variety of functions including a structural role in a number of lipoprotein particles, as well as that of regulating their metabolism. Moreover, apo E is produced by most organs (the largest amount of apo E mRNA is found in the brain, second only to the liver), and its synthesis is increased following injury and is implicated in the

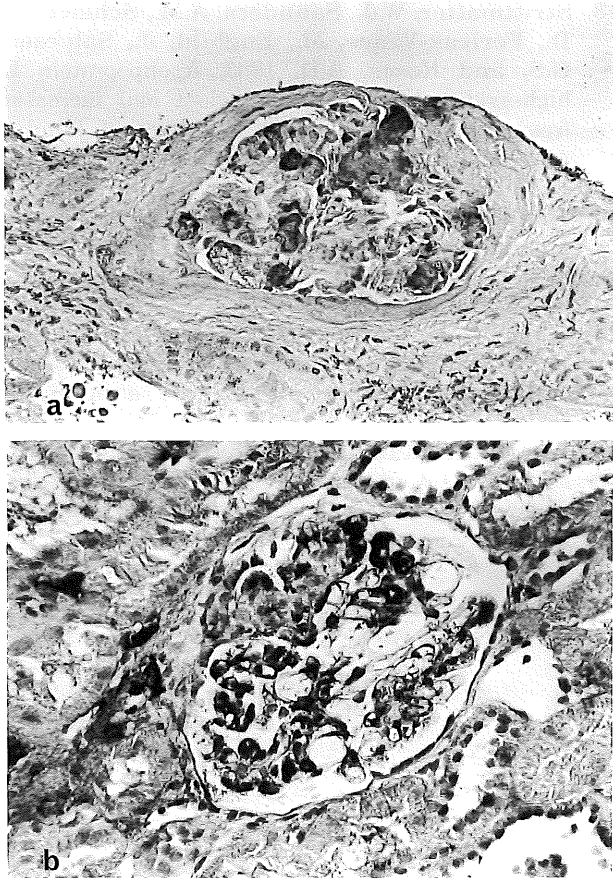


Fig. 1. The distribution of apolipoprotein E: strong staining for apo E was observed on amyloid deposits in the glomeruli. ($\times 350$)

a) Immunohistochemistry with Apo E antibody for a biopsy specimen with the patient in primary amyloidosis.

b) Immunohistochemistry with Apo E antibody for a biopsy specimen with the patient in secondary amyloidosis.

growth and repair of the nervous system during development or after injury^{13,14}). Recently, it was reported that apo E may be an essential cofactor for amyloid fibril formation and that it plays an important role in the distribution of amyloid deposits in various tissues^{1,7,11,13,16}). This idea is supported by recent immunohistochemical and molecular biological findings which showed positive immunostaining of amyloid deposits or expression of apo E mRNA in Alzheimer's disease and Down's syndrome associated amyloid, primary systemic amyloidosis and secondary amyloidosis coexistent with dialysis or RA^{1,7,11,13,15,16}). In this study, we examined 11 patients with renal amyloidosis immunohistochemically. The present results provide further confirmation that apo E is distributed in the glomeruli, vascular wall and/or the interstitial area, and suggest that apo E staining correlates highly with the presence of amyloid deposits. It is speculated that amyloid-

associated proteins such as P component, glucosaminoglycans and α_1 -anti-chymotrypsin reduce the solubility and stabilize the β -pleated structure in amyloid fibrils or their precursor proteins, and so play an important role in the formation of amyloid deposits⁸). Our results suggest that apo E, in addition to the above proteins, may function as 'pathological molecular chaperones'¹⁴). Further studies using models of amyloid formation or in situ hybridization are needed to clarify the interactions between pathological chaperones and amyloid deposits.

At present, patients with renal amyloidosis have been treated by controlling precursor proteins or by the prevention and removal of amyloid deposits^{2,10,12}), but these therapies are not very effective. If the functions of pathological chaperones such as apo E, P component and glucosaminoglycans can be clarified, anti-pathological chaperones may be of therapeutic benefit.

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REFERENCES

1. Allsop, D., Landon, M., Kidd, M., Lowe, J.S., Reynolds, G.P. and Gardner, A. 1986. Monoclonal antibodies raised against a subsequence of senile plaque core protein react with plaque cores, plaque periphery and cerebrovascular amyloid in Alzheimer's disease. *Neurosci. Lett.* **68**: 252-256.
2. Brandwein, S.R., Sipe, J.D. and Cohen, A.S. 1994. Combined treatment with terbutaline and aminophylline inhibits experimental amyloidosis in mice. *Arthritis Rheum.* **37**: 1757-1760.
3. Cohen, A.S. and Calkins, E. 1959. Electron microscopic observations on a fibrous component in amyloid of diverse origins. *Nature* **183**: 1202-1203.
4. Çolakoglu, M., Sungur, C., Sungur, A., Akpolat, T., Kansu, E., Yasavul, ü., Turgan, Ç. and Caglar, S. 1995. Pattern of proteinuria in patients with renal amyloidosis secondary to familial mediterranean fever. *Nephron* **69**: 124.
5. Hermansen, L.F., Bergman, T., Jörnvall, H., Husby, G., Ranløv, I. and Sletten, K. 1995. Purification and characterization of amyloid-related transthyretin associated with familial amyloidotic cardiomyopathy. *Eur. J. Biochem.* **227**: 772-779.
6. Kyle, R.A., Greipp, P.R. and O'Fallon, W.M. 1986. Primary systemic amyloidosis: multivariate analysis for prognostic factors in 168 cases. *Blood* **68**: 220-224.
7. Livneh, A., Zemer, D., Langevitz, P., Laor, A., Sohar, E. and Pras, M. 1994. Colchicine treatment of AA amyloidosis of familial mediterranean fever. *Arthritis Rheum.* **37**: 1804-1811.
8. Pasternack, J.M., Abraham, C.R., Dyke, B.J.V., Potter, H. and Younkin, S.G. 1989. Astrocytes in Alzheimer's disease gray matter express α_1 -anti-chymotrypsin mRNA. *Am. J. Pathol.* **135**: 827-834.
9. Quinton, R., Siersema, P.D., Michiels, J.J. and

- ten Kate, F.J.W.** 1992. Renal AA amyloidosis in a patient with Bence Jones proteinuria and ankylosing spondylitis. *J. Clin. Pathol.* **45**: 934–936.
10. **Sakhuja, V. and Chugh, K.S.** 1988. Renal amyloidosis-is it preventable?. *Int. J. Artif. Organs* **11**: 63–64.
11. **Saunders, A.M., Schmader, K., Breitner, J.C.S., Benson, M.D., Brown, W.T., Goldfarb, L., Goldgaber, D., Manwaring, M.G., Szymanski, M.H., McCown, N., Dole, K.C., Schmechel, D.E., Strittmatter, W.J., Pericak-Vance, M.A. and Roses, A.D.** 1993. Apolipoprotein E ϵ 4 allele distributions in late-onset Alzheimer's disease and in other amyloid-forming diseases. *Lancet* **342**: 710–711.
12. **Spiegel, D.M., Costante, N., Janiga, A.M., Haas, M. and Soltani, K.** 1992. Deposition and removal of cutaneous beta₂-microglobulin. *Am. J. Nephrol.* **12**: 330–335.
13. **Strittmatter, W.J., Saunders, A.M., Schmechel, D., Pericak-Vance, M., Enghild, J., Salvesen, G.S. and Roses, A.D.** 1993. Apolipoprotein E: high-avidity binding to β -amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc. Natl. Acad. Sci. USA* **90**: 1977–1981.
14. **Wisniewski, T. and Frangione, B.** 1992. Apolipoprotein E: a pathological chaperone protein in patients with cerebral and systemic amyloid. *Neurosci. Lett.* **135**: 235–238.
15. **Wong, C.W., Quaranta, V. and Glenner, G.G.** 1985. Neuritic plaques and cerebrovascular amyloid in Alzheimer disease are antigenically related. *Proc. Natl. Acad. Sci. USA* **82**: 8729–8732.
16. **Yamada, T., Kakihara, T., Gejyo, F. and Okada, M.** 1994. A monoclonal antibody recognizing apolipoprotein E peptides in systemic amyloid deposits. *Ann. Clin. Lab. Sci.* **24**: 243–249.