Satoshi FUJII, Yukio TAKESHIMA, Koji ARIHIRO, Mayumi KANEKO and Kouki INAI 2nd Department of Pathology, Hiroshima University School of Medicine, 1–2–3 Kasumi, Minami-ku, Hiroshima 734–8551, JAPAN

# ABSTRACT

We examined breast cancers from 67 female patients to ascertain the possible correlation between RER or LOH status, age and bilaterality using eight microsatellite markers on chromosomes 2p, 3p, 16q, 17p and 17q. The frequencies of RER in young patients (25–35 years old), patients with double primary disease (43–77 years old) and patients with contralateral metastases (46–72 years old) were 35%, 63%, and 80%, respectively, while that in elderly patients (60–81 years old) was 0%. In contrast, there were no statistically significant differences in the frequency of LOH between these groups. Our results suggest that RER might play an important role in the occurrence of breast cancer at a younger age and in bilateral breast cancer.

Key words: Replication error (RER), Loss of heterozygosity (LOH), Young, Bilateral breast cancer

Epidemiological studies have indicated that a positive family history, obesity and hormonal imbalance are etiological risk factors for breast cancer, and it has recently been suggested that somatic or inherited genetic alterations play an important role in the occurrence, development and progression of this disease <sup>5-7,13,17,21,28,37,38,41,43,45,46,51,52,57,59,62,64</sup>. There have been now many reports of genetic alterations in breast cancer. However, it remains unclear whether these are critical events in the early stages of carcinogenesis. The correlation between risk factors and genetic alterations is also unclear.

In contrast, a distinctive type of genetic alteration based on misalignment mutagenesis has been reported in colorectal and gastric cancer. This type of mutagenesis involves microsatellites which are defined as tandem arrays of short, simple nucleotide sequences dispersed within the human genome. It is now believed that such repeat sequences indicate a predisposition to mutation and that alterations in microsatellites reflect genuine genetic instability, although it is still unknown whether this genetic instability has a general effect on activating oncogenes or inactivating tumor suppressor genes. The phenotype associated with genetic instability has been termed "microsatellite instability" or "replication error (RER)".

Colorectal cancer was the first type of human malignancy in which RER was detected. RER has been very frequently demonstrated in patients with hereditary non-polyposis colorectal cancer (HNPCC). Moreover, it has been found that these patients have germline mutations in the known mismatch repair genes<sup>8,42)</sup>. Also, non-HNPCC colorectal cancers in patients aged 35 years or younger exhibit a higher prevalence of RER than those in elderly patients<sup>32)</sup>. RER has subsequently been found in many tumor types including breast cancer<sup>11,18,48)</sup>. If similar abnormalities of the mismatch repair gene system affect the genesis of breast cancer, RER might occur in the early stages of breast cancer in young patients. Moreover, bilateral breast cancer may have a background of RER because patients with multiple primary cancers have frequently been reported to show RER<sup>20,39)</sup>.

Loss of heterozygosity (LOH) at microsatellite loci on various chromosomes is a feature of microsatellite instability, and various rates of LOH have been reported in breast cancer. However, there are few reports concerning the correlation between LOH status and the age of the patients or bilaterality.

Therefore, we examined the prevalence of RER and LOH in breast cancer, in relation to the age of the patients and bilaterality, to elucidate the role of microsatellite instability in the tumorigenesis of breast cancer.

# MATERIALS AND METHODS Patient selection

The correlation between RER and LOH status, age and bilaterality was evaluated in 67 female patients with breast cancer, identified from our surgical files over the period 1986 to 1996. We classified the patients into five groups: young (25–35 years old, 20 patients, Group I; 36–39

Group	No. of patient	Age <sup>a</sup>	Size <sup>b</sup> (range)		Histological type <sup>c</sup>			$\mathrm{LN}^{\mathrm{d}}$		
				NIDC	IDC	ILC	Special type <sup><math>e</math></sup>	(+)	(_)	
Young patients										
Group I	20	31	2.4(0.7-6.0)	0	19	0	1	9	11	
Group II	18	37	3.0(0.9-7.0)	3	14	0	1	9	9	
Elderly patients										
Group III	16	69	2.4(1.0-6.0)	1	14	1	0	2	14	
Bilateral cancer										
Group IV	8	60	2.8(0.3-10.5)	1	13	2	0	3	5	
Group V	5	61	4.4(0.8-15.0)	0	5	0	0	5	0	

**Table 1.** Clinicopathological Profiles of Patients with Breast Cancer

NIDC: nonivasive ductal carcinoma; IDC: invasive ductal carcinoma; ILC: invasive lobular carcinoma.

"Age, average age of patients; "Size, average maximum tumor size (cm); "Pathological classification of breast cancer was confirmed according to the general rules for clinicopathological records of breast cancer of the Japan Breast Cancer Society; "LN, No. of patients with lymph node metastasis; "Special type, other histological types."

years old, 18 patients, GroupII), elderly (60-81 years old, 16 patients, Group III) and those with bilateral disease (43-77 years old, 13 patients). The patients with bilateral breast cancer included 8 with independent primary carcinomas of both breasts (double primary disease, 43-77 years old, GroupIV) and 5 with primary carcinoma of one breast which had metastasized to the contralateral breast (46-72 years old, GroupV). Of the 16 elderly patients, 14 had no lymph node metastasis. Of the 20 patients in Group I, 9 had lymph node metastases and of the 18 patients in Group II, 9 had lymph node metastases. The clinicopathological profiles of the patients are summarized in Table 1. Patients with family history of breast cancers were not included in the present study.

Sections of paraffin-embedded cancerous tissues were stained with hematoxylin and eosin and examined routinely under the microscope according to the guidelines of the Japan Breast Cancer Society.

## **DNA** extraction

From tissues fixed in 10% formalin and embedded in paraffin,  $10\mu$ m-thick unstained sections were cut. After dewaxation with xylene and 100% ethanol, microscope slides of the cancerous area were prepared by scraping off only cancerous cells with sterile needles. Slides from areas with a high proportion (>50%) of cancerous cells were prepared for microscopic DNA analysis. Using the same method, we extracted matched normal DNA from lymph node tissue free of metastases. Genomic DNA was prepared by the standard method<sup>53</sup>.

#### Polymerase chain reaction-based microsatellite analysis

Sixty-seven matched DNA pairs obtained from normal and cancerous tissues were examined for RER and LOH at eight microsatellite loci by polymerase chain reaction (PCR) amplification. Eight primers were used to target and examine two microsatellite loci on chromosome 16 (D16S301 and D16S303)<sup>10,55)</sup>, four on chromosome 17 (D17S796, TP53, D17S855 and D17S579)<sup>3,23)</sup>, one locus on chromosome 2  $(D2S136)^{58)}$  and one on chromosome 3 (D3S1067)<sup>24)</sup>. Each PCR was performed in  $25\mu$ l reaction mixture containing 200-500ng DNA, 2.5µl 10×buffer (500mM KCl, Tris-HCl 100mM (pH8.8),  $15 \mathrm{mM}$ MgCl<sub>2</sub>, 1%Triton), 0.05mM dNTP, 1.25 units Taq DNA polymerase, 0.05nM of the counterpart of each primer and  $2\mu Ci$  [ $\alpha$ -<sup>32</sup>P-dCTP]. After heating at 95°C for 8 min, 35 PCR cycles comprising 1 min at each annealing temperature, 2 min at 72°C for strand elongation and 40s at 95°C for denaturing were performed. A final elongation was performed over 7 min at 72°C. The reaction product  $(4\mu l)$  was then denatured and electrophoresed in 6% denaturing polyacrylamide gel containing urea. After electrophoresis, the gel was fixed on paper, dried and exposed to X-ray film for 24-48h. If a shift and/or gain of electrophoretic bands was observed with the DNA allele from the cancer, that cancer was categorized as showing RER. We defined a patient whose cancer showed RER using at least one marker, as a patient with RER. If the signal intensity of one allele in the cancer was reduced to less than 50% of the normal intensity observed among informative cases, that patient was categorized as showing LOH.

## Statistical analysis

Correlations between the RER or LOH status and clinicopathological parameters (histological type, tumor size, lymph node metastasis and stage) were analyzed using chi-square method or Fisher's exact test.

#### RESULTS

A summary of the RER and LOH status of the young patients are shown in Table 2 and 3, and the RER and LOH status of the patients with bilateral disease are summarized in Table 4. Typical electrophoretic photographs are shown in

Table 2. The Frequency of RER in Young Patients

Group	n	Microsatellite markers									
	11	D16S301	D16S303	D17S796	TP53	D17S855	D17S579	D2S136	D3S1067	. 10041	
I	20	$1/20(5)^{a}$	0/20(0)	2/20(10)	1/20(5)	1/20(5)	4/20(20)	0/20(0)	0/20(0)	7/20(35) -	
II	18	0/18(0)	0/18(0)	0/18(0)	0/18(0)	1/18(6)	2/18(11)	0/18(0)	0/18(0)	2/18(11)	
III	16	0/16(0)	0/16(0)	0/16(0)	0/16(0)	0/16(0)	0/16(0)	0/16(0)	0/16(0)	0/16(0) —	

<sup>a</sup>Number of cases with RER/total cases (%). <sup>b</sup>p<0.05.

Table 3.	The	Frequency	of LOH	in Y	Young Patients
					<i>(</i> )

Group	n	Microsatellite markers									
		D16S301	D16S303	D17S796	TP53	D17S855	D17S579	D2S136	D3S1067	iotai	
I	20	$1/20(5)^{a}$	0/20(0)	2/17(12)	1/20(5)	1/20(5)	1/18(6)	0/20(0)	1/18(6)	7/20(35) —	
II	18	2/17(12)	0/18(0)	1/13(8)	2/18(11)	1/18(6)	0/17(0)	0/18(0)	2/17(12)	5/18(28) -	
III	16	0/16(0)	0/16(0)	0/16(0)	2/14(14)	2/16(13)	1/16(6)	2/16(13)	0/16(0)	5/16(31)	

<sup>a</sup>Number of cases with LOH/number of infomative cases (%). <sup>b</sup>not significant.





Fig. 1. Replication Error and Loss of Heterozygosity in Breast Cancer Specimens from Young Patients

For each patient, lane N shows DNA from non-cancerous tissue and lane T shows DNA from the breast cancer. The tumors from patients 31, 32, 38, 39 and 40 (upper row) exhibit RER, while those from patients 31 and 33 (lower row) exhibit LOH. The shift and loss of band in electrophoretic mobility was reproducible in replicate experiments.



Patients with double primary disease Patients with contralateral metastases

# **Fig. 2.** Replication Error in Breast Cancer Specimens from Patients with Bilateral Disease

For each patient, lane N shows DNA from non-cancerous tissue, and lanes R and L DNA from the tumors in the right and left breasts. Patients 1, 2, 3 and 5 had double primary disease. In these patients, the tumor in one breast exhibits RER at D16S301, whereas both tumors in patients with metastasis to the contralateral breast (patients 6, 7, 9 and 10) exhibit RER at TP53.



Fig. 3. Percentage of RER and LOH in Breast Cancer Specimens from Different Groups

Tab	le 4. Result of Mici	rosatellite Assa	y in the Patie	ents with Bila	teral Breast	Cancer
Patients with Double	Primary Desease					

Patients'	Age	Histolgy	LN	Size	Stage	Microsatellite marker							
No.						D16S301	D16S303	D17S796	TP53	D17S855	D17S579	D2S136	D3S1067
1R	63	IDC	(+)	10.5	III	he	he	he	he	he	he	he	he
1L	63	IDC	(+)	1.7	$\mathbf{III}$	RER	he	he	he	he	he	he	he
$2\mathrm{R}$	57	IDC	(_)	4.5	Ι	he	he	he	he	he	he	he	he
2L	57	IDC	(-)	1.7	Ι	RER	he	LOH	LOH	he	he	he	he
3R	69	IDC	(+)	NA	NA	he	he	LOH	he	he	he	ho	he
3L	69	IDC	()	NA	NA	RER	he	he	he	he	$\mathbf{he}$	ho	he
$4\mathrm{R}$	43	IDC	(_)	NA	NA	he	he	he	he	he	he	he	he
4L	43	IDC	(_)	2.6	Ι	RER	he	he	he	he	$\mathbf{he}$	he	he
$5\mathrm{R}$	74	IDC	()	1.5	Ι	RER	he	he	he	he	he	LOH	he
5L	74	ILC	()	2.3	Ι	he	he	he	he	he	$\mathbf{he}$	he	he
11R	47	ILC	(_)	1.2	Ι	he	LOH	ho	he	he	he	he	he
11L	44	IDC	(_)	1.2	Ι	LOH	LOH	ho	he	he	$\mathbf{he}$	he	he
12R	54	IDC	(-)	2.4	Ι	he	he	he	he	he	$\mathbf{he}$	he	he
12L	54	IDC	()	0.3	Ι	he	he	he	he	he	he	he	he
13R	77	NIDC	()	5	Ι	he	he	LOH	LOH	he	he	he	he
13L	77	IDC	(+)	0.9	Ι	he	he	he	he	he	he	he	he

Patients with Contralateral Metastases

Patients'	Age	Histolgy	LN	Size	Stage	Microsatellite marker							
No.						D16S301	D16S303	D17S796	TP53	D17S855	D17S579	D2S136	D3S1067
6R	48	IDC	NA	3	NA	he	he	he	RER	he	he	he	he
6L	46	IDC	(+)	NA	NA	he	he	he	RER	he	he	he	he
7R	54	IDC	NA	3	NA	he	he	he	RER	he	he	he	he
7L	53	IDC	(+)	15	II	he	he	he	RER	he	he	he	he
8R	64	IDC	(+)	<b>5</b>	$\Pi$	he	he	he	he	he	LOH	he	he
8L	67	IDC	NA	1.2	NA	he	he	LOH	he	he	LOH	LOH	he
9R	72	IDC	NA	0.8	NA	he	he	he	RER	he	he	ho	he
9L	72	IDC	(—)	4.5	Ι	he	he	LOH	RER	he	he	ho	he
10R	64	IDC	(+)	2.5	III	he	$\mathbf{he}$	he	RER	he	he	ho	he
10L	66	IDC	(+)	NA	NA	he	he	he	RER	he	he	ho	he

LN: lymph node metastasis; he: non-LOH; RER: replication error; LOH: loss of heterozygosity; ho: not informative; NA: not available; IDC: invasive ductal carcinoma; ILC: invasive lobular carcinoma.

Fig. 1 and 2. The percentages of RER and LOH observed in each group are shown in Fig. 3. The percentage of RER in young patients was considerably higher than that in elderly patients (Fig. 3). The difference in RER between 25 to 35 year-old and elderly patients (Group I and III) was statistically significant (p<0.05). RER was observed more frequently in both patients with double primary disease and those with contralateral metastases (Group IV and V) than in elderly patients (Group III) (p<0.01).

In contrast, the percentage of LOH in young patients was similar to that in elderly ones (Fig. 3). A tendency towards a higher percentage of LOH was observed in patients with double primary disease, but there were no significant differences in the percentage of LOH between the groups.

The microsatellite markers D2S136 and D3S1067 were used to identify changes in the MSH2 and MLH1 genes in this study. There were few patients with alterations at these microsatellite loci (Table 3 and 4) (1 in Group I at D3S1067, 2 in Group II at D3S1067, 2 in Group II at D3S1067, 2 in Group II at D2S136, 1 in Group IV at D2S136 and 1 in Group V at D2S136), that is, there was no striking association between the presence of RER and alterations at microsatellite loci on the MSH2 and MLH1 genes.

Patients with double primary disease showed a predisposition towards alterations at D16S301 and D16S303 on chromosome 16q more frequently. In 5 of these 8 the tumor on one side exhibited RER at D16S301 (Table 4). On the other hand, patients with metastatic tumors exhibited a predisposition towards more frequent alterations at D17S796 and TP53 on chromosome 17p (Table 4). In 4 of 5 patients with contralateral metastases, both tumors showed RER at TP53.

There was a few patients with genetic alterations at microsatellite locus (D17S855) on the BRCA1 gene (Table 2, 3 and 4) (2 in Group I, 2 in Group II, 2 in Group III, 2 in Group III and none in Groups IV or V).

No significant correlation between RER or LOH and any clinicopathological parameter was observed in this study.

#### DISCUSSION

Cancers with RER can be classified into three types. The first is a subset of sporadic colorectal cancers. The great majority of these patients have no family history of cancer, are over 60 years of age, and have no detectable germline mutations in their mismatch repair genes<sup>26,34,54)</sup>. The second type is a subset of colorectal, endometrial and other cancers arising in related patients fulfilling the criteria for HNPCC<sup>1,44,48)</sup>. These patients have a strong family history of cancer, are often young, and most have germline mutations in their mismatch repair genes. The third type is a subset of many tumor types including cancers of the breast, as well as pancreas and stomach<sup>9,11,18,36,60)</sup>.

It has been reported that RER in sporadic human breast cancers was low<sup>2,14,30,44,50,56,63</sup>). However, in the present study, RER was more frequently observed in younger patients than in elderly patients. This discrepancy may reflect the difference of criteria of RER in the previous study<sup>2,14,30,50,56,63)</sup>. The significant difference of the RER rate, however, may have some significant meanings. It has been reported that RER correlates with lobular carcinoma, lymph node involvement and poor prognosis<sup>2,14,30</sup>. Despite this, the present study found no evidence that RER correlates with lobular carcinoma, lymph node involvement or noninvasive ductal carcinoma. It has also been suggested that RER is an early event in the genesis of occasional breast cancers<sup>50,56,63)</sup>. Moreover, another study in our laboratory indicated the presence of RER in preneoplastic breast cancer lesions<sup>25)</sup>, and in the present study, RER was not seen in any of the 16 elderly patients, 14 of whom had no lymph node metastasis. This may reflect a difference in etiology between young and elderly patients in the early carcinogenic process.

It is well known that individuals with mutations in one of the known mismatch repair genes (including MSH2, MLH1 and PMS1) have an increased likelihood of accumulating genetic alterations that lead to tumorigenesis<sup>8,19,33,40</sup>. It has been reported that a subset of breast cancer occurs due to the inheritance of a mutant mismatch repair gene<sup>47)</sup>. However, mutations in mismatch repair genes have not been reported in sporadic breast cancer. In the present study, RER or LOH were infrequently found at microsatellite loci on the MSH2 and MLH1 genes in all groups. However, these mismatch repair genes, including unknown genes, are likely to be involved in RER in young patients and those with bilateral breast cancer. Endometrial carcinoma which is the second commonest malignancy of the HNPCC syndrome is reported to show RER more frequently<sup>48)</sup>. However, it is also reported that primary endometrial carcinoma has low frequency of mutations in MLH1 and  $MSH2^{29}$ . These findings suggest that other mismatch repair genes are responsible for the RER phenotype in endometrial carcinoma as well as breast cancer. It will therefore be necessary to examine mutations in other mismatch repair genes. Also, it has been reported that there were frequent replication errors in post hepatitis B viral cirrhosis, even though hepatocellular carcinoma showed high frequency of LOH using microsatellite markers<sup>49)</sup>. These findings suggest RER was generated from another system different from germline and/or somatic mutation in mismatch repair genes.

In general, breast cancer in younger women (40 years of age or younger) has a poorer prognosis

than that in older women<sup>12)</sup>. It is probable that breast cancers in younger women proliferate more than those in postmenopausal aggressively women. Therefore, it is supposed that markers of poor prognosis exist at the molecular level without significant histopathologic differences. However, the prognosis of cancers with RER remains undetermined. For example, occasional colorectal cancers with RER and a right-sided anatomical distribution have a favorable prognosis<sup>26,34,54</sup>, and endometrial carcinomas with RER have a diploid or near diploid DNA content and a low recurrence rate<sup>48)</sup>. In contrast, RER has been reported to be more frequently observed in the poorly differentiated type of gastric cancer and in patients with multiple primary cancers<sup>18,20)</sup>. In our study, there was no significant correlation between RER and any clinicopathological factor (histological grade. and tumor size lymph node metastasis). Consequently, it is still unknown whether RER in breast cancer is a biological marker of poor prognosis.

RER has been frequently found in the tumors of patients with multiple primary cancers. Genetic instability is therefore believed to play an important role in the development of multiple primary cancers<sup>20,39)</sup>. Our results showed that RER occured more frequently in patients with double primary disease than in those with single cancers. Moreover, it was interesting that RER was found more frequently in the patients with contralateral metastases, and that RER was frequently detected at microsatellite loci on the p53 gene in these patients. Since abnormalities of the p53 gene have been reported to be more common in bilateral breast cancer than in unilateral breast cancer<sup>27</sup>, it is suggested that RER in bilateral breast cancer and breast cancer with contralateral metastases might correlate with an abnormality of the p53gene.

Inherited mutations in the BRCA1 and BRCA2 genes are known to confer a predisposition to breast cancer<sup>16,61)</sup>. BRCA1 mutation has been associated with estimated life-time risks of approximately 85 percent for breast cancer and 50 percent for ovarian cancer<sup>15,35)</sup>. Recently, alterations in the BRCA1 gene were identified in approximately 10 percent of a cohort of young women who were diagnosed before the age of 35 and were not selected on the basis of family history<sup>31)</sup>. Thus, mutations of the BRCA1 gene are thought to play an important role in breast-ovarian cancer families. Although the prevalence of BRCA1 gene mutations in young Japanese women with breast cancer has not been reported, the proportion of Japanese breast cancer and breast-ovarian cancer families who inherit the mutated BRCA1 allele appears to be small<sup>22)</sup>. Loss of heterozygosity at microsatellite loci on the BRCA1 and BRCA2 genes might correlate with larger tumor size and

higher histological grade in some breast cancers<sup>4</sup>). In the present study, we found only a few patients with alteration at one microsatellite locus (D17S855) on the *BRCA1* gene in every groups, but it will be necessary to examine alterations at many microsatellite loci on the *BRCA1* gene as well as investigating mutations using direct sequencing and on the basis of family history.

In summary, our preliminary data indicated that the percentage of RER in young patients (25-35 years old) and those with bilateral breast cancer was significantly higher than in elderly patients, while there was no significant correlation between the incidence of LOH and the ages of the patients or bilaterality. These findings suggest that RER may play an important role in early onset (occurrence at a young age) and bilateral breast cancer. Genetic analysis including RER or LOH in middle age patients (40-59 years old) is warranted. It is possible that genetic instability has general effects on activating oncogenes or inactivating tumor suppressor genes. However, further studies will be required to determine the true role of RER in the natural history of breast cancer.

> (Received February 26, 1998) (Accepted May 26, 1998)

#### REFERENCE

- Aaltonen, L.A., Peltomaki, P., Mecklin, J.-P., Jarvinen, H., Jass, J.R., Green, J.S., Lynch, H.T., Watson, P., Tallqvist, G., Juhola, M., Sistonen, P., Hamilton, S.R., Kinzler, K.W., Vogelstein, B. and Albert de la Chapelle. 1994. Replication errors in benign and malignant tumours from hereditary nonpolyposis colorectal cancer patients. Cancer Res. 54: 1645–1648.
- Aldaz, C.M., Chen, T., Sahin, A., Cunningham, J. and Bondy, M. 1995. Comperative Allelotype of in situ and invasive human breast cancer. Cancer Res. 55: 3976–3981.
- Anderson, L.A., Friedman, L., Osborne-Lawrence, S., Lynch, E., Weissenbach, J., Bowcock, A. and King, M-C. 1993. High-density genetic map of the BRCA1 region of chromosome 17q12-q21. Genomics 17: 618-623.
- Beckmann, M.W., Picard, F., An, H.X., van. Roeyen, C.R., Dominik, S.I., Mosny, D.S., Schnurch, H.G., Bender, H.G. and Niederacher, D. 1996. Clinical impact of detection of loss of heterozygosity of BRCA1 and BRCA2 markers in sporadic breast cancer. Br. J. Cancer. 73: 1220–1226.
- Bergthorsson, J.T., Eiriksdottir, G., Barkardottir, R.B., Egilsson, V., Arason, A. and Ingvarsson, S. 1995. Linkage analysis and allelic imbalance in human breast cancer kindreds using microsatellite markers from the short arm of chromosome 3. Hum. Genet. 96: 437–443.
- 6. Bieche, I., Champene, M-H. and Lidereau, R.

1995. Loss and gain of distinct regions of chromosome 1q in primary breast cancer. Clin. Cancer Res. 1: 123–127.

- Brenner, A.J. and Aldaz, C.M. 1995. Chromosome 9p allelic loss and p16/CDKN2 in breast cancer and evidence of p16 inactivation in immortal breast epithelial cells. Cancer Res. 55: 2892–2895.
- Bronner, C.E., Baker, S.M., Morrison, P.T., Warren, G., Smith, L.G., Lescoe, M.K., Kane, M., Earabino, C., Lipford, J., Lindblom, A., Tannergard, P., Bollag, R.J., Godwin, A.R., Ward, D.C., Nordenskjold, M., Fishel, R., Kolodner, R. and Liskay, R.M. 1994. Mutation in the DNA mismatch repair gene homologue hMLH1 is associated with hereditary non-polyposis colon cancer. Nature 368: 258-261.
- 9. Burks, R.T., Kessis, T.D., Cho, K.R. and Hedrick, L. 1994. Microsatellite instability in endometrial carcinoma. Oncogene 9: 1163–1166.
- Callen, D.F., Doggett, N.A., Stallings, R.L., Chen, L.Z., Whitmore, S.A., Lane, S.A., Nanccarrow, J.K., Apostolou, S., Thompson, A.D., Lapsys, N. M., Eyre, H.J., Baker, E.G., Shen, Y., Holman, K., Phillips, H., Richards, R. I. and Sutherland, G.R. 1992. High-resolution cytogenetic-based physical map of human chromosome 16. Genomics 13: 1178–1185.
- Chong, J-M., Fukayama, M., Hayashi, Y., Takizawa, T., Koike, M., Konishi, M., Kikuchi-Yanoshita, R. and Miyaki, M. 1994. Microsatellite instability in the progression of gastric carcinoma. Cancer Res. 54: 4595–4597.
- Chung, M., Chang, H.R., Bland, K.I. and Wanebo, H.J. 1996. Younger women with breast carcinoma have a poor prognosis than older women. Cancer 77: 97–103.
- Cleton-Jansen, A-M., Collins, N., Lakhani, S.R., Weissenbach, J., Devilee, P., Cornelisse, C.J. and Stratton, M.R. 1995. Loss of heterozygosity in sporadic breast tumours at BRCA2 locus on chromosome 13q12-q13. Br. J. Cancer 72: 1241–1244.
- Contegiacomo, A., Palmirotta, R., De Marchis, L., Pizzi, C., Mastrnzo, P., Delrio, P., Petrella, G., Figliolini, M., Bianco, A.R., Frati, L., Cama, A. and Mariani-Costantini, R. 1995. Microsatellite instability and pathological aspects of breast cancer. Int. J. Cancer 64: 264–268.
- Ford, D. and Easton, D.F. 1995. The genetics of breast and ovarian cancer. Br. J. Cancer 72: 805-812.
- Hall, J.M., Lee, M.K., Newman, B., Morrow, J.E., Anderson, L.A., Huey, B. and King, M.C. 1990. Linkage of early-onset familial breast cancer to chromosome 17q21. Science 250: 1684–1689.
- Hampton, G.M., Mannermaa, A., Winquist, R., Alavaikko, M., Blanco, G., Taskinen, P.J., Kiviniemi, H., Newsham, I., Cavenee, W.K. and Evans, G.A. 1994. Loss of heterozygosity in sporadic human breast carcinoma: A common region between 11q22 and 11q23.3. Cancer Res. 54:

4586-4589.

- Han, H-J., Yanagisawa, A., Kato, Y., Park, J-G. and Nakamura, Y. 1993. Genetic Instability in pancreatic cancer and poory differentiated type of gastric cancer. Cancer Res. 53: 5087–5089.
- Hemminiki, A., Peltomaki, P., Mecklin, J.-P., Jarvinen, H., Salovaara, R., Nystrom-Lahti, M., Albert de la Chapelle and Aaltonen, L.A. 1994. Loss of the wild type MLH1 gene is a feature of hereditary nonpolyposis colorectal cancer. Nature Genet. 8: 405-410.
- Horii, A., Han, H-J., Shimada, M., Yanagisawa, A., Kato, Y., Ohta, H., Yasui, W., Tahara, E. and Nakamura, Y. 1994. Frequent replication errors at microsatellilte loci in tumors of patients with multiple primary cancers. Cancer Res. 54: 3373–3375.
- Hung, T.H.-M., Yeh P.L.-H., Martin, M.B., Straub, R.E., Gilliam, T.C., Caldwell, C.W. and Skibba, J.L. 1995. Genetic alteration of microsatellites on chromosome 18 in human breast carcinoma. Diagn. Mol. Pathol. 4: 66–72.
- Inoue, R., Fukutomi, T., Ushijima, T., Matsumoto, Y., Sugimura, T. and Nagao, M. 1995. Germline mutation of BRCA1 in Japanese breast cancer families. Cancer Res. 55: 3521–3524.
- Jones, M.H. and Nakamura, Y. 1992. Detection of loss of heterozygosity at the human TP53 locus using a dinucleotide repeat polymorphism. Genes Chromosomes and Cancer 5: 89–90.
- Jones, M.H., Yamakawa, K. and Nakamura, Y. 1992. Isolation and characterization of 19 dinucleotide repeat polumorphism on chromosome 3p. Hum. Mol. Genet. 1: 131-133.
- 25. Kaneko, M., Arihiro, K., Takeshima, Y., Fujii, S. and Inai, K. Loss of heterozygosity and microsatellite instability in epithelial hyperplasia of the breast. Journal of Experimental Therapeurics and Oncology (in-press).
- Kim, H., Jen, J., Vogelstein, B. and Hamilton, S.R. 1994. Clinical and pathological characteristics of sporadic colorectal carcinomas with DNA replication errors in microsatellite sequences. Am. J. Pathol. 145: 148–156.
- Kinoshita, T., Ueda, M., Enomoto, K., Ikeda, T., Kikuchi, K., Ishii, S. and Kitajima, M. 1995. Comparison of p53 gene abnormalities in bilateral and unilateral breast cancer. Cancer 76: 2504–2509.
- Koreth, J., Bethwaite, P.B. and McGee, J.O. 1995. Mutation at chromosome 11q23 in human non-familial breast cancer: a microdissection microsatellite analysis. J. Pathol. 176: 11–18.
- Kowalski, L.D., Mutch, D.G., Herzog, T.J., Rader, J.S. and Goodfellow, P.J. 1997. Mutational analysis of MLH1 and MSH2 in 25 prospectively-aquired RER+ endometrial cancer. Genes. Chromosomes. Cancer 18: 219-227.
- 30. Kranik, P., Plummer, S., Casey, G., Myles, J., Tubbs, R., Crowe, J. and Williams, B.R. 1995. Microsatellite instability at a single locus (D11S988) on chromosome 11p15.5 as a late event

in mammary tumorigenesis. Hum. Mol. Genet. 4: 1889–1894.

- Langston, A.A., Malone, K.E., Thompson, J.D., Daling, J.R. and Ostrander, E.A. 1996. BRCA1 mutations in a population-based sample of young women with breast cancer. N. Engl. J. Med. 334: 137-142.
- 32. Liu, B., Farrington, S.M., Petersen, G.M., Hamilton, S.R., Parsons, R., Papadopoulos, N., Fujiwara, T., Jen, J., Kinzler, K.W., Wyllie, A.H., Vogelstein, B. and Dunlop, M.G. 1995. Genetic instability occurs in the majority of young patients with colorectal cancer. Nature Med. 1: 348–352.
- Liu, B., Parsons, R.E., Hamilton, S.R., Petersen, G.M., Lynch, H.T., Watson, P., Markowitz, S., Willson, J.K.V., Green J., Albert de la Chapelle, Kinzler, K.W. and Vogelstein, B. 1994. hMSH mutations in hereditary nonpolyposis colorectal cancer kindreds. Cancer Res. 54: 4590–4594.
- 34. Lothe, R.A., Peltomaki, P., Meling, G.I., Aaltonen, L.A., Nystrom-Lahti, M., Pylkkanen, L., Heimdal, K., Andersen, T.I., Moller, P., Rognum, T.O., Fossa, S.D., Haldorsen, T., Langmark, F., Brogger, A., Chapelle, A.D.L. and Borresen, A.-L. 1993. Genomic instability in colorectal cancer. Cancer Res. 53: 5849–5852.
- 35. Merajver, S.D., Frank, T.S., Xu, J., Pham, T.M., Calzone, K.A., Bennett-Baker, P., Chamberlain, J., Boyd, J., Garber, J.E., Collins, F.S. and Weber, B.L. 1995. Germline BRCA1 mutations and loss of the wild-type allle in tumors from families with early onset breast and ovarian cancer. Clin. Cancer Res. 1: 539–544.
- Merlo, A., Mabry, M., Gabrielson, E., Vollmer, R., Baylin, S.B. and Sidransky, D. 1994. Frequent microsatellite instability in primary small cell lung cancer. Cancer Res. 54: 2098–2101.
- 37. Munn, K.E., Walker, R.A., Menasce, L. and Varley, J.M. 1996. Allelic imbalance in the region of the BRCA1 gene in ductal carcinoma in situ of the breast. Br. J. Cancer 73: 636–639.
- 38. Nagai, H., Negrini, M., Carter, S.L., Gillum, D.R., Rosenberg, A.L., Schwartz, G.F. and Croce, C.M. 1995. Detection and cloning of a common region of loss of heterozygosity at chromosome 1p in breast cancer. Cancer Res. 55: 1752–1757.
- Nakashima, H., Honda, M., Inoue, H., Shibuta, K., Arinaga, S., Era, S., Ueo, H., Mori, M. and Akiyoshi, T. 1995. Microsatellite instability in multiple gastric cancers. Int. J. Cancer 64: 239–242.
- 40. Nicolaides, N.C., Papadopoulous, N., Liu, B., Wei, Y.-F., Carter, K.C., Ruben, S.M., Rosen, C.A., Haseltine, W.A., Fleischmann, R.D., Fraser, C.M., Adams, M.D., Venter, J.C., Dunlop, M.G., Hamilton, S.R., Petersen, G.M., Albert de la Chapelle, Vogelstein, B. and Kinzler, K.W. 1994. Mutations of two PMS homologues in hereditary nonpolyposis colon cancer. Nature **371**: 75–80.

- Orphanos, V., McGown, G., Hey, Y., Boyle, J.M. and Santibanez-Koref, M. 1995. Proximal 6q, a region showing allele loss in primary breast cancer. Br. J. Cancer 71: 290–293.
- 42. Papadopoulos, N., Nicolaides, N.C., Wei, Y-F., Ruben, S.M., Carter, K.C., Rosen, C.A., Haseltine, W.A., Fleischmann, R.D., Fraser, C.M., Petersen, G.M., Watson, P., Lynch, H.T., Peltomaki, P., Mecklin, J-P., Albert de la Chapelle, Kinzler, K.W. and Vogelstein, B. 1994. Mutation of a mutL homolog in hereditary colon cancer. Science 263: 1625–1629.
- Patel, U., Grundfest-Broiniatowski, S., Gupta, M. and Banerjee, S. 1994. Microsatellite instabilities at five chromosomes in primary breast tumors. Oncogene 9: 3695–3700.
- 44. Peltomaki, P., Lothe, R.A., Aaltonen, L.A., Pylkkanen, L., Nystrom-Lahti, M., Seruca, R., David, L., Holm, R., Ryberg, D., Haugen, A., Brogger, A., Borresen, A.-L. and Albert de la Chapelle. 1993. Microsatellite instability is associated with tumours that characterize the hereditary non-polyposis colorectal carcinoma syndrome. Cancer Res. 53: 5853–5855.
- Radford, D.M., Fair, K.L., Phillips, N.J., Ritter, J.H., Steinbrueck, T., Holt, M.S. and Donis-Keller, H. 1995. Allelotyping of ductal carcinoma in situ of the breast: deletion of loci on 8p, 13q, 16q, 17p and 17q. Cancer Res. 55: 3399–3405.
- 46. Radford, D.M., Fair, K.L., Thompson, A.M., Ritter, J.H., Holt, M., Steinbrueck, T. and Wallace, M. 1993. Allelic loss on chromosome 17 in ductal carcinoma in situ of breast. Cancer Res. 53: 2947–2950.
- 47. Risinger, J.I., Barrett, J.C., Watson, P., Lynch, H.T. and Boyd, J. 1996. Molecular genetic evidence of the occurence of breast cancer as an integral tumor in patients with the hereditary nonpolyposis colorectal carcinoma syndrome. Cancer 77: 1836–1843.
- Risinger, J.I., Berchuck, A., Kohler, M.F., Watson, P., Lynch, H.T. and Boyd, J. 1993. Genetic instability of microsatellites in endometrial carcinoma. Cancer Res. 53: 5100–5103.
- Salvucci, M., Lemoine, A., Azoulay, D., Sebagh, M., Bismuth, H., Reynes, M., May, E. and Debuire, B. 1996. Frequent microsatellite instability in post hepatitis B viral cirrhosis. Oncogene 13: 2681–2685.
- Shaw, J.A., Walsh, T., Chappell, S.A., Carey, N., Johnson, K. and Walker, R.A. 1996. Microsatellite instability in early sporadic breast cancer. Br. J. Cancer 73: 1393–1397.
- 51. Skirnisdottir, S., Eirikisdottir, G., Baldursson, T., Barkardottir, R.B., Eglisson, V. and Invarrson, S. 1995. High frequency of allelic imbalance at chromosome region 16q22–23 in human breast cancer: correlation with high PgR and low S phase. Int. J. Cancer 64: 112–116.
- 52. Spirin, K.S., Simpson, J.F., Takeuchi, S., Kawa-

mata, N., Miller, C.W. and Koeffler, H.P. 1996. p27/Kip1 mutation found in breast cancer. Cancer Res. 56: 2400–2404.

- 53. Takeshima, Y., Inai, K., Bennet, W.P., Metcalf, R.A., Welsh, J.A., Yonehara, S., Hayashi, Y., Fujihara, M., Yamakido, M., Akiyama, M., Tokuoka, S., Land, C.E. and Harris, C.C. 1994. p53 mutations in lung cancers from Japanese mustard gas workers. Carcinogenesis 15: 2075–2079.
- Thibodeau, S.N., Bren, G. and Shaid, D. 1993. Microsatellite instability in cancer of the proximal colon. Science 260: 816–819.
- Thompson, A.D., Shen, Y., Holman, K., Sutherland, G.R., Callen, D.F. and Richards, R.I. 1992. Isolation and characterisation of (AC)<sub>n</sub> microsatellite genetic markers from human chromosome 16. Genomics 13: 402–408.
- 56. Toyama, T., Iwase, H., Yamashita, H., Iwata, H., Yamashita, T., Ito, K., Suchi, M., Nakamura, T. and Kobayashi, S. 1996. Microsatellite instability in sporadic human breast cancer. Int. J. Cancer 68: 447–451.
- 57. Tsuda, H., Callen, D.F., Fukutomi, T., Nakamura, Y. and Hirohashi, S. 1994. Allele loss on chromosome 16q24.2-qter occurs frequently in breast cancers irrespectively of differences in phenotype and extent of spread. Cancer Res. **54:** 513–517.
- Weissenbach, J., Gyapay, G., Dib, C., Vignal, A., Morissette, J., Millasseau, P., Vaysseix, G. and Lathrop, M. 1992. A second-generation linkage map of the human genome. Nature 359: 794–801.
- Wick, W., Petersen, I., Schmutzler, R.K., Wolfarth, B., Lenartz, D., Bierhoff, E., Hummerich, J., Muller, D.J., Stangl, A.P., Schramm, J., Wiestler, O.D. and von Deimling, A. 1996. Evidence for a novel

tumor suppressor gene on chromosome 15 associated with progression to a metastatic stage in breast cancer. Oncogene **12**: 973–978.

- Wooster, R., Cleton-Jansen, A.-M., Collins, N., Mangion, J., Cornelis, R.S., Cooper, C.S., Gusterson, B.A., Ponder, B.A.J., von Deimling, A., Wiestler, O.D., Cornelisse, C.J., Devilee, P. and Stratton, M.R. 1994. Instability of short tandem repeats (microsatellites) in human cancers. Nature Genet. 6: 152–156.
- Wooster, R., Neuhausen, S.L., Mangion, J., Quirk, Y., Ford, D., Collins, N., Nguyen, K., Seal, S., Tran, T., Averill, D., Fields, P., Marshall, G., Narod, S., Lenoir, G.M., Lynch, H., Feunteun, J., Devilee, P., Cornelisse, C.J., Menko, F.H., Daly, P.A., Ormiston, W., Mcmanus, P., Pye, C., Lewis, C.M., Cannonalbright, L.A., Peto, J., Ponder, B.A.J., Skolnick, M.H., Easton, D.F., Goldgar, D.E. and Stratton, M.R. 1994. Localization of a breast cancer susceptibility gene, BRCA2, to chromosome 13q12–13. Science 265: 2088–2090.
- 62. Yaremko, M.L., Recant, W.M. and Westbrook, C.A. 1995. Loss of heterozygosity from the short arm of chromosome 8 is an early event in breast cancers. Genes Chromosomes and Cancer 13: 186-191.
- Yee, C.J., Roodi, N., Verrier, C.S. and Parl, F.F. 1994. Microsatellite instability and loss of heterozygosity in breast cancer. Cancer Res. 54: 1641–1644.
- 64. Zenklusen, J.C., Bieche, I., Lidereau, R. and Conti, C.J. 1994. (C-A)n microsatellite repeat D7S522 is the most commonly detected region in human primary breast cancer. Proc. Natl. Acad. Sci. USA. 91: 12155–12158.