

A Japanese Pedigree of Familial Cerebral Cavernous Malformations — A Case Report —

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ABSTRACT

Familial cerebral cavernous malformations (FCCM) are autosomal-dominant vascular malformations. At present, 3 cerebral cavernous malformation genes (KRIT1/CCM1, MGC4607/CCM2, and PDCD10/CCM3) have been identified. Few genetic analyses of Japanese FCCM have been reported. A Japanese pedigree of 4 patients with FCCM has been reported that includes the genetic analysis of one of the patients. All 4 patients showed multiple lesions in the brain. Surgical removal was performed at our hospital due to enlargement or hemorrhage of the intracranial lesions in a 21-year-old female (Case 1) and a 30-year-old male (Case 2). The histological diagnoses were cavernous malformations. A 62-year-old female (Case 4), the mother of Cases 1, 2, and 3, suffered from intramedullary hemorrhage at T6-7 and surgical removal was performed at another hospital. Only one patient, a 32-year-old female (Case 3), did not show symptoms. The genetic analysis of Case 2 demonstrated heterozygous partial deletions of exons 12-15 of the KRIT1 gene.

Key words: Cerebral cavernous malformation, Familial cerebral cavernous malformation, Genetic analysis, Japanese

Cerebral cavernous malformations (CCM) are congenital vascular anomalies of the central nervous system that show a prevalence of 0.1-0.5% in the general population²². The lesions are characterized by abnormally enlarged capillary cavities without intervening brain parenchyma²³. CCM occurs in both a sporadic and an inherited form. The proportion of familial cerebral cavernous malformations (FCCM) has been estimated to be as high as 50% in Hispano-American CCM patients²² and close to 10-40% in Caucasian patients^{12,22}. FCCM are characterized by the presence of multiple lesions and are inherited in an autosomal-dominant fashion^{11,12,22}. While 3 CCM genes have been identified (KRIT1/CCM1, MGC4607/CCM2, PDCD10/CCM3)^{1,4,13,14,24}, few reports have conducted a genetic analysis of Japanese FCCM. In this study, we present a Japanese pedigree of FCCM and report the genetic analysis of 1 patient.

CASE REPORTS

Case 1 (III:5, Fig. 1) : A 21-year-old female with a history of loss of consciousness at 19 years old

suffered general convulsion and was admitted to our hospital. Neurological and physical examination revealed no abnormalities, especially in the skin and mucosa. Computed tomography (CT) demonstrated 3 high-density areas in the bilateral frontal and right temporal lobe (Fig. 2A). T2-weighted magnetic resonance imaging (MRI) revealed 3 cores of mixed signal intensity areas with hypointensity rims in the bilateral frontal and right temporal lobe, and T2 star-weighted MRI revealed one small hypointensity area in the right cingulate gyrus (Fig. 2B-C). Bilateral carotid angiography showed no vascular anomalies. The right temporal lesion was removed surgically at 3 months after the first admission. Twenty-one months after the first surgery, two lesions in the bilateral frontal lobe were removed surgically due to their enlargement (Fig. 3A). Six months after the second surgery, she suffered intracerebral hemorrhage from the residual lesion in the right cingulate gyrus (Fig. 3B), and T2 star-weighted MRI revealed a small *de novo* lesion in the left midbrain (Fig. 3C). Emergency surgery was performed to evacuate a hematoma and the residual lesion in the right

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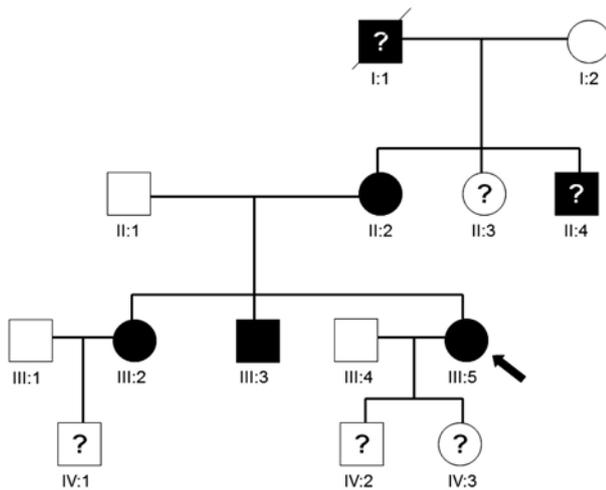


Fig. 1. The family tree

Closed circles or closed squares indicate the affected members, while open circles and open squares indicate the unaffected members. Question marks inside closed squares show the members who are probably affected. Question marks inside open circles and squares show the members who are not known to be affected. A diagonal line indicates a deceased member.

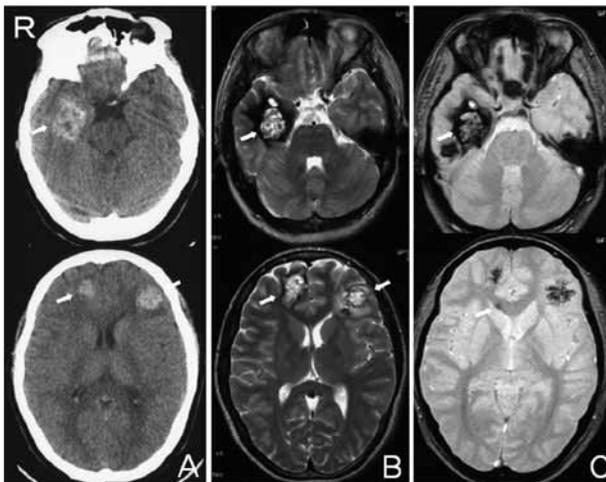


Fig. 2. Preoperative computed tomography (CT) scans (A) and T2-weighted magnetic resonance imaging (MRI) of Case 1 (B) show 3 cavernous malformations in the bilateral frontal and right temporal lobe (arrows). Preoperative T2 star-weighted MRIs of Case 1 (C) show a small cavernous malformation in the right cingulate gyrus (arrows). The temporal lesion was removed at the first surgery.

cingulate gyrus was removed. Histological diagnoses were cavernous malformations in all of the lesions. She was discharged from the hospital with no neurological deficit. Follow-up MRI showed the lesion in the midbrain was unchanged, and no more *de novo* lesions were observed at 4 years after the third surgery (Fig. 3D).

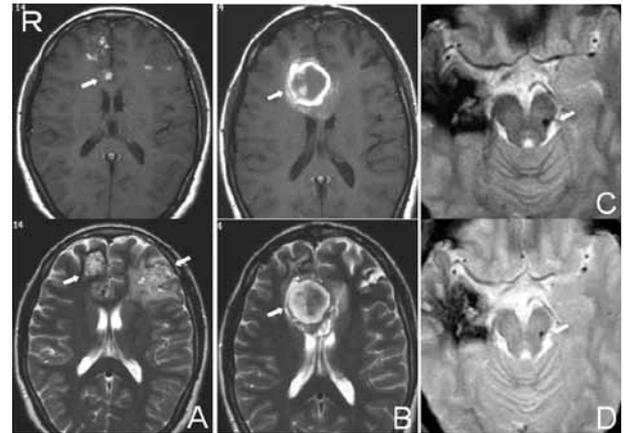


Fig. 3. T1 and T2-weighted MRIs of Case 1 at 21 months after the first surgery (A) show apparent enlargements of the two lesions in the bilateral frontal lobe. They were removed at the second surgery. T1 and T2-weighted MRIs of Case 1 at 6 months after the second surgery (B) show intracerebral hemorrhage from the residual lesion in the right cingulate gyrus. It was removed at the third surgery. T2 star-weighted MRIs of Case 1 at 6 months after the second surgery (C) show a small *de novo* lesion in the left midbrain. T2 star-weighted MRIs of Case 1 at 4 years after the third surgery (D) show no changes.

Case 2 (III:3, Fig. 1) : A 30-year-old male with no medical history, the elder brother of Case 1, suffered from loss of consciousness and was admitted to our hospital at 6 years after the first admission of Case 1. Neurological and physical examination revealed no abnormalities, especially in the skin and mucosa. T2-weighted MRI revealed 3 cores of mixed signal intensity areas with hypointensity rims in the right frontal, right parietal and right caudate nucleus (Fig. 4A), and T2 star-weighted MRI revealed 5 small hypointensity areas in the bilateral cerebral hemisphere (Fig. 4B). T2-weighted MRI sagittal views revealed no lesion in the spinal cord. Bilateral carotid angiography showed no vascular anomalies. At 5 months after onset, two lesions in the right frontal and right parietal lobe were removed surgically due to their enlargement (Fig. 4C). The histological diagnoses were cavernous malformations. He was diagnosed with FCCM from his familial history. He was discharged from the hospital with no neurological deficit. We recommended that other family members receive MRI examinations and we obtained informed consent for the genetic analysis from Case 2. We performed the genetic analysis by outsourcing to GeneDX®, a genetic testing company founded in 2000 by 2 scientists from the National Institutes of Health in the United States. A blood sample was submitted to GeneDX. Genetic analysis demonstrated the following. First, the sequence

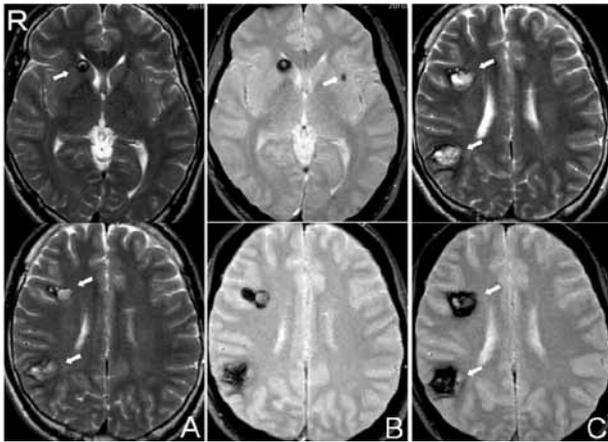


Fig. 4. Preoperative T2-weighted MRIs of Case 2 (A) show 3 cavernous malformations in the bilateral frontal and right temporal lobe (arrows). Preoperative T2 star-weighted MRIs of Case 2 (B) show 5 small hypointensity lesions in the bilateral cerebral hemisphere (arrows). T2 and T2 star-weighted MRIs of Case 2 at 5 months after the onset show enlargement of 2 lesions in the right frontal and right parietal lobe. They were removed surgically.

analysis of exons 14, 16, and 18 of the KRIT1 gene and their flanking splice sites revealed no mutation. Second, the detection of deletions and duplications of the KRIT1, MGC4607, and PDCD10 genes was performed by exon-focused oligonucleotide array comparative genomic hybridization analysis. The analysis revealed heterozygous partial deletions of exons 12-15 of the KRIT1 gene. The reference genomic sequence coordinate was chromosome 7: 91850937-91856230, based on the Genome Reference Consortium build 37 (GRCh37). Hybridization data were analyzed with Genomic Workbench v5 software (Agilent Technologies). No additional deletions were detected in the MGC4607 or PDCD10 genes. Third, the sequence analysis of exons 1-10 of the MGC4607 genes and their flanking splice sites revealed no mutation.

Case 3 (III:2, Fig. 1) : Case 3 was a 32-year-old female with no symptoms. She was the elder sister of Cases 1 and 2. T2 star-weighted MRI revealed 14 small hypointensity lesions in the bilateral cerebral hemisphere, left cerebellar hemisphere, and brain stem at 7 years after the first admission of Case 1 (Fig. 5A-B). Follow-up MRI showed no change at 2 years after the first examination.

Case 4 (II:2, Fig. 1) : Case 4 was a 62-year-old female with no symptoms. She was the mother of Cases 1, 2 and 3. T2 star-weighted MRI revealed 55 small hypointensity lesions in the bilateral cerebral hemisphere, left cerebellar hemisphere,

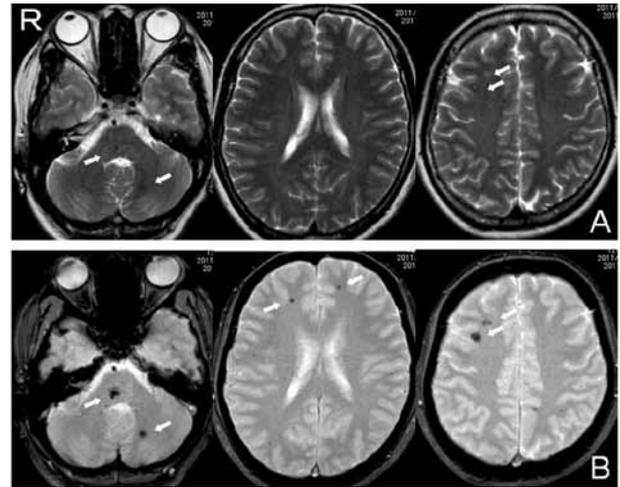


Fig. 5. T2-weighted MRIs of Case 3 (A) show 6 lesions in the bilateral cerebral hemisphere, left cerebellar hemisphere, and brain stem. Axial T2 star-weighted MRIs of Case 3 (B) show 14 lesions in the bilateral cerebral hemisphere, left cerebellar hemisphere, and brain stem.

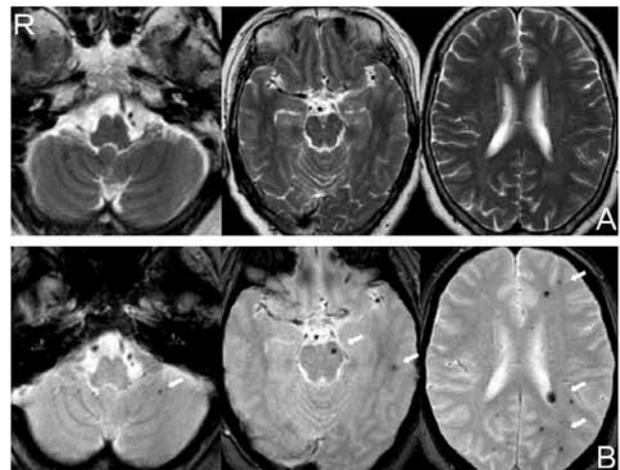


Fig. 6. T2-weighted MRIs of Case 4 (A) show 9 lesions in the bilateral cerebral hemisphere, left cerebellar hemisphere, and brain stem. T2 star-weighted MRIs of Case 4 (B) show 55 lesions in the bilateral cerebral hemisphere, left cerebellar hemisphere, and brain stem.

and brain stem (Fig. 6A-B). She had also been followed up at our hospital. However, she did not receive MRI examination of the spinal cord. At 1 year after the diagnosis, she suffered from paraplegia and was transferred to another hospital. T1-weighted and T2-weighted MRI revealed intramedullary hemorrhage at T6-7 and surgery was performed at that hospital. It was assumed that she had a lesion of CM lesion in the spinal cord. Her husband had multiple cutaneous vascular malformations on his face, head and neck, although MRI revealed no lesions of CM lesions in the brain or the spinal cord. Therefore, detailed family histories were taken from her

after the diagnosis. Her younger brother had previously shown the presence of a multiple mass lesion in the brain at the other hospital. Her younger sister had not received MRI. Her mother had received MRI, which revealed no abnormal lesions in the brain. Her father may have died from stroke, but the details were unclear. The family tree is shown in Fig. 1.

DISCUSSION

We have reported a Japanese pedigree of FCCM that includes 4 cases. These cases indicate four important clinical issues. First, all 4 cases had multiple lesions, and T2 star-weighted MRI was useful for the diagnosis. Second, the probability of showing symptoms was high (3/4) in all of these cases. Third, inheritance seemed to be autosomal-dominant from their family tree. Finally, genetic analysis of Case 2 demonstrated heterozygous partial deletions of exons 12-15 of the KRIT1 gene.

There are few case reports of Japanese FCCM compared with patients from other countries, such as Italy³⁾, the United States^{2,17)}, Brazil⁶⁾, Switzerland⁹⁾, Spain and Portugal¹⁹⁾. In addition, only one report conducted genetic analysis of Japanese FCCM²⁷⁾. Thus, the pathogenesis, natural history, inheritance fashion, and gene mutations related to Japanese FCCM are unclear. A previous review of 17 Japanese pedigrees and 37 cases of FCCM reported 17 cases of multiple lesions²⁶⁾, although T2 star-weighted MRI may reveal more cases with multiple lesions. The same review showed that surgical removals were performed in 22 cases due to hemorrhages, focal deficits, or epilepsies²⁶⁾, which indicated a high probability of showing symptoms of FCCM. FCCM is reported to be inherited in an

autosomal-dominant fashion^{11,12,22)}, thus many unrecognized cases of FCCM may exist in Japanese multiple CCM. Taking detailed family histories is important for a precise diagnosis. Currently 3 CCM genes have been identified: KRIT1 (Krev Interaction Trapped 1) at 7q21.2 (CCM1); MGC4607 at 7p13 (CCM2); and PDCD10 (Programmed Cell Death 10) at 3q25.2-q27 (CCM3)^{1,4,13,14,24)}. KRIT1 accounts for 40%-56% of FCCM^{2,14,28)}, MGC4607 accounts for 20%-33% of FCCM^{1,2,4,14)}, and PDCD10 accounts for 6%-10% of FCCM cases^{1,15,28)}. Other CCM genes or a significant fraction of CCM mutations that are not detected by routine direct DNA sequence analysis account for 5%-30% of FCCM cases^{5,8,15-17,28)}. There are several reports on large deletions or duplications within the three known CCM genes^{8,10,16-18,20,21,27)}, and deletions of CCM2 were the most frequent^{4,16)}, with similar location and type of CCM mutation in the same pedigree in many cases^{8,10,16,18,20,21,27)}. Oligonucleotide array comparative genomic hybridization has been used to identify deletions and duplications that are not detected by routine direct DNA sequence analysis^{25,29)}. The technique revealed heterozygous partial deletions of exons 12-15 of the KRIT1 gene in our Case 2. We predict that genetic analysis of Cases 1, 3, and 4 would reveal mutations similar to those in the previous reports mentioned above. However, it is unclear whether this location of the deletion of the KRIT1 is rare or not, because few reports gave details about deletions of the KRIT1^{8,16,27)}. Recently, a genetic analysis of 10 cases of Japanese FCCM was reported (Table 1)²⁷⁾. It showed that no CCM1, CCM2 or CCM3 mutations were identified in 4 patients, and surgical removal was performed due to symptoms in 2 cases with mutations other than CCM1,

Table 1. Reported cases of genetic analysis of Japanese FCCM

Author (Year)	Age (years)/ gender	Location	Exon	Type	procedure
Tsutsumi et al (2013)	60/F	CCM1	Exon 16	HD	
	37/F	CCM1	Exon 16	HD	
	30/F	UV	UV	UV	surgery
	46/F	UV	UV	UV	
	65/M	CCM3	Exon 5	N	
	37/M	CCM2	Exon 1	Mi	
	45/M	CCM2	Exon 2	D	
	79/M	CCM2	Exon 2	D	
	29/M	UV	UV	UV	
	15/M	UV	UV	UV	surgery
Present case	30/M	CCM1	Exon12-15	HD	surgery

FCCM: familial cerebral cavernous malformations, F: female, M: male, D: deletion,

HD: heterozygous deletion, Mi: missense, N: nonsense, UV: unverified

CCM2 or CCM3. The author assumed that cases associated with CCM1, CCM2, or CCM3 mutations may have benign clinical courses in contrast to previous reports²⁵. However, this assumption was invalid for our cases. In our cases, the larger genomic deletion spanning exons 12-15 may relate to symptomatic CCA. Further genetic analyses of Japanese FCCM are important in order to clarify the relationship between mutation patterns and their clinical courses.

CONFLICTS OF INTEREST DISCLOSURE

The authors have no personal financial or institutional interest in any of the drugs, materials, or devices mentioned in this article. All authors who are members of The Japan Neurosurgical Society (JNS) have registered online Self-reported COI Disclosure Statement Forms through the website for JNS members.

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REFERENCES

- Bergametti, F., Denier, C., Labauge, P., Arnoult, M., Boetto, S., Clanet, M., et al. 2005. Mutations within the programmed cell death 10 gene cause cerebral cavernous malformations. *Am. J. Hum. Genet.* **76**: 42-51.
- Craig, H.D., Gunel, M., Cepeda, O., Johnson, E.W., Ptacek, L., Steinberg, G.K., et al. 1998. Multilocus linkage identifies two new loci for a mendelian form of stroke, cerebral cavernous malformation, at 7p15-13 and 3q25.2-27. *Human Molecular Genetics* **7**: 1851-1858.
- D'Angelo, R., Marini, V., Rinaldi, C., Origone, P., Dorcaratto, A., Avolio, M., et al. 2011. Mutation analysis of CCM1, CCM2 and CCM3 genes in a cohort of Italian patients with cerebral cavernous malformation. *Brain Pathol.* **21**: 215-224.
- Denier, C., Goutagny, S., Labauge, P., Krivosic, V., Amoult, M., Cousin, A., et al. 2004. Mutations within the MGC4607 gene cause cerebral cavernous malformations. *Am. J. Hum. Genet.* **74**: 326-337.
- Denier, C., Labauge, P., Bergametti, F., Marchelli, F., Riant, F., Amoult, M., et al. 2006. Genotype-phenotype correlations in cerebral cavernous malformations patients. *Ann. Neurol.* **60**: 550-556.
- Domingues, F., Gasparetto, E.L., Andrade, R., Noro, F., Eiras, A., Gault, J., et al. 2008. Familial cavernous malformations: Rio de Janeiro study and review of the recommendations for management. *Arq. Neuropsiquiatr.* **66**: 795-799.
- Dubovsky, J., Zabramski, J.M., Kruth, J., Spetzler, R.F., Rich, S.S., Orr, H.T., et al. 1995. A gene responsible for cavernous malformations of the brain maps to chromosome 7q. *Human Molecular Genetics* **4**: 453-458.
- Gaetzner, S., Stahl, S., Sürücü, O., Schaafhausen, A., Halliger-Keller, B., Bertalanffy, H., et al. 2007. CCM1 gene deletion identified by MLPA in cerebral cavernous malformation. *Neurosurg. Rev.* **30**: 155-159.
- Graeni, C., Stepper, F., Sturzenegger, M., Merlo, A., Verlaan, D.J., Andermann, F., et al. 2010. Inherited cavernous malformations of the central nervous system: clinical and genetic features in 19 Swiss families. *Neurosurg. Rev.* **33**: 47-51.
- Guarnieri, V., Muscarella, L.A., Amoroso, R., Quattrone, A., Abate, M.E., Coco, M., et al. 2007. Identification of two novel mutations and of a novel critical region in the KRIT1 gene. *Neurogenetics* **8**: 29-37.
- Labauge, P., Laberge, S., Brunereau, L., Levy, C. and Tournier-Lasserre, E. 1998. Hereditary cerebral cavernous angiomas: clinical and genetic features in 57 French families. *Lancet* **352**: 1892-1897.
- Labauge, P., Denier, C., Bergametti, F. and Tournier-Lasserre, E. 2007. Genetics of cavernous angiomas. *Lancet Neurol.* **6**: 237-244.
- Laberg-le Couteulx, S., Jung, H.H., Labauge, P., Houtteville, J.P., Lescoat, C., Cecillon, M., et al. 1999. Truncating mutations in CCM1, encoding KRIT1, cause hereditary cavernous angiomas. *Nat. Genet.* **23**:189-193.
- Liquori, C.L., Berg, M.J., Siegel, A.M., Huang, E., Zawistowski, J.S., Stoffer, T., et al. 2003. Mutations in a gene encoding a novel protein containing a phosphotyrosine-binding domain cause type 2 cerebral cavernous malformations. *Am. J. Hum. Genet.* **73**: 1459-1464.
- Liquori, C.L., Berg, M.J., Squitieri, F., Ottenbacher, M., Sorlie, M., Leedom, T.P., et al. 2006. Low frequency of PDCD10 mutations in a panel of CCM3 probands: potential for a fourth CCM locus. *Hum. Mutat.* **27**: 118.
- Liquori, C.L., Berg, M.J., Squitieri, F., Leedom, T.P., Ptacek, L., Johnson, E.W., et al. 2007. Deletions in CCM2 are a common cause of cerebral cavernous malformations. *Am. J. Hum. Genet.* **80**: 69-75.
- Liquori, C.L., Penco, S., Gault, J., Leedom, T.P., Tassi, L., Esposito, T., et al. 2008. Different spectra of genomic deletions within the CCM genes between Italian and American CCM patients cohorts. *Neurogenetics* **9**: 25-31.
- Muscarella, L.A., Guarnieri, V., Coco, M., Belli, S., Parrella, P., Pulcrano, G., et al. 2010. Small deletion at the 7q21.2 locus in a CCM family detected by Real-Time Quantitative PCR. *J. Biomed. Biotechnol* 2010. pii: 854737. doi: 10.1155/2010/854737.
- Ortiz, L., Costa, A.F., Bellido, M.L., Solano, F., Garcia-Moreno, J.M., Gamero, M.A., et al. 2007. Study of cerebral cavernous malformation in Spain and Portugal: high prevalence of a 14bp deletion in exon 5 of MGC4607(CCM2 gene). *J. Neurol.* **254**:322-326.
- Penco, S., Ratti, R., Bianchi, E., Citterio, A., Patrosso, M.C., Marocchi, A., et al. 2009. Molecular screening test in familial forms of cerebral cavernous malformation: the impact of the Multiplex Ligation-dependent Probe Amplification approach. *J. Neurosurg.* **110**: 929-934.
- Pileggi, S., Buscone, S., Ricci, C., Patrosso, M.C.,

- Marocchi, A., Brunori, P., et al.** 2010. Genetic variations within KRIT1/CCM1, MGC4607/CCM2 and PDCD10/CCM3 in a large Italian family harbouring a Krit1/CCM1 mutation. *J. Mol. Neurosci.* **42**: 235-242.
22. **Rigamonti, D., Hardley, M.N., Drayer, B.P., Johnson, P.C., Hoenig-Rigamonti, K., Knight, J.T., et al.** 1988. Cerebral cavernous malformations: incidence and familial occurrence. *N. Engl. J. Med.* **319**: 343-347.
23. **Russell, D.S. and Rubinstein, L.J.** 1989. *Pathology of Tumors of the Nervous System*, 5th edition. pp.730-736. Williams & Wilkins, Baltimore.
24. **Sahoo, T., Johnson, E.W., Thomas, J.W., Kuehl, P.M., Jones, T.L., Dokken, C.G., Touchman, J.W., Galione, C.J., Lee-Lin, S.Q., Kosofsky, B., et al.** 1999. Mutations in the gene encoding KRIT1, a Krev-1/rap1a binding protein, cause cerebral cavernous malformations (CCM1). *Hum. Mol. Genet.* **8**:2325-2333.
25. **Saillour, Y., Cossée, M., Leturcq, F., Vasson, A., Beugnet, C., Poirier, K., et al.** 2008. Detection of exonic copy-number changes using a highly efficient oligonucleotide-based comparative genomic hybridization-array method. *Hum. Mutat.* **29**: 1083-1090.
26. **Sunada, I., Nakabayashi, H., Tsuchida, K. and Tanaka, A.** 2001. A case of familial cerebral cavernous angioma and review of Japanese cases. *No Shinkei Geka. Apr.* **29**: 359-365.
27. **Tsutsumi, S., Ogino, I., Miyajima, M., Ikeda, T., Shindo, N., Yasumoto, Y., et al.** 2013. Genomic causes of multiple cerebral cavernous malformations in a Japanese population. *J. Clin. Neurosci.* **20**: 667-669.
28. **Verlaan, D.J., Roussel, J., Laurent, S.B., Elger, C.E., Siegel, A.M. and Rouleau, G.A.** 2005. CCM3 mutations are uncommon in cerebral cavernous malformations. *Neurology* **65**:1982-1983.
29. **Wong, L.J., Dimmock, D., Geraghty, M.T., Quan, R., Lichter-Konecki, U., Wang, J., et al.** 2008. Utility of oligonucleotide array-based comparative genomic hybridization for detection of target gene deletions. *Clin. Chem.* **54**: 1141-1148.