

A crotofolane-type diterpenoid and a rearranged nor-crotofolane-type diterpenoid with a new skeleton from the stems of *Croton cascarilloides*

Susumu Kawakami ^a, Katsuyoshi Matsunami ^a, Hideaki Otsuka ^{a,*}, Takakazu Shinzato ^b, Yoshio Takeda ^c, Masatoshi Kawahata ^d, Kentaro Yamaguchi ^d

^a *Department of Pharmacognosy, Graduate School of Biomedical Sciences, Hiroshima University, 1-2-3 Kasumi, Minami-ku, Hiroshima 734-8553, Japan*

^b *Subtropical Field Science Center, Faculty of Agriculture, University of the Ryukyus, 1 Senbaru, Nishihara-cho, Nakagami-gun, Okinawa 903-0213, Japan*

^c *Faculty of Pharmacy, Yasuda Women's University, 8-13-1 Yasuhigashi, Asaminami-ku, Hiroshima 731-0153, Japan*

^d *Department of Analytical Chemistry, Faculty of Pharmaceautical Sciences, Tokushima Bunri University, Kagawa, 1314-1 Shido, Sanuki City, Kagawa 769-2193, Japan*

* Corresponding author.

E-mail address: hotsuka@hirohisma-u.ac.jp (H. Otsuka).

ABSTRACT

From the stems of *Croton cascarilloides* collected in the Okinawa Islands, a structurally rare crotofolane-type diterpenoid (**1**) and a rearranged nor-crotofolane, a new skeletal diterpenoid (**2**), were isolated. The structures were determined by X-ray crystallographic analyses, establishing their absolute stereostructures for the first time. Compound **2** was probably biosynthesized from **1** through several steps, such as decarboxylation, oxidation, C-C bond migration, etc.

Keywords:

Croton cascarilloides

Euphorbiaceae

Crotofolane

Diterpenoid

Crotofolane-type diterpenoids have fused 5-, 6- and 7-membered rings and are expected to be biosynthesized from cembrane via lathyrane through cross annular cyclization.¹ These diterpenoids have been found in only three *Croton* species, Jamaican *C. coryliformis*,^{1,2} Kenyan *C. dichogamus*,³ and Congolese *C. humanianus*.⁴

Our phytochemical investigation of the stems (14.5 kg) of *Croton cascarilloides* Rauschel, collected in the Okinawa Islands, led to the isolation of a crotofolane-type diterpenoid and a rearranged nor-molecular species of it having a new skeleton. A MeOH extract of branches of *C. cascarilloides* was washed with *n*-hexane and then evaporated to a gummy mass, which was then suspended in H₂O and extracted with CH₂Cl₂. The CH₂Cl₂-soluble fraction was separated by normal and reversed-phase silica gel column chromatographies, Sephadex LH-20 column chromatography, and then HPLC to afford compounds **1**⁵ and **2** (30.0 mg and 3.5 mg, respectively).⁶

Compound **1**⁵ was isolated as colorless plates and its elemental composition was determined to be C₂₅H₃₂O₇. The IR spectrum of **1** showed absorptions for ester carbonyl and lactone carbonyl groups. ¹³C-NMR of compound **1** revealed 25 resonances, five of which were assignable to 2-methylbutanoic acid. The remaining 20 signals comprised those of three methyls, two methylenes, seven methines, one tetra- and one disubstituted double bond, respectively, and three quaternary carbons. Precise inspection

of two-dimensional NMR spectra led to the conclusion that compound **1** was a diterpenoid with an unusual carbon skeleton. Thus, X-ray crystallographic analysis of **1** was performed and the relative stereostructure of **1** was established to be a derivative of crotofolane-type diterpenoid (Figs. 1 and 2).⁷ The positive Cotton effect in the CD spectrum empirically indicated the absolute configuration at the 9-position was *S*⁸ and chirality analysis of the 2-methylbutanoic acid moiety by HPLC established the absolute configuration of **1**, as shown in Fig. 1.⁹ This is the first report of the absolute structure of a crotofolane and the absolute configuration of the pentanolide portion, presumed based on the empirical rule for the CD spectrum, was proved to be correct.

Compound **2**⁶ was isolated as colorless plates and its elemental composition was determined to be C₂₄H₃₂O₈. The ¹³C-NMR spectrum displayed 24 signals, including five attributable to 2-methylbutanoic acid. Thus, the core skeleton was constituted of 19 carbons. X-ray crystallographic analysis revealed that compound **2** has a new skeleton, such as that of a rearranged mononor-crotofolane, as shown in Figs. 1 and 3.¹⁰ Compound **2** was probably derived from some crotofolane, like compound **1**, through several steps, such as decarboxylation, oxidation, C-C bond migration, etc.

Supplementary data

Supplementary X-ray crystallographic data for **1** (CCDC 761004) and **2** (CCDC 761005) can be obtained free of charge via www.ccdc.cam.ac.uk/conts/retrieving.html (or from the Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: (+44) 1223-336-033; or deposit@ccdc.cam.ac.uk).

Acknowledgements

The authors are grateful for access to the superconducting NMR instrument at the Analytical Center of Molecular Medicine of the Hiroshima University Faculty of Medicine and an Applied Biosystem QSTAR XL system ESI (Nano Spray)-MS at the Analysis Center of Life Science of the Graduate School of Biomedical Sciences, Hiroshima University.

References and Notes

1. Chan, W. R.; Prince, E. C.; Manchand, P. S.; Springer, J. P.; Clardy, J. *J. Am. Chem. Soc.* **1975**, *97*, 4439.
2. Burke, B. A.; Chan, W. R.; Pascoe, K. O.; Blout, J. F.; Manchand, P. S. *Tetrahedron Lett.* **1979**, 3345.

3. Jogia, M. K.; Andersen, R. A.; Párkány, L.; Clardy, J.; Dublin, H. T.; Sinclari, A. R. *E. J. Org. Chem.* **1989**, 54, 1654.
4. Tchissambou, L.; Chiaroni, A.; Riche, C.; Khung-Huu, F. *Tetrahedron* **1990**, 46, 5199.
5. Compound **1**: colorless plates (2-PrOH), m.p. 152-153 °C, $[\alpha]_D^{26} +81.8$ (c 1.52, CHCl₃). IR (KBr) ν_{\max} cm⁻¹: 3478, 2972, 2929, 2879, 1769, 1739, 1659, 1457, 1185, 1143, 1014, 804. UV (MeOH) λ_{\max} nm (log ϵ): 218 (4.00). ¹H-NMR (CDCl₃, 400 MHz): δ 5.38 (1H, d, $J = 5$ Hz, H-1), 5.20 (1H, s, H-18a), 5.17 (1H, s, H-18b), 5.14 (1H, dddd, $J = 13, 4, 2, 2$ Hz, H-9), 4.53 (1H, ddd, $J = 4, 2, 2$ Hz, H-13), 3.19 (1H, s, H-5), 3.15 (1H, d, $J = 13$ Hz, H-13), 3.06 (1H, br d, $J = 12$ Hz), 2.49 (1H, dd, $J = 14, 8$ Hz, H-3a), 2.73 (1H, ddd, $J = 13, 4, 4$ Hz, H-10a), 2.49 (1H, qdd, $J = 7, 7, 7$ Hz, H-2'), 2.44 (1H, dd, $J = 2, 2$ Hz, -OH), 2.18 (1H, dqdd, $J = 8, 7, 7, 5$ Hz, H-2), 1.90 (3H, br s, H₃-17), 1.74 (1H, ddq, $J = 14, 7, 7$ Hz, H-3'a), 1.70 (1H, dd, $J = 14, 10$ Hz, H-2b), 1.50 (1H, ddq, $J = 14, 7, 7$ Hz, H-3'b), 1.27 (1H, dddd, $J = 13, 13, 4, 2$ Hz, H-10b), 1.17 (3H, d, $J = 7$ Hz, H₃-5'), 1.06 (3H, s, H₃-20), 0.98 (3H, d, $J = 7$ Hz, H₃-19), 0.93 (3H, dd, $J = 7, 7$ Hz, H₃-4'). ¹³C-NMR (CDCl₃, 100 MHz): δ 178.0 (C-1'), 173.4 (C-16), 162.0 (C-8), 148.9 (C-12), 128.2 (C-15), 115.2 (C-18), 78.4 (C-9), 75.8 (C-1), 72.7 (C-11), 68.8 (C-14), 60.1 (C-4), 57.8 (C-5), 55.9 (C-6), 44.4

(C-7), 44.1 (C-10), 41.2 (C-2'), 36.4 (C-3), 32.7 (C-2), 31.7 (C-13), 26.6 (C-3'), 19.3 (C-20), 16.2 (C-5'), 12.3 (C-19), 11.4 (C-4'), 9.7 (C-17). CD $\Delta\epsilon$ (nm): +1.36 (249), -1.27 (210) (c 4.31×10^{-5} , MeOH). HR-ESI-MS (positive-ion mode) m/z : 467.2017 [M + Na]⁺ (C₂₅H₃₂O₇Na requires 467.2040).

6. Compound **2**: colorless plates (CHCl₃), m.p. 202-203 °C, $[\alpha]_D^{26}$ +78.7 (c 0.13, CHCl₃). IR (KBr) ν_{\max} cm⁻¹: 3479, 2968, 2926, 2855, 1761, 1721, 1634, 1461, 1193, 804. ¹H-NMR (CDCl₃, 400 MHz): δ 5.78 (1H, d, $J = 5$ Hz, H-1), 5.39 (1H, br s, H-18a), 5.23 (1H, s, H-18b), 4.39 (1H, d, $J = 6$ Hz, H-5), 4.17 (1H, br t-like, $J = 8$ Hz, H-11), 3.11 (1H, br d, $J = 13$ Hz, H-13), 2.92 (1H, d, $J = 13$ Hz, H-7), 2.46 (1H, dd, $J = 15, 7$ Hz, H-10a), 2.39 (3H, s, H₃-17), 2.38 (1H, overlapped, H-2'), 2.34 (1H, dd, $J = 14, 7$ Hz, H-3a), 2.26 (1H, d, $J = 6$ Hz, -OH at C-5), 2.22 (1H, m, H-2), 2.07 (1H, dd, $J = 15, 10$ Hz, H-10b), 1.68 (1H, ddq, $J = 14, 7, 7$ Hz, H-3'a), 1.60 (1H, br d, $J = 3$ Hz, -OH at C-11), 1.56 (1H, dd, $J = 14, 10$ Hz, H-3b), 1.43 (1H, ddq, $J = 14, 7, 7$ Hz, H-3'b), 1.27 (3H, s, H₃-20), 1.12 (3H, d, $J = 7$ Hz, H₃-5'), 0.92 (3H, d, $J = 7$ Hz, H₃-19), 0.90 (3H, dd, $J = 7, 7$ Hz, H₃-4'). ¹³C-NMR (CDCl₃, 100 MHz): δ 202.0 (C-9), 175.5 (C-1'), 174.7 (C-15), 146.1 (C-12), 113.8 (C-18), 88.0 (C-6), 75.5 (C-1), 75.4 (C-5), 67.9 (C-11), 66.0 (C-14), 64.8 (C-4), 60.1 (C-8), 48.0 (C-7), 41.2 (C-2'), 35.8 (C-10), 34.4 (C-2), 34.3 (C-3), 32.4 (C-13), 26.6 (C-3'), 26.0 (C-17),

22.0 (C-20), 16.9 (C-5'), 12.6 (C-19), 11.8 (C-4'). HR-ESI-MS (positive-ion mode)

m/z : 471.1937 [M + Na]⁺ (C₂₄H₃₂O₈Na requires 471.1689).

7. X-ray diffraction study on compound **1**: C₂₅H₃₂O₇ · C₃H₈O, $M = 504.60$, crystal size:

$0.50 \times 0.30 \times 0.15 \text{ mm}^3$, space group: orthorhombic, $P2_12_12_1$, $T = 120 \text{ K}$, $a =$

$10.1775(10) \text{ \AA}$, $b = 10.4348(10) \text{ \AA}$, $c = 25.908(3) \text{ \AA}$, $V = 2751.5(5) \text{ \AA}^3$, $Z = 4$, $D_c =$

1.218 Mg/m^3 , $F(000) = 1088$. The data were measured using a Bruker APEX II

CCD diffractometer, using MoK α graphite-monochromated radiation ($\lambda = 0.71073$

\AA) in the range of $3.14 < 2\theta < 53.4$. Of 13566 reflections collected, 3212 were

unique ($R_{\text{int}} = 0.0224$), data/restraints/parameters 3212 / 0 / 334. The structure was

solved by a direct method using the SHELXS-97.¹¹ The refinement and all further

calculations were carried out using SHELXL-97.¹¹ The H atoms were included at

calculated positions and treated as riding atoms using the SHELXL default

parameters. The non-H atoms were refined anisotropically using weighted

full-matrix least-squares on F^2 . Final goodness-of-fit on $F^2 = 1.048$, $R_1 = 0.0344$,

$wR_2 = 0.0868$ based on $I > 2\sigma(I)$ and $R_1 = 0.0378$, $wR_2 = 0.0891$ based on all data.

The largest difference peak and hole were 0.335 and -0.228 e\AA^{-3} , respectively.

8. Fragoso-Serrano, M.; Gibbons, S.; Pereda-Miranda, R. *Planta Med.* **2005**, 71, 278.

9. Compound **1** (2 mg) was dissolved in 100 μL of 50% aqueous 1,4-dioxane and then

100 μL of a 10% KOH solution was added. The reaction mixture was kept at 100 $^{\circ}\text{C}$ for 3 h and then the cooled solution was neutralized by the addition of IR-120B (H^+) ion-exchange resin. An aliquot (20 μL) was analyzed by a HPLC system equipped with an optical rotation detector on an ODS column with a solvent system of 20% CH_3CN in H_2O , containing 0.5% trifluoroacetic acid. A peak appeared at 16.4 min which showed positive chirality and was identified as that of authentic (*S*)-(+)-2-methylbutanoic acid.

10. X-ray diffraction study on compound **2**: $\text{C}_{24}\text{H}_{32}\text{O}_8$, $M = 448.50$, crystal size: $0.30 \times 0.15 \times 0.15 \text{ mm}^3$, space group: monoclinic, $P2_1$, $T = 120 \text{ K}$, $a = 9.9294(12) \text{ \AA}$, $b = 9.1267(11) \text{ \AA}$, $c = 12.5443(15) \text{ \AA}$, $\beta = 98.650(1)^\circ$, $V = 1123.9(2) \text{ \AA}^3$, $Z = 2$, $D_c = 1.325 \text{ Mg/m}^3$, $F(000) = 1088$. Of 5560 reflections collected in the range of $3.28^\circ < 2\theta < 54.1^\circ$, 2416 were unique ($R_{\text{int}} = 0.0154$), data/restraints/parameters 2416 / 1 / 296. The structure was solved in a similar manner to as for compound **1**. Final goodness-of-fit on $F^2 = 1.056$, $R_1 = 0.0315$, $wR_2 = 0.0794$ based on $I > 2\sigma(I)$ and $R_1 = 0.0335$, $wR_2 = 0.0809$ base on all data. The largest difference peak and hole were 0.285 and -0.208 e\AA^{-3} , respectively.

11. Sheldrick, G. M. *Acta. Cryst.* **2008**, A64, 112.

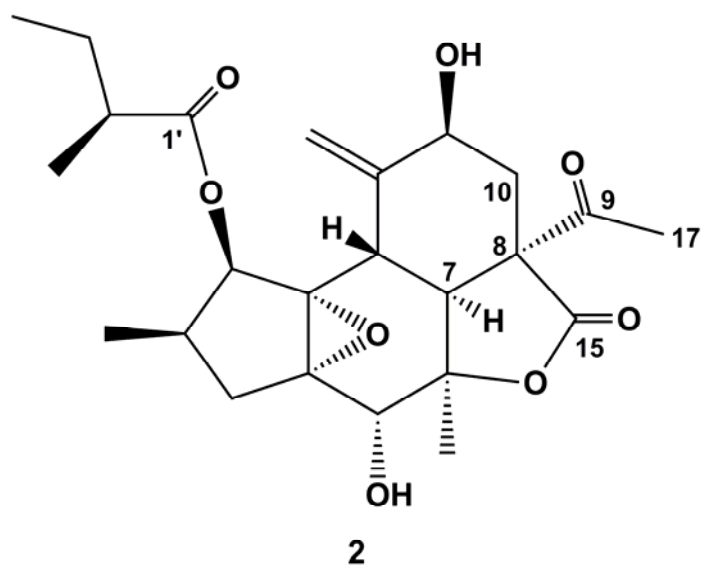
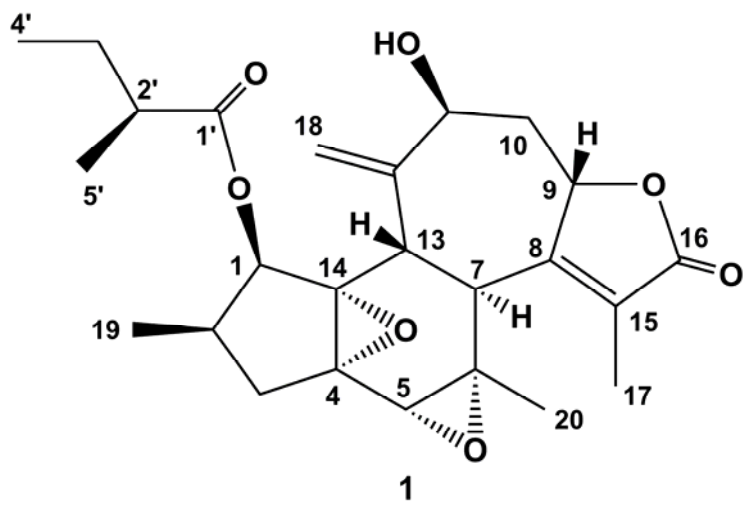


Fig. 1 Structures of Compounds 1 and 2

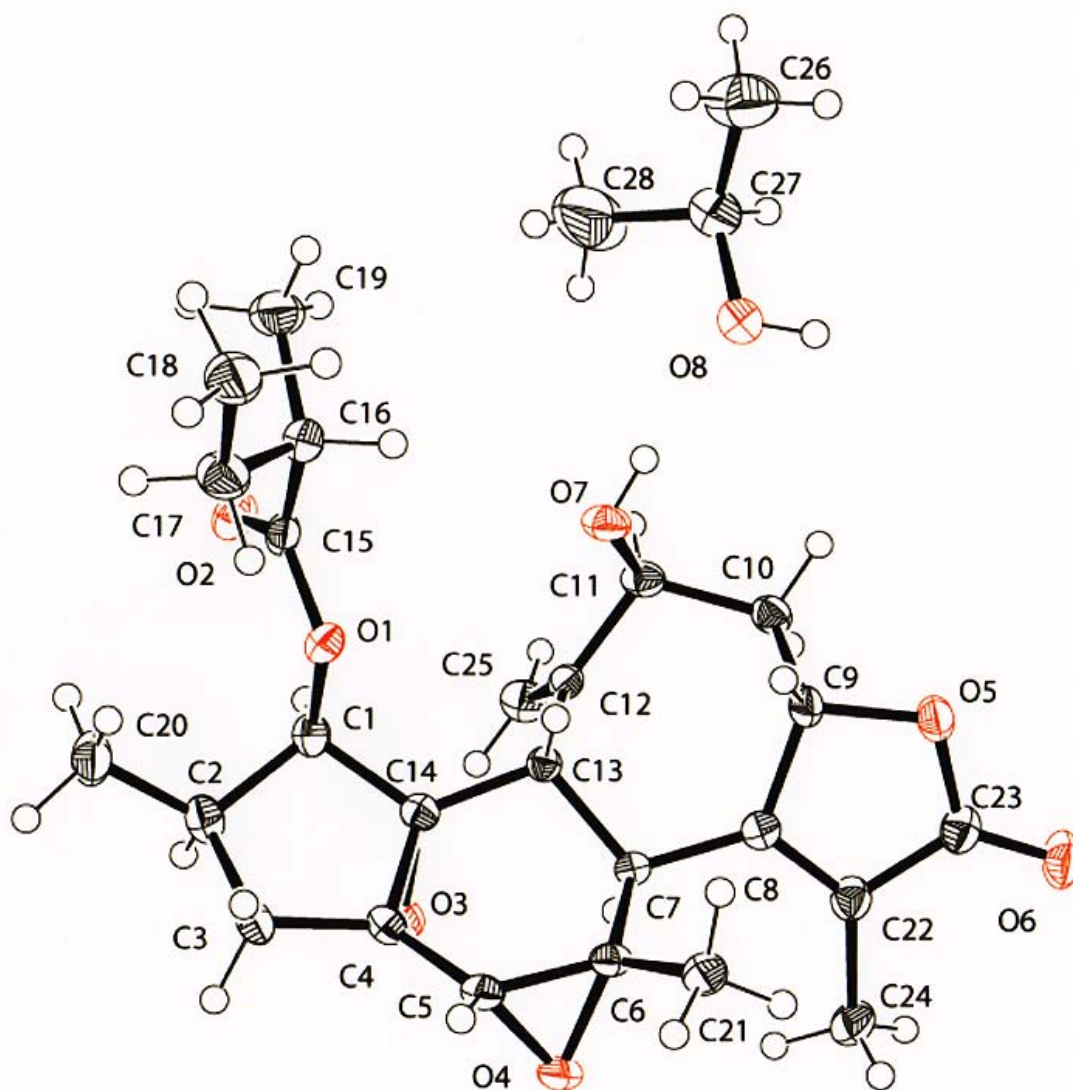


Fig. 2 ORTEP drawing of compound 1

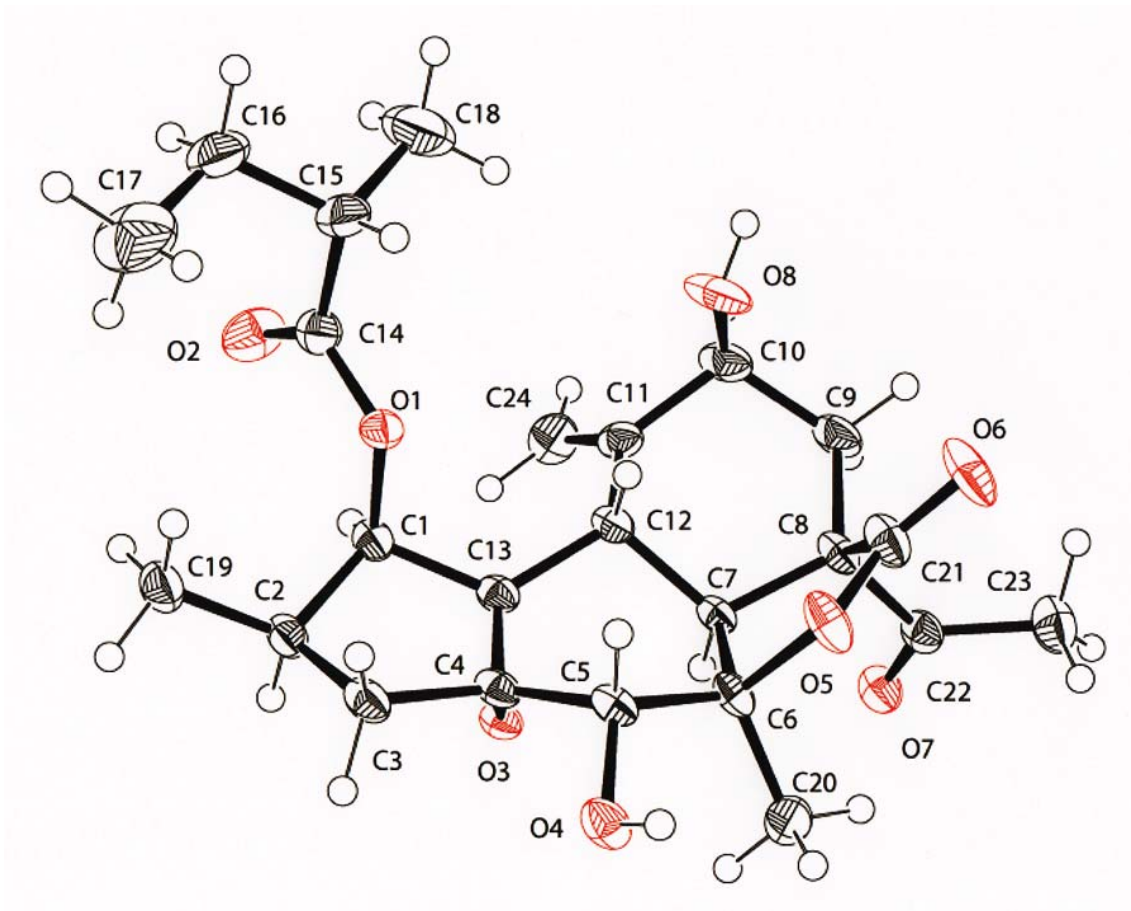


Fig. 3 ORTEP drawing of compound 2