Genomic Characterization of *Ralstonia solanacearum* Phage ϕRSB1, a T7-like Wide-host-range Phage

Takeru Kawasaki, Mio Shimizu, Hideki Satsuma, Akiko Fujiwara, Makoto Fujie, Shoji Usami and Takashi Yamada*

Department of Molecular Biotechnology, Graduate School of Advanced Sciences of Matter, Hiroshima University, Higashi-Hiroshima 739-8530, Japan

Running title: NOTES

*Corresponding author:
Dr. Takashi Yamada
Department of Molecular Biotechnology,
Graduate School of Advanced Sciences of Matter,
Hiroshima University, 1-3-1 Kagamiyama,
Higashi-Hiroshima 739-8530, Japan
Tel/Fax: +81-82-424-7752.
E-mail: tayamad@hiroshima-u.ac.jp

Abstract

ϕRSB1 is a wide-host-range, T7-like bacteriophage that infects and efficiently lyses the phytopathogenic bacterium *Ralstonia solanacearum*. The ϕRSB1 genome comprises 43,079-bp dsDNA (61.7% G+C) with 325-bp terminal repeats and contains 47 open reading frames. Strong activity of tandem early promoters and wide specificity of phage promoters of ϕRSB1 were demonstrated.

Text

The phytopathogenic gram-negative bacterium *Ralstonia solanacearum* causes bacterial wilt disease in many important crops (10). Recently, Yamada et al. (9, 12, 21) isolated and characterized various kinds of bacteriophage that specifically infect *R. solanacearum* strains belonging to different races and/or biovars. These phages may be useful as a tool not only for molecular biological studies of *R. solanacearum* pathogenicity but also for diagnosis and biocontrol of bacterial wilt. In this study, we report the genome and characteristic features of a new phage ϕRSB1. ϕRSB1 was isolated from a soil sample from a tomato crop field, and was selected for its ability to form large clear plaques on plate cultures of *R. solanacearum* strain M4S (for details of bacterial strains see 21). Plaques formed on assay plates (CPG) were 1.0-1.5 cm in diameter. This phage has a wide host range and infected 13 of 15 strains tested, including strains of races 1, 3 and 4 and of biovars 3, 4, and N2. Under laboratory conditions (in standard one-step growth assays) host cells of *R. solanacearum* strains lyses after 2.5-3 h p.i. (with an eclipse period of 1.5-2h), releasing approximately 30-60 pfu new phage particles per cell (burst size) (data not shown). Electron microscopic observation of negatively stained phage particles revealed short-tailed icosahedral structures resembling those of the family *Podoviridae*. The particles consisted of a head approximately 60 nm in diameter and a stubby

tail 20 nm in length (data not shown).

The ϕRSB1 genome is linear double-stranded (ds) DNA of approximately 43.0 kbp in size as determined by pulsed-field gel electrophoresis (data not shown). Since no oligomeric forms were formed after heat-treatment, cohesive ends are absent. The sequence of the ϕRSB1 genome was determined using DNA purified from phage particles by shotgun sequencing and primer walking strategy (9). Sequences were assembled into a circular contig of 42,754 bp, suggesting the presence of long terminal repeats. The precise sequence of the repeat was determined by direct sequencing of genomic DNA with outward-directed primers, located outside the possible terminal repeat region. The final sequence of the ϕRSB1 genome is 43,079 bp, and includes direct terminal repeats of 325 bp. The ϕRSB1 genome size is comparable with that of *Pseudomonas aeruginosa* phage ϕKMV (42,159 bp; accession no. AJ505558) and a little larger than coliphages T7 (39,937 bp; accession no. NC_00164) and T3 (38,208 bp; accession no. NC_003298). The size of the ϕRSB1 terminal repeats is smaller than that of ϕKMV (414 bp) but larger than that of T7 (160 bp) or T3 (231 bp). The G + C content of the genome is 61.7%. This value is lower than the G + C values of the large and small replicons of the *R. solanacearum* GMI1000 genome (67.04% and 66.86%, respectively) (18). Potential ORFs consisting of more than approximately 50 codons and starting with ATG, GTG, or TTG were identified using the Orfinder and DNASIS programs. The presence of a Shine-Dalgarno ribosome-binding sequence preceding the initiation codon was taken into account for ORF prediction. Possible functions were assigned to ORFs by searching through databases using BLAST, BLASTX, and BLASTP programs (1). Accordingly, a total of 47 potential ORFs oriented in the same direction were assigned on the genome (Fig. 1A and Supplementary data). To find homologous sequences, nucleotide sequences from ϕRSB1 DNA were used to searched databases with BLAST and BLASTX programs. Patchy or local

homologies were detected in the genomic sequences of various phages including *Xanthomonas oryzae* phages Xop411 (accession no. DQ777876) and Xp10 (AY299121; 22), *Pseudomonas aeruginosa* phages ϕKMV (accession no. AJ505558; 14) , LKD16 and LKA1 (6), *Erwinia amylovora* phage Era103 (accession no. EF160123), and *Burkholderia cepacia* phage BcepF1 (accession no. AY616033). All of these are members of *Podoviridae*. The genome of coliphage T7, the representative of T7-like viruses of the *Podoviridae,* generally consists of three functional gene clusters; one for early functions (class I), one for DNA metabolism (class II), and the other for structural proteins and virion assembly (class III) (8). These gene clusters are essentially conserved in the ϕRSB1 genome. Figure 1A shows putative ORFs identified on the ϕRSB1 genome compared with ORFs from other phages; *Xanthomonas* phage Xop411 (giving the highest local similarities), *Pseudomonas* phage ϕ KMV(showing marginal similarity but longest regions of similarity), and coliphage T7. The mosaic genetic relationship of ϕRSB1 indicates frequent recombinations on the ϕRSB1 ancestral genome during its evolution, in the way suggested for tailed phages and their prophages (2-5, 11).

One of the characteristic features found in the ϕRSB1 gene organization is that the predicted gene for RNA polymerase (RNAP) of ϕRSB1 (*orf26*) is not located in the early gene region (class I) but at the end of the class II region (Fig. 1A). Another exception is the gene for DNA ligase (DNAL); *orf25* encoding the ϕRSB1 DNA ligase is in front of the RNAP ORF (*orf26*), whereas the gene encoding T7 DNAL is downstream of the gene for RNAP at the end of the class I cluster (8). In *Pseudomonas* phages ϕKMV, LKD16, and LKA1, the DNA ligase gene is upstream of the gene for DNAP in the class II gene cluster (Fig. 1A).

Similarly to the T7 genome, structural proteins are predicted to be encoded in the class III gene cluster of the ϕRSB1 genome. Purified ϕRSB1 particles gave at least 10 protein bands

on SDS-PAGE (Fig. 2). Each band was extracted from the gel and was subjected to N-terminal amino acid sequencing (19). The N-terminal sequence of each protein always started from the second amino acid residue of its corresponding ORF, except for ORF35, which included the first methionine (Fig. 2). In addition to known structural proteins, ORF35, ORF36 and ORF46 were identified as structural proteins. In this way, all predicted protein in the class III-structural region were identified, except for the scaffolding protein (ORF31) and a possible tail fiber protein ORF38, which may be lost during purification of the phage particles. In the case of the largest structural protein (170-180 kDa), determination of the N-terminal sequence was unsuccessful using standard methods, possibly because of modification at the N-terminus. However, it most likely corresponds to ORF37, judged from its exceptionally large size (174 kDa); there is no other candidate for this size. ORF37 may encode a tail protein with a transglycosylase domain.

T7-like phages are generally known as absolute lytic phages, with a few exceptions, such as integrase-coding phages, e.g. prophage 3 of *Pseudomonas putida* (17), and the cyanophage P-SSP7 (16). Sometimes nucleotide sequences related to T7-like phages are found in conjunction with other temperate phages such as λ–like phages that are integrated in various bacterial genomes (2-5, 11). BLAST and BLASTX database searches using the ϕRSB1 sequence revealed a significantly homologous region (at the nucleotide sequence level) in the genome of *Burkholderia pseudomallei* 1710b (accession no. CP000124). A matrix comparison plot showed that this homology is extended to a 20 kbp region (1710b positions 1,740,980-1,761,000) (data not shown). In the 1710b genome, this region is embedded in a large (85 kbp) prophage sequence (1710b positions 1,719,000-1,804,000), which is related to λ–like phages such as *B. pseudomallei* phage ϕ1026b (7) and *B. thailandensis* phage ϕE125 (20). The homologous region of 1710b-prophage contains 8 ORFs encoding a DNA primase,

DNA helicase, DNA ligase, DNA polymerase, exonuclease, and RNA polymerase etc. These correspond to the class II genes of ϕRSB1 as shown in Fig. 1B. Interestingly, the putative ϕRSB1 promoters (see below) were also found in this 1710B region (positions 1743597-1743650 and 1744983-1745040; Fig. 1B). Both *Ralstonia* and *Burkholderia* belong to *Betaproteobacteria* and may share common bacteriophages (9). Database search also showed that a 10 kbp genomic region of the *R. solanacearum* GMI1000 (positions 1,661,000-1,672,000) contains a 1589-aa ORF (RSO5240) showing significant similarity to ϕRSB1 ORF37, which encodes a putative transglycosylase-tail protein (Supplementary data, E-value e-131). Amino acid sequence similarity extends to the entire region consisting of the N-terminal transglycosylase and C-terminal core or tail domains (1606 aa in ϕRSB1 ORF37). This GMI1000 ORF is associated with two IS transposase sequences (ISRSO8 transposase A and B; RSO5237 and RSO5236, respectively) on the left side. Immediately to the right of this ORF, there is an ORF (RSO5241) for a putative integrase, which is closely associated with arginine tRNA (AGA), a possible *att* sequence (Fig. 1C). This structure indicates horizontal acquisition of this ϕRSB1 ORF by host cells, as well as some involvement of the phage integrase/*att* sequence and transposons in such an event. In the context of lysogenic conversion or introduction of a new fitness factor by phage in the pathogenic bacteria, the functions of ϕRSB1 ORF37 are interesting.

As shown in Fig. 3, several putative transcription promoter and terminators were identified in apparently noncoding regions (more than 100 bp long) in the ϕRSB1 genome. A typical prokaryotic promoter sequence (resembling *E. coli* σ^{70}) was repeated five times (p1-p5) in a left 1,000-bp region without ORFs (Fig. 1A and Fig. 3A). In addition, a few other putative sequences of host σ^{70} promoter (p6-p8) were detected in front of ORFs 1, 17, and 39 (Fig. 1A and Fig. 3A). There are possible ρ–independent terminator-like sequences (Fig. 3C).

A terminator-like sequence (t2) present after ORF13 is located in the region that separates class I and class II genes (Fig. 1A). Another possible terminator (t3) is located immediately downstream of ORF32, encoding the major capsid protein. t4 is located in front of a putative promoter p8 for ORF40 and ORF41 (similar to the large subunit of a terminase). A final terminator (t5) was defined behind the last ORF47. The terminator positions of t2, t3, and t5 are consistent with those reported in *Pseudomonas* phages ϕKMV (14), LKD16 and LKA1 (6) (Fig. 1A). Searching for core promoter-like sequences conserved in phages T3, T7, or SP6 in the ϕRSB1 intergenic regions could not find any significant ones. Instead, three sets of common sequence elements were found in front of ORFs 16, 18, and 32 (designated ϕp1, ϕp2, and ϕp3, Fig. 1A). We found a set of sequence elements consisting of a GC-rich stretch, and TTGT, TCTGG, and CGGGCAC motifs preceding an AG-rich Shine-Dalgarno sequence (Fig. 3B). Activity of transcriptional promoters of both host- and phage-types thus predicted on the ϕRSB1 genome was examined using a GFP-expressing single-copy plasmid pRSS12 (13), where the *lac* promoter for *gfp* expression was replaced with a ϕRSB1 promoter sequence. When we tested bacterial σ^{70}–type promoters p1-p4, p1-p5, and p1-p6, which are located tandemly at the beginning of the class I gene cluster, transformed cells of *R. solanacearum* strains always showed strong GFP fluorescence. Fluorescence was 3-15 times greater than that of pRSS12 with a *lac* promoter. Results with strain MAFF301558 as the host are shown in Table 1. Increased GFP intensity from p1-p4 via p1-p5 to p1-p6 clearly demonstrates actual promoter activities of these ϕRSB1 early promoters. ϕp1 is located at the beginning of the class II gene cluster, after the possible terminator t2 and in front of ORF16 encoding possible DNA primase. ϕp3 is located upstream of ORF32, which encodes the major capsid protein. Both ϕp1 and ϕp3 also function as promoters for bacterial RNA polymerase in ϕRSB1-uninfected *R. solanacearum* cells, but show lower activity compared with p1-p6

(Table 1). The promoter activity was also examined in *R. solanacearum* cells after infection with ϕRSB1. As strain MAFF301558 was found to be a low-efficient host, giving fewer titers of phage progeny, the host was changed to strain M4S. After infection with ϕRSB1, GFP fluorescence intensity retained at almost the same levels in cells containing the promoters, p1-p5, or p1-p6, whereas cells with ϕp1 or ϕp3 showed increased GFP fluorescence after 30 min p.i. to 90 min p.i. (Table 1). At 120 min p.i. cell lysis began. These results indicate that ϕp1 and ϕp3 are functional in transcription by both bacterial and phage RNAPs. Bacterial σ^{70}–type promoters are not shut down but continue to function after infection.

The occurrence of host σ^{70}-type promoter sequences in the late gene clusters, class II and class III, and the low specificity of phage promoters further imply that expression of ϕRSB1 genes is highly dependent on the host RNAP. To determine whether host RNAP is involved in late stages of ϕRSB1 infection, rifampicin was added to ϕRSB1-infected cultures at various times post infection and the number of progeny phage was determined. The results are shown in Fig. 4. In samples that were incubated with rifampicin, more than 90% of phage progeny was obtained when the drug was added at 90 min p.i. or later, and no or very few progeny phages were obtained when the drug was added 75 min p.i. or earlier. These results indicate that a switch from host RNAP to ϕRSB1 RNAP occurs between 75 min p.i. and 90 min p.i. and that late stages of ϕRSB1 replication are independent of rifampicin. The late genes can be transcribed by rifampicin-resistant ϕRSB1 RNAP, at least in the presence of rifampicin.

This study was supported by the Industrial Technology Research Grant Program (04A09505) from the New Energy and Industrial Technology Development Organization (NEDO) of Japan. The sequence data for ϕRSB1 genomic DNA has been deposited in the DDBJ database under accession no. AB451219.

REFERENCES

1. **Altschul, S. F., T. L., Madden, A. A., Schaffer, Z., Zhang, W., Miller, and D. J. Lipman.** 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Res. **25:** 3389-3402.
2. **Brussow, H., C. Canchaya, and W.-D. Hardt.** 2004. Phages and the evolution of bacterial pathogens: from genomic rearrangements to lysogenic conversion. Microbiol. Mol. Biol. Rev. **68:** 560-602.
3. **Canchaya,C., C. Proux, G. Fournous, A. Bruttin and H. Brussow.** 2003. Prophage genomics. Microbiol. Mol. Biol. Rev. **67:** 238-276.
4. **Casjens, S.** 2003. Prophages and bacterial genomics: what have we learned so far? Mol. Microbiol. **49:** 277-300.
5. **Casjens, S.** 2005. Comparative genomics and evolution of the tailed-bacteriophages. Curr. Opin. Microbiol. **8:**451-458.
6. **Ceyssens, P.-J., R. Lavigne, W. Mattheus, A. Chibeu, K. Hertveldt, J. Mast, J. Robben, and G. Volckaert.** 2006. Genomic analysis of *Pseudomonas aeruginosa* phages LKD16 and LKA1: establishment of the ϕKMV subgroup within the T7 subgroup. J. Bacteriol. **188:**6924-6931.
7. **DeShazer, D.** 2004. Genomic diversity of *Burkholderia pseudomallei* clinical isolates: subtractive hybridization reveals a *Burkholderia mallei*-specific prophage in *B. pseudomallei* 1026b. J, Bacteriol. **186:** 3938-3950.
8. **Dunn, J. J., and F. W. Studier.** 1983. Complete nucleotide sequence of bacteriophage T7 DNA and the locations of T7 genetic elements. J. Mol. Biol. **166:**477-535.
9. **Fujiwara, A., T. Kawasaki, S. Usami, M. Fujie, and T. Yamada.** 2008. Genomic

characterization of Ralstonia *solanacearum* phage ϕRSA1 and its related prophage (ϕRSX) in strain GMI1000. J. Bacteriol. **190:**143-156.

10. **Hayward, A. C.** 1991. Biology and epidemiology of bacterial wilt caused by *Pseudomonas solanacearum*. Annu. Rev. Phytopathol. **29:** 65-87.
11. **Hendrix, R. W., G. F. Hatfull, and M. C. M. Smith.** 2003. Bacteriophages with tails:chasing their origins and evolution. Res. Microbiol. **154:** 253-257.
12. **Kawasaki, T., S. Nagata, A. Fujiwara, H. Satsuma, M. Fujie, S. Usami, and T. Yamada.** 2007. Genomic characterization of the filamentous integrative bacteriophage ϕRSS1 and ϕRSM1, which infect *Ralstonia solanacearum*. J. Bacteriol. **189:**5792-5802.
13. **Kawasaki, T., H. Satsuma, M. Fujie, S. Usami, and T. Yamada.** 2007. Monitoring of phytopathogenic *Ralstonia solanacearum* cells using green fluorescent protein-expressing plasmid derived from bacteriophage ϕRSS1. J. Biosci. Bioeng. **104:**451-456.
14. **Lavigne, R., M. V. Burkal'tseva, J. Robben, N. N. Sykilinda, L. P. Kurochkina, B. Grymonprez, B. Jonckx, V. N. Krylov, V. V. Mesyanzhinov, and G. Volckaert.** 2003. The genome of bacteriophage ϕKMV, a T7-like virus infecting *Pseudomonas aeruginosa*. 2003. Virology 312:49-59.
15. **Liao, Y. D., J. Tu, and T. T. Kuo.** 1987. Regulation of transcription of the Xp10 genome in bacteriophage-infected *Xanthomonas campestris* pv. *oryzae*. J. Virol. **61:**1695-1699.
16. **Lindell, D., M. B. Sullivan, Z. I. Johnson, A. C. Tolonen, F. Rohwer, and S. W. Chisholm.** 2004. Transfer of photosynthesis genes to and from *Prochlorococcus* viruses. Proc. Natl. Acad. Sci. USA **101:** 11013-11018.
17. **Molineux I. J.** 1999. T7-like phages (*Podoviridae*), p.1722-1729. *In* A. Granoff and R. Webster (ed), Encyclopedia of virology. Academic Press, Ltd., London, UK.
18. **Salanoubat, M., S. Genin, F. Artiguenave, J. Gouzy, S. Mangenot, M. Ariat, A.**

Billault, P. Brottier, J. C. Camus, L. Cattolico, M. Chandler, N. Choisene, S. Claudel-Renard, N. Cunnac, C. Gaspin, M. Lavie, A. Molsan, C. Robert, W. Saurin, T. Schlex, P. Siguier, P. Thebault, M. Whalen, P. Wincker, M. Levy, J. Weissenbach, and C. A. Boucher. 2002. Genome sequence of the plant pathogen *Ralstonia solanacearum*. Nature **415:** 497-502.

19. **Songsri, P., S. Hiramatsu, M. Fujie, and T. Yamada.** 1997. Proteolytic processing of *Chlorella* virus CVK2 capsid proteins. Virology **227:** 252-254.

20. **Woods, D. E., J. A. Jeddeloh, D. L. Fritz, and D. DeShazer.** 2002. *Burkholderia thailandensis* E125 harbors a temperate bacteriophage specific for *Burkholderia mallei*. J. Bacteriol. **184:**4003-4017.

21. **Yamada, T., T. Kawasaki, S. Nagata, A. Fujiwara, S. Usami, and M. Fujie.** 2007. Isolation and characterization of bacteriophages that infect the phytopathogen *Ralstonia solanacearum*. Microbiology **153:** 2630-2639.

22. **Yuzenkova, J., S. Nechaev, J. Berlin, D. Rogulja, K. Kuznedelov, R. Inman, A. Mushegian, and K. Severinov.** 2003. Genome of *Xanthomonas oryzae* bacteriophage XP10: an odd T-odd phage. J. Mol. Biol. **330:**735-748.

Figure legends

FIG. 1. Genetic organization and comparative analyses of ϕRSB1 genome. (A) Comparison of the genomes of ϕRSB1 and T7-like phages. In each alignment, corresponding ORFs (horizontal arrows) are connected by shading. Three functional gene clusters, class I (green), class II (Yellow), and class III (orange) are indicated above the ϕRSB1 and the T7 maps and

corresponding ORFs are colored. Putative bacterial promoters, phage promoters, and terminators of transcription are indicated under the ϕRSB1 map by p, ϕp, and t, respectively. Promoters and terminators are also shown in the ϕKMV and T7 maps. Xop411, *Xanthomonas oryzae* phage (44,520 bp, accession no. DQ777876); ϕKMV, *Pseudomonas aeruginosa* phage (42,159 bp, AJ50558); coliphage T7 (39,937 bp, NC_00164). DNAP, DNA polymerase; DNAL, DNA ligase; RNAP, RNA polymerase; MCP, major capsid protein; LYS, lysozyme. (B) Class II region of ϕRSB1 genome (ORF16-ORF26) often shows high homology with phage or prophage sequences of different phage groups. The ϕRSB1 region is aligned with the corresponding prophage sequence of *Burkholderia pseudomallei* 1710b (accession no. CP000124). (C) Region on the GMI1000 genome (positions 1,661,000-1,672,000) containing a large ϕRSB1-ORF37-like ORF (RSO5240), which encodes a putative transglycosylase protein. This GMI1000 region is flanked by two ORFs encoding ISRSO8 transposase A and B on the left side and by integrase (RSO5241) and argnine tRNA (AGA) on the right side.

FIG. 2. Identification of ϕRSB1 virion proteins. Proteins from purified ϕRSB1 particles were separated by SDS-PAGE (10% gel) and stained with Coomassie blue. Molecular size of each marker protein (Amersham full-range MW markers and a LMW gel filtration calibration kit) is indicated on the left. N-terminal amino acid sequence (5 residues) determined for each ϕRSB1 protein band is shown on the right with its corresponding ORF. Amino acids in parenthesis are obscure residues. Although the N-terminal sequence could not determined, the largest protein, approximately 175-180 kDa, is predicted to be ORF37, as there is no other candidate for this large size. A few small proteins were lost from the gel during electrophoresis.

FIG. 3. Predicted regulatory sequences found in the ϕRSB1 genome. (A) *E. coli* σ^{70}-promoter -like sequences, (B) putative promoter sequences for ϕRSB1-encoded RNAP, and (C) putative terminators.

FIG. 4. Late stages of ϕRSB1 development are resistant to rifampicin. Cells of *R. solanacearum* M4S were infected with ϕRSB1 at moi 5. At the indicated times p.i., aliquots of the infected culture were withdrawn, and incubated with 100 µg/ml rifampicin for 2.5 h before $CHCl_3$-treatment and determination of phage titers (pfu) (open circle). For control, phage titers were also determined at indicated times after $CHCl_3$ addition without rifampicin-treatment (closed circle). The method is according to Liao et al. (15).

TABLE 1. Expression of *gfp* by ϕRSB1 promoters

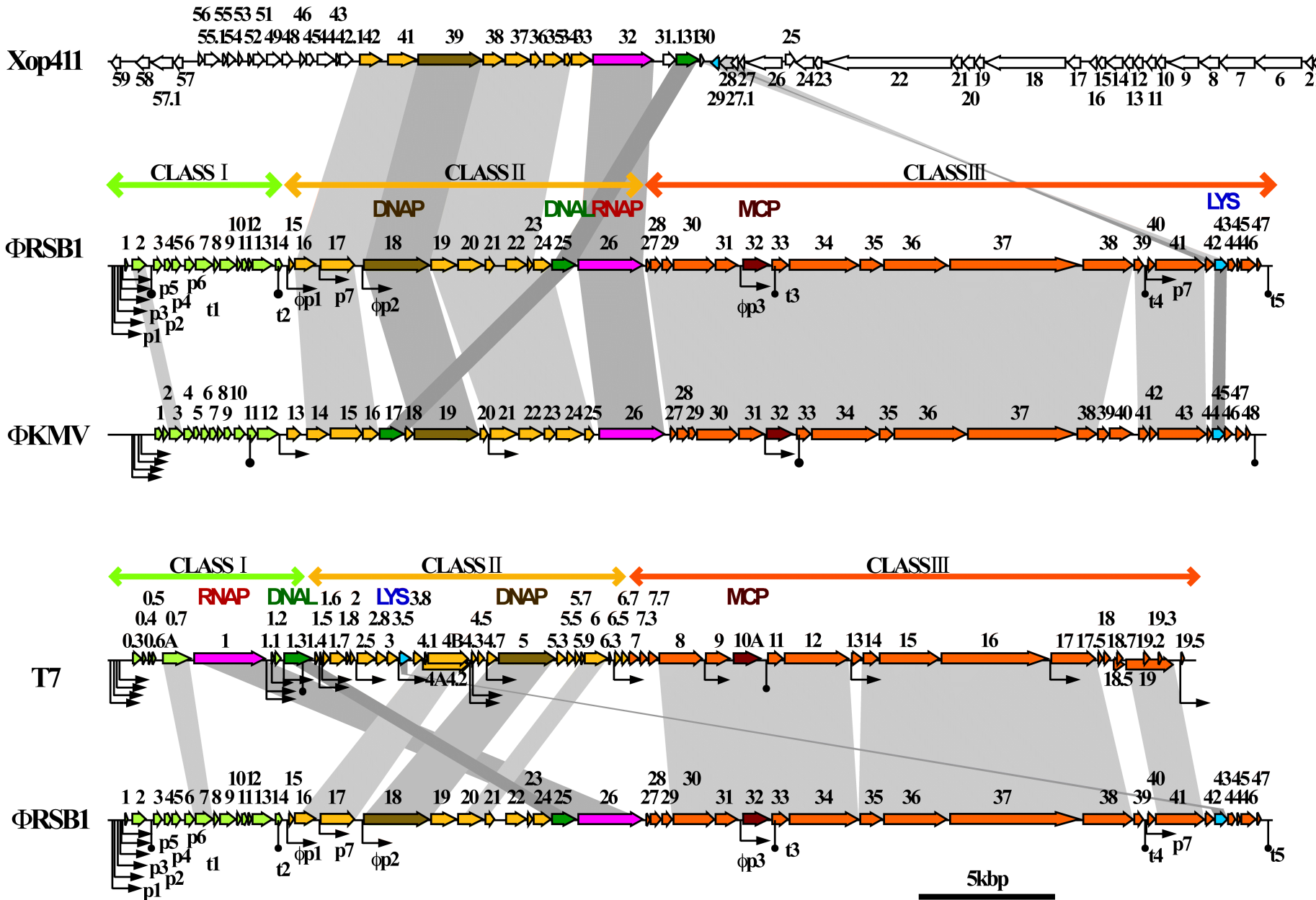
Promoter (positions)	Relative intensity of GFP fluorescence[a]					
	Srain MAFF301558	Strain M4S				
		Time (min) post infection of ϕRSB1				
		0	30	60	90	120
lac	3.5 ± 0.2 (1.0)[b]	1.8 ± 0.1	ND[c]	2.0 ± 0.1	ND	2.0 ± 0.1
p1-p4 (425-845)	10.0 ± 0.3 (3.0)[b]	ND	ND	ND	ND	ND
p1-p5 (425-921)	14.3 ± 0.5 (4.1)[b]	4.2 ± 0.2	4.4 ± 0.2	6.0 ± 0.3	7.2 ± 0.3	9.0 ± 0.4
p1-p6 (425-995)	53.0 ± 0.9 (15.2)[b]	7.3 ± 0.3	8.4 ± 0.3	12.0 ± 0.4	11.7 ± 0.4	13.6 ± 0.5
ϕp1 (6895-7009)	11.0 ± 0.5 (3.2)[b]	6.7 ± 0.3	16.5 ± 0.5	19.0 ± 0.6	32.8 ± 0.8	15.1 ± 0.5
ϕp3 (23515-23675)	7.2 ± 0.3 (2.1)[b]	1.4 ± 0.1	4.1 ± 0.2	5.9 ± 0.2	11.4 ± 0.4	7.8 ± 0.3

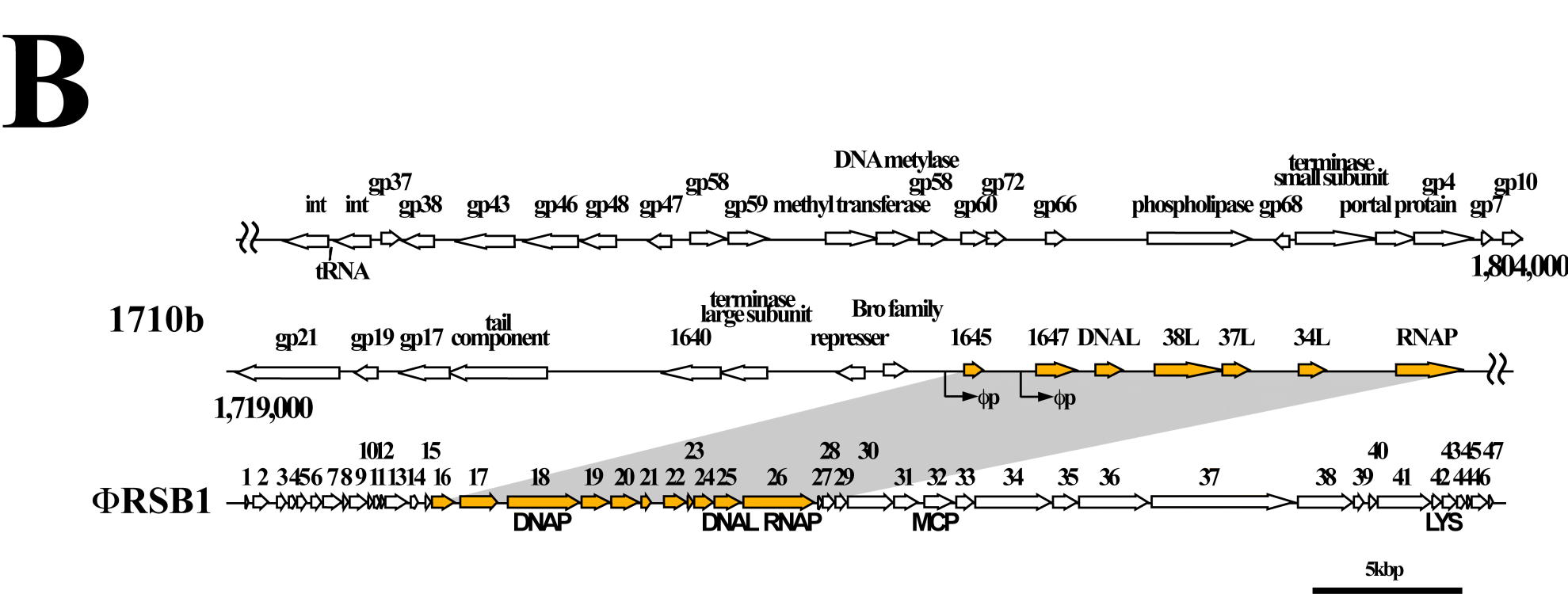
[a] The values are means ± standard errors for data from three independent experiments.

[b] The ratio to *lac*-value (1.0) is in parenthesis.

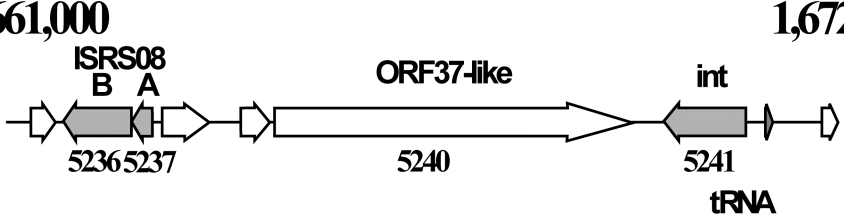
[c] ND: Not determined.

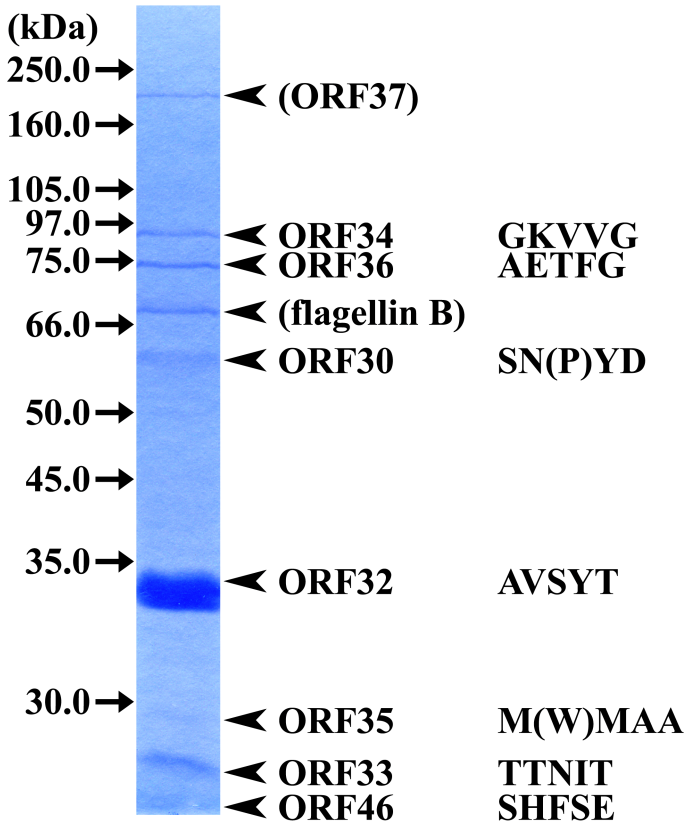
A


Xop411
ΦRSB1
ΦKMV
T7
CLASS I
CLASS II
CLASS III
DNAP
DNAL
RNAP
MCP
LYS
5kbp


B
DNA metylase
terminase
small subunit
gp37
gp58
gp58
gp72
gp4
gp10
int
int
gp38
gp43
gp46
gp48
gp47
gp59
methyl transferase
gp60
gp66
phospholipase
gp68
portal protain
gp7
tRNA
1,804,000
1710b
terminase
large subunit
Bro family
tail
gp21
gp19
gp17
component
1640
represser
1645
1647
DNAL
38L
37L
34L
RNAP
1,719,000
ϕp
ϕp
ΦRSB1
DNAP
DNAL
RNAP
MCP
LYS
5kbp

C


1,661,000
1,672,000
ISRS08
B
A
ORF37-like
int
5236
5237
5240
5241
tRNA


(kDa)
250.0
160.0
105.0
97.0
75.0
66.0
50.0
45.0
35.0
30.0
(ORF37)
ORF34
ORF36
(flagellin B)
ORF30
ORF32
ORF35
ORF33
ORF46
GKVVG
AETFG
SN(P)YD
AVSYT
M(W)MAA
TTNIT
SHFSE

A

E. coli σ^{70}-like promoters

	Positions	
p1	508-552	CAGAGC**TTGACA**GACACACACAGCACAGC---**TACAGT**GGAGTCACAC
p2	589-633	CAGAGC**TTGACA**GAGCGAACGGACTAGCA---**TACAGT**GGAGTCCAGC
p3	709-753	GCACGC**TTGACA**GACGGTACGGACTGAGC---**TACAGT**GAAGCCTAGC
p4	801-845	GTGCGC**TTGACA**AGGCGGACGGCAGGTAG---**TACACT**GCAGTCCAGT
p5	877-925	GGATGC**TTGACA**GAGTATCGTCGGGTTGAACG**TGAAGC**GGCGAACGTGC
p6	934-982	CGCGCT**TTGACC**TACAGGGTACCACTAGGGAC**TGTAGC**GAGTGCAGGCA
p7	8113-8156	GACAAC**TCGACA**ACATGAGCTTCGGCTT----**TGTCAT**TCCGATTTCG
p8	38455-38498	TCCGTC**CTGATC**GCAGTATACACCGTCG----**TACAAT**TTTACTTCCT
	E. coli σ^{70}	------**TTGACA**(-**35**)---------------**-TATAAT**(-**10**)------

B

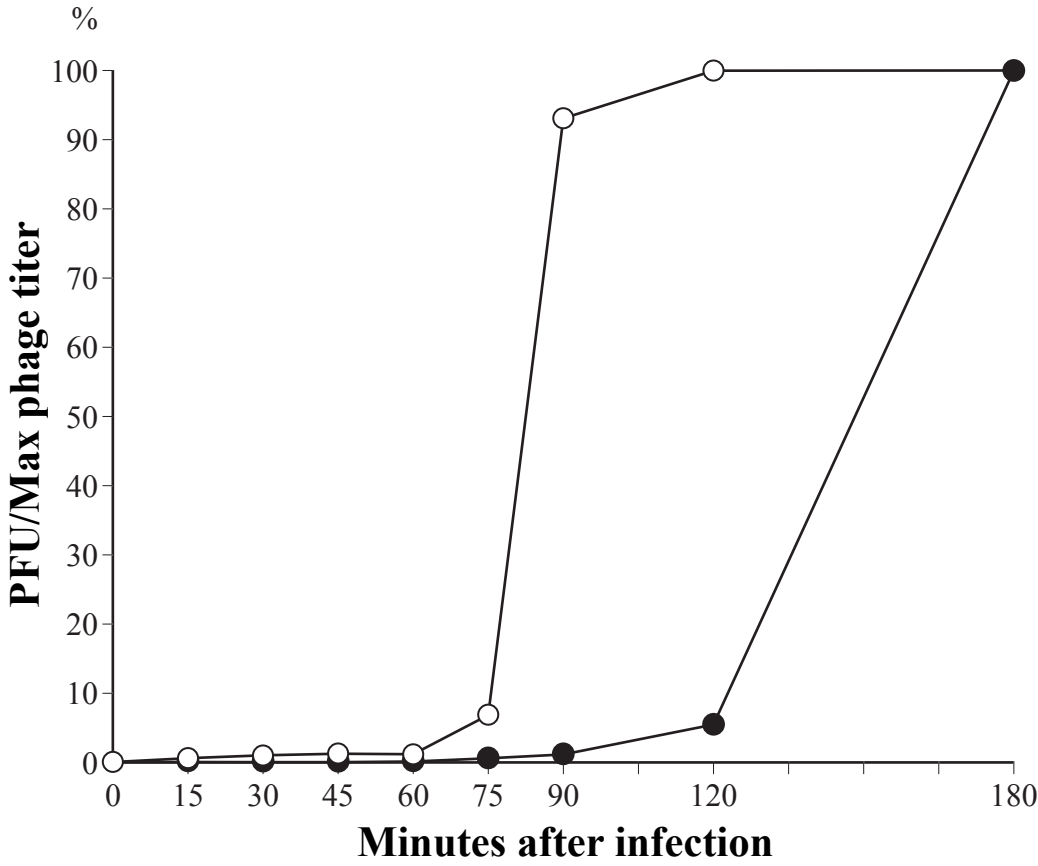
Putative phage promoter sequences

	Positions	
ϕp1	6926-6960	CGA**GCCGGGC**ATCATCC**TTGC**CCTGCT**TCTGG**AGATTCTAGGAG**CGGGCAC**CTT---30b---ATG
ϕp2	9688-9721	AGCT**GGGGGCGGGGC**TA**TTGT**GCATA-**TCTGG**CGG---------**CGGGTAT**GCT---29b---ATG
ϕp3	23561-23593	AGGCT**CGGGCGGGCC**TT**TTGT**CGTT--**TCTGG**CTCCCCT-----**TGGGACAC**AAA--68b---ATG
	Consensus	----**GCstretch**----**TTGT------TCTGG**------------**CGGGCAC**------------ATG

C

Putative phage terminators

	Positions	
t1	1808-1853	GACGTACATGCCCATTCTGCGGAGTGGGCATGTGCATT
t2	6465-6501	ACTCGCGGCTCGGATAACGAGCTTCTGAGCCGCGCAC
t3	24690-24727	CCCAACTGCCCGTCACACTCACAAGGTGGCGGGCATTTTT
t4	38428-38460	TGGGGATACCCGATTGCTGATTGGGTATCCGTC
t5	42791-42864	GTGTGCGCGCGTGCATGTGCGCGTACGTGTGTGCGCATGAGCGCGTGCATGTGCGTGCCTGCGCGCGAGTAGCA


%
100
90
80
70
60
50
40
30
20
10
0
PFU/Max phage titer
0
15
30
45
60
75
90
120
180
Minutes after infection